

ORIGINAL ARTICLE

Spozazomicins A–C, novel antitrypanosomal alkaloids produced by an endophytic actinomycete, *Streptosporangium oxazolinicum* K07-0460^T

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Three novel antitrypanosomal alkaloids, named spozazomicins A–C, were isolated by silica gel column chromatography and HPLC from the culture broth of a new endophytic actinomycete species, *Streptosporangium oxazolinicum* K07-0460^T. The structures of the spozazomicins were elucidated by NMR and X-ray crystal analyses and shown to be new types of pyochelin family antibiotic. Spozazomicin A showed potent and selective antitrypanosomal activity with an IC₅₀ value of 0.11 μg ml⁻¹ *in vitro* without cytotoxicity against MRC-5 cells (IC₅₀=27.8 μg ml⁻¹).

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Keywords: antitrypanosome; endophytic actinomycete; spozazomicins; *Streptosporangium oxazolinicum*

INTRODUCTION

It has become more and more difficult to find new bioactive compounds derived from actinomycete strains isolated from soils, so the focus of attention has switched to find novel actinomycetes from other sources. We have studied endophytic actinomycetes as the producers of new bioactive compounds. On the other hand, we have identified several useful microbial metabolites, such as staurosporine, pyrindicin and dityromycin by a chemical screening approach to detect alkaloids using Dragendorff's reagent.^{1–3} In the course of chemical screening for alkaloids from the culture broths of endophytic actinomycetes, we have now discovered three new alkaloid antibiotics, spozazomicins A–C (1–3) (Figure 1), produced by a new actinomycete species, *Streptosporangium oxazolinicum* K07-0460^T isolated from an orchid collected in Okinawa prefecture, Japan. Spozazomicins A (1) and B (2) exhibit antitrypanosomal activities *in vitro*. We report here the fermentation, isolation, physico-chemical properties and structural elucidation of 1–3. We also report the antitrypanosomal profiles of 1–3 (Figure 1) and discuss structure-activity relationships. The taxonomic study of the spozazomicin producing strain will be reported in the accompanying paper.⁴

RESULTS

Isolation

The procedure for isolation of 1–3, using the Dragendorff reaction, is summarized in Scheme 1. The 4-day-old culture broth (8.4 l) was

added with 8.4 l of ethanol, followed by centrifugation at 3000 r.p.m. The supernatant was concentrated under reduced pressure to remove ethanol and then adjusted to pH 9 with aqueous ammonia followed by extraction with 4.0 l of *n*-butyl acetate. The organic layer was partitioned with 0.1 M hydrochloric acid (2.0 l). The water layer was subsequently adjusted to pH 9 with aqueous ammonia and extracted with 2.0 l of ethyl acetate. The ethyl acetate extract (211.0 mg) was chromatographed on silica gel (20ϕ×70 mm, 0.063–0.200 mm, Merck, Darmstadt, Germany) eluting with a solvent mixture of chloroform and methanol (100:0 v/v (140 ml) and 50:1, v/v (70 ml)). Each fraction was concentrated *in vacuo* to dryness to yield 40.6 and 78.3 mg of crude materials, respectively. The material of fraction 100:0 was purified by HPLC on a Pegasil ODS column (20ϕ×250 mm, Senshu Scientific, Tokyo, Japan) with 10% acetonitrile/0.1% TFA at 8 ml min⁻¹ detected at UV 210 nm. The fraction eluted at 18.5 min was concentrated *in vacuo* to dryness to afford 3 (36.1 mg) as white powder. The material of fraction 50:1 (78.3 mg) was purified by HPLC on a Pegasil ODS column (20ϕ×250 mm) with 45% acetonitrile at 8 ml min⁻¹ detected at UV 210 nm. The fractions eluted at 20.1 and 23.3 min were concentrated *in vacuo* to dryness to afford 2 (5.3 mg) and 1 (24.4 mg), both as white powders.

Physico-chemical properties

The physico-chemical properties of 1–3 are summarized in Table 1. They were readily soluble in methanol and chloroform. They showed

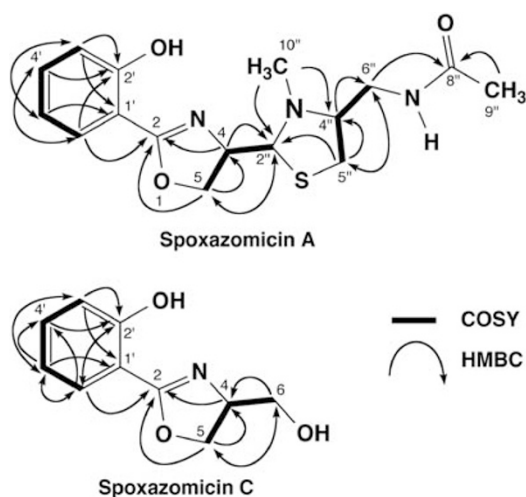
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Table 2 ^1H and ^{13}C NMR spectral data of spoxazomicins A–C (1–3)

Spoxazomicin A (1)				Spoxazomicin B (2)		Spoxazomicin C (3)	
Position	^{13}C	^1H (J in Hz)	^{13}C	^1H (J in Hz)	^{13}C	^1H (J in Hz)	
1'	111.7	—	111.7	—	111.9	—	
2'	161.1	—	161.0	—	161.0	—	
3'	117.6	6.94 (1H, br. d, 8.2)	117.5	6.94 (1H, dd, 0.9, 8.4)	117.5	6.93 (1H, dd, 0.9, 8.6)	
4'	134.6	7.38 (1H, ddd, 0.9, 7.3, 8.2)	134.6	7.38 (1H, ddd, 1.5, 7.2, 8.4)	134.4	7.37 (1H, ddd, 1.5, 7.4, 8.6)	
5'	119.8	6.88 (1H, br. dd, 7.3, 7.7)	119.8	6.87 (1H, ddd, 0.9, 7.2, 7.8)	119.7	6.87 (1H, ddd, 0.9, 7.4, 7.9)	
6'	129.2	7.64 (1H, dd, 0.9, 7.7)	129.2	7.64 (1H, dd, 1.5, 7.8)	129.2	7.63 (1H, dd, 1.5, 7.9)	
2	167.7	—	167.7	—	167.4	—	
4	72.8	4.57 (1H, m)	69.7	4.72 (1H, m)	68.5	4.44 (1H, m)	
5	70.8	4.54 (1H, dd, 9.3, 13.9)	70.5	4.49 (1H, dd, 8.6, 9.3)	70.1	4.50 (1H, m)	
		4.41 (1H, dd, 7.2, 13.9)		4.29 (1H, dd, 8.3, 8.6)		4.33 (1H, m)	
6	—	—	—	—	64.6	3.70 (1H, d, 4.5)	
2''	77.9	4.18 (1H, d, 4.2)	78.3	4.47 (1H, d, 6.6)			
4''	72.7	3.11 (1H, m)	69.1	3.57 (1H, m)			
5''	34.7	2.98 (1H, dd, 5.7, 11.1)	33.2	3.04 (1H, dd, 5.7, 10.2)			
		2.85 (1H, dd, 8.1, 11.1)		2.75 (1H, dd, 5.3, 10.2)			
6''	42.6	3.47 (1H, dd, 4.2, 13.6)	39.3	3.45 (1H, dd, 6.0, 13.5)			
		3.21 (1H, dd, 6.6, 13.6)		3.36 (1H, dd, 7.5, 13.5)			
8''	173.5	—	173.4	—			
9''	22.6	1.98 (3H, s)	22.5	1.93 (3H, s)			
10''	43.1	2.51 (3H, s)	36.8	2.48 (3H, s)			

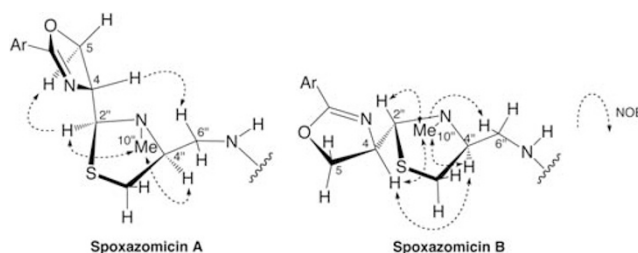
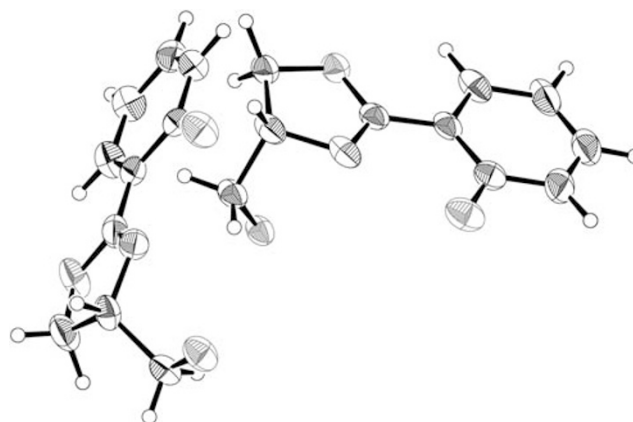
^1H NMR: 300 MHz, ^{13}C NMR: 75 MHz, All compounds were measured in CD_3OD .


Figure 2 ^1H - ^1H COSY and HMBC correlations of spoxazomicins A (1) and C (3).

and two sp^3 methylene carbons. The ^1H and ^{13}C NMR spectra of **3** were similar to those of the 2-phenyl-4,5-dihydrooxazole moiety of **1**. The 2D NMR spectra indicated that the structure of **3** was as shown in Figure 2 and **3** was designated as spoxazomicin C. The structure was confirmed by single-crystal X-ray crystallographic analysis using the crystals obtained from a mixed solution of EtOAc/hexane (1:3). The ORTEP drawing is shown in Figure 4.

Biological activities

In vitro antitrypanosomal activities of **1–3** were evaluated together with the activities of some standard antitrypanosomal drugs (Table 3). Among the tested spoxazomicins, **1** showed the highest antitrypanosomal activity against the GUTat 3.1 strain of *T. b. brucei*, with an


Figure 3 NOE correlations of spoxazomicins A (1) and B (2).

Figure 4 ORTEP (Oak Ridge Thermal Ellipsoid Plot Program) drawing for spoxazomicin C (3).

IC_{50} value of $0.11 \mu\text{g ml}^{-1}$, which has 14–21-fold more potent activity than those of the clinically used antitrypanosomal drugs, suramin and eflornithine. Compound **2** also showed comparatively potent antitrypanosomal activity, with an IC_{50} value of $0.55 \mu\text{g ml}^{-1}$, but **3**

Table 3 *In vitro* antitrypanosomal activity against *T. b. b.* GUTat 3.1 and cytotoxicity in human MRC-5 cells of spoxazomicins and two commonly used antitrypanosomal drugs

Compound	IC_{50} ($\mu\text{g ml}^{-1}$)		
	Antitrypanosomal activity <i>T. b. b.</i> GUTat 3.1	Cytotoxicity MRC-5	Selectivity index MRC-5/ <i>T. b. b.</i>
Spoxazomicin A (1)	0.11	27.84	253.1
Spoxazomicin B (2)	0.55	21.11	38.4
Spoxazomicin C (3)	3.37	14.02	3.8
Eflornithine ^a	2.27	> 100	> 44.1
Suramin ^a	1.58	> 100	> 63.3

Abbreviation: *T. b. b.*, *Trypanosoma brucei brucei*.^aClinically used antitrypanosomal drugs.

showed weak antitrypanosomal activity, with an IC_{50} value around $3 \mu\text{g ml}^{-1}$.

The cytotoxicity of spoxazomicins was evaluated against a human diploid embryonic cell line (MRC-5). The compounds **1** and **2** were weakly cytotoxic with IC_{50} values of $21\text{--}28 \mu\text{g ml}^{-1}$, and **3** showed an IC_{50} value of $14 \mu\text{g ml}^{-1}$. To compare the overall antitrypanosomal activity and cytotoxicity, we introduced a selectivity index (SI: cytotoxicity (IC_{50} for the MRC-5 cells)/antitrypanosomal activity (IC_{50} for the GUTat 3.1 strain)), as presented in Table 3. The SI of **1** was the highest, with a ratio of 253. Compound **2** showed a medium SI, with the ratio of 38, whereas **3** showed the lowest SI with a ratio of 3.8.

Compounds **1–3** showed no antibacterial and antifungal activities at $10 \mu\text{g}$ on the paper disc method against the following microorganisms: *Staphylococcus aureus* (ATCC6538p), *Bacillus subtilis* (ATCC6633), *Kocuria rhizophila* (ATCC9341), *Mycobacterium smegmatis* (ATCC607), *Escherichia coli* NIHJ (KB213), *Escherichia coli* NIHJ JC-2 (IFO12734), *Pseudomonas aeruginosa* (IFO3080), *Xanthomonas campestris* pv. *oryzae* (KB88), *Proteus mirabilis* (NBRC3849), *Proteus vulgaris* (NBRC3167), *Bacteroides fragilis* (ATCC23745), *Candida albicans* (KF1), *Saccharomyces cerevisiae* (ATCC9763), *Aspergillus niger* (ATCC6275), *Pyricularia oryzae* (KF180) and *Mucor racemosus* (IFO4581).

DISCUSSION

We have isolated new types of pyochelin family antibiotics (**1–3**) from a new endophytic actinomycete species, *Streptosporangium oxazolanicum* K07-0460^T. Antitrypanosomal activity of **1** and **2** was much more potent and selective than **3**. Therefore, the 4-(acetylamino) methyl-3-methyl-2-thiazolidine moiety might be important for conferring activity and selectivity. The antitrypanosomal activities of **1** was substantially more potent than **2**, which suggested the S^* configuration of C-2'' might have a significant role.

Pyochelin family antibiotics have been reported to have various activities. Pyochelin was an iron-chelating growth promoter for *Pseudomonas aeruginosa*.⁵ Thiazostatins A and B had antioxidant activities.⁶ Watasemycins A and B had anti-*Proteus mirabilis* activity and weak antibacterial activities against some Gram-positive bacteria.⁷ However, this is the first report of antitrypanosomal activities of pyochelin family antibiotics. The results reveal that **1** and **2** are the promising lead compounds for developing a new type of antitrypanosomal drugs. Further investigations of the antitrypanosomal and other biological activities of **1–3** are in progress.

METHODS

General

¹H, ¹³C and 2D NMR spectra were measured in CD₃OD using a Varian XL-300 spectrometer (Varian, Palo Alto, CA, USA). NOE experiments were measured using a Varian Inova 600 spectrometer. Chemical shifts were expressed in p.p.m. and were referenced to CD₃OD (3.31 p.p.m.) in the ¹H NMR spectra and to CD₃OD (49.0 p.p.m.) in the ¹³C NMR spectra. FAB-MS and EI-MS spectra were measured on a JEOL JMS AX-505 HA mass spectrometer (JEOL, Akishima, Japan). IR spectra (KBr) were taken on a Horiba FT-210 Fourier transform Infrared spectrometer (Horiba, Kyoto, Japan). UV spectra were measured with a Beckman DU640 spectrophotometer (Beckman, Fullerton, CA, USA). Optical rotation was measured on a JASCO model DIP-181 polarimeter (JASCO, Hachioji, Japan). X-ray crystal analysis was measured on a RASA-5R (Rigaku, Akishima, Japan).

Fermentation

The strain K07-0460 was grown and maintained on the agar slant containing starch 1.0%, NZ amine 0.3%, yeast extract 0.1%, meat extract 0.1%, CaCO₃ 0.3% and agar 1.5 % (adjusted to pH 7.0 before sterilization). A loop of cells of strain K07-0460 was inoculated into 10 ml of seed medium consisting of starch 2.4%, glucose 0.1%, peptone 0.3%, meat extract 0.3%, yeast extract 0.5% and CaCO₃ 0.3% (adjusted to pH 7.0 before sterilization) in a test tube. The test tube was incubated on a rotary shaker (300 r.p.m.) at 27 °C for 6 days. A 1-ml portion of the first seed culture was transferred to a 500-ml Erlenmeyer flask containing 100 ml of the seed medium (the same components described above), and the fermentation was carried out on a rotary shaker (210 r.p.m.) at 28 °C for 3 days.

For production of **1–3**, a 1-ml portion of the second seed culture was transferred to 500-ml Erlenmeyer flasks (total 42 flasks), each containing 200 ml of production medium (the same components as the seed medium), and the fermentation was carried out on a rotary shaker (210 r.p.m.) at 28 °C for 4 days.

Antitrypanosomal and cytotoxic activity *in vitro*

In vitro antitrypanosomal activities against *Trypanosoma brucei brucei* strain GUTat 3.1 and cytotoxicity against human diploid embryonic cell line MRC-5 were measured as described previously.⁸

Antimicrobial activity

Antimicrobial activity was measured by a paper disk method (6 mm, Advantec, Tokyo, Japan) as described previously.⁹

Crystallographic analysis

The crystal contained two molecules of **3**, $2(C_{10}H_{11}NO_3)$, was the monoclinic space group C2(#5) with $a=28.736(5)$ Å, $b=4.714(4)$ Å, $c=14.117(6)$ Å, $\beta=90.0000^\circ$, $V=1912(1)$ Å³, $Z=8$, $d_{\text{calc}}=1.342$ g cm⁻³, $\mu=8.35$ cm⁻¹ and $T=296$ K. X-ray intensity data were collected on a Rigaku AFC5R diffractometer employing graphite-monochromated Cu K α radiation ($\lambda=1.54178$ Å) and the ω -2 θ scan technique. The structure was solved by direct methods. For refinement, 2070 unique reflections with $F^2 > 2.0\sigma(F^2)$ were used. Full-matrix least-squares refinement were based on F^2 , minimizing the quantity $\sum w(F_o^2 - F_c^2)^2$ with $w=1/\sigma^2(F_o^2)$, GOF=1.20, $R1=0.052$ and $Rw=0.152$. Crystallographic data of **3** has been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 812009. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (Fax: C44(0)-1223-336033 or E-mail: deposit@ccdc.cam.ac.uk).

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