

Chemical Modification of Pseurotin A: One-pot Synthesis of Synerazol and Pseurotin E and Determination of Absolute Stereochemistry of Pseurotin E

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Abstract Two natural products, synerazol and pseurotin E, were synthesized from the natural product pseurotin A in 58% and 57% yields, respectively in one-pot procedures. This work also establishes the absolute stereochemistry of pseurotin E.

Keywords pseurotin A, synerazol, pseurotin E, chemical modification, absolute stereochemistry

Pseurotin A (**1**) is a microbial secondary metabolite isolated from the fermentation broth of *Pseudeurotium ovalis* STOLK (Ascomycetes) in 1976 [1]. It has various

biological activities, including induction of cell differentiation [2], inhibition of chitin synthase [3], inhibition of monoamine oxidase [4], and apomorphine-antagonistic activity [5]. Pseurotin A (**1**) possesses a highly substituted 1-oxa-7-azaspiro[4,4]non-2-ene-4,6-dione skeleton. The structure and absolute stereochemistry of **1** have been determined by a single-crystal X-ray analysis of its 12,13-dibromo derivative [6]. The core structure of **1** is found in other natural products, too (Fig. 1). These pseurotin-related natural products also possess interesting biological activities, including apomorphine-antagonistic activity in the cases of pseurotin D [5] and F [7], anti-angiogenic activity [8a], antifungal activity [8b] and synergistic activity [8b] in the case of synerazol (**2**) isolated

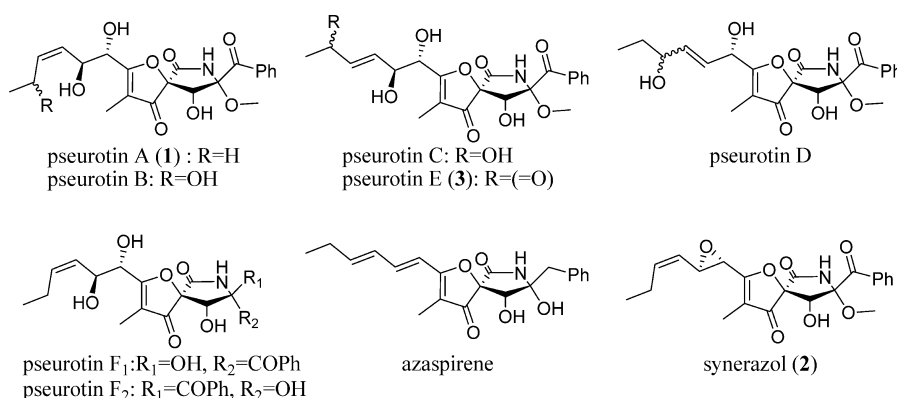


Fig. 1 Structure of pseurotin-related natural products.

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from the fermentation broth of *Aspergillus fumigatus* SANK 10588 in 1991 [8b], and anti-angiogenic activity in the case of azaspirene [9]. Another member of the pseurotin family, pseurotin E (**3**), was isolated from the fermentation broth of *Pseudeurotium ovalis* STOLK in 1981 [10], though its absolute stereochemistry and biological activity were not established.

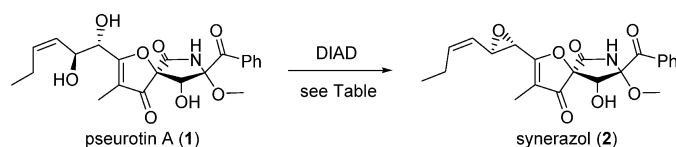
Elegant total syntheses of **1** [11, 12], pseurotin F₂ [11, 12], **2** [13] and azaspirene [14, 12] have been reported by Hayashi *et al.* and Tadano *et al.* However, these syntheses involved many steps, and are unsuitable for large-scale application to provide samples for biological testing. On the other hand, the supply of pseurotin-related natural products from fermentation is also limited, except for **1**. The reported fermentation yields are 40 mg/liter for **1** [10], 1.4 mg/liter for **2** [8b], and 6 mg/liter for **3** [10]. Hence, we thought chemical modification of **1** might be an attractive way to obtain large amounts of pseurotin-related natural products. In this paper, we wish to report one-pot synthesis of synerazol (**2**) and pseurotin E (**3**) from pseurotin A (**1**) and determination of the absolute stereochemistry of **3**. Various chemical modifications of pseurotin-related natural products, including acetylation (pseurotin A [1] and synerazol [16]), hydrogenation (pseurotin A [15], B [10], C [10]), oxidative cleavage (pseurotin A [1], B [10], C [10]),

dibromination [1] (pseurotin A), formation of acetonide [1] (pseurotin A), reduction of carbonyl group [15] (pseurotin A), and synthesis of pseurotin A from synerazol for the purpose of the determination of its absolute stereochemistry [16], have been reported.

Compound **1** was obtained from the fermentation broth of *Aspergillus* sp. The synthesis of **2** from **1** is illustrated in Table 1. First of all, epoxidation of the 1,2-diol of **1** under the reported Mitsunobu condition [17] did not afford **2** (entry 1). However, this reaction proceeded when Cy₃P was replaced with Ph₃P (entry 2). To optimize the reaction, the effects of solvents and temperature were examined. Dichloroethane (DCE) gave a better yield (entry 3). Decreasing the temperature reduced the formation of minor by-products (58% yield, entry 4). Synthetic **2** exhibited properties identical to those of the natural product [8b] (¹H-NMR, ¹³C-NMR, and optical rotation [18]). We were able to obtain **2** on a 200 mg scale.

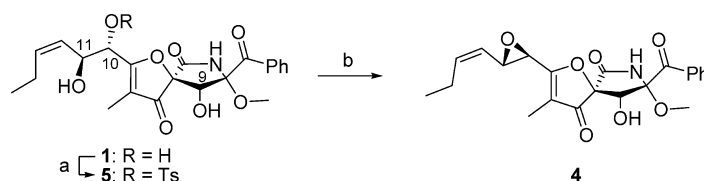
We decided to synthesize the (10*R*,11*S*)-diastereoisomer of **2** to confirm the diastereoselectivity in the synthesis of **2**. The (10*R*,11*S*)-diastereoisomer **4** is generated if the hydroxyl group at the 10 position is eliminated under the above Mitsunobu conditions. NMR spectra of the (10*R*,11*S*)-diastereoisomer **4** have not been reported, although its total synthesis was achieved [13]. Selective

Table 1 Synthesis of synerazol by Mitsunobu reaction



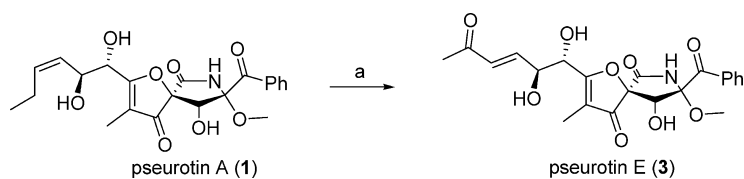
Entry	Phosphine	Solvents	Temp (°C)	Time (hours)	Yield of 2 (%)
1	Cy ₃ P	THF	22 to 40	25	0
2	Ph ₃ P	THF	22 to 60	28	14
3	Ph ₃ P	DCE	30	4.5	43
4	Ph ₃ P	DCE	15	7	58
5	Ph ₃ P	DCM	15	3	54

DIAD: diisopropyl azodicarboxylate; Cy: cyclohexyl; DCE: dichloroethane.



Scheme 1 Synthesis of **4**^a.

^a Reagents: (a) TsCl, Et₃N, *n*-Bu₂SnO, DCM, rt, 17 hours, 33%; (b) K₂CO₃, MeOH, 4 °C, 2 hours, 27%.



Scheme 2 Synthesis of pseurotin E^a.

^a Reagents: (a) Grubbs catalyst (second generation), methyl vinyl ketone, DCM, 40°C, 2.5 hours, 57%.

tosylation reaction of the 1,2-diol of **1** was achieved in the presence of dibutyltin oxide [19] to give **5** (Scheme 1). Regioselectivity of the tosyl group was assigned on the basis of ¹H-NMR multiplicity (H-C11 (ddd, *J*=5.1, 7.8, 9.5 Hz), H-C10 (d, *J*=7.8 Hz), and H-C9 (d, *J*=12.4 Hz)). Epoxidation under basic conditions gave **4**. The optical rotation of **4** ($[\alpha]_D^{26} -53.9^\circ$ (*c* 0.075, CHCl₃)) is in good agreement with literature data [13] ($[\alpha]_D^{22} -49.2^\circ$ (*c* 0.02, CHCl₃)). The (10*R*,11*S*)-diastereomer **4** [20] proved readily distinguishable from **2** by ¹H-NMR, ¹³C-NMR, and HPLC, thereby confirming the complete diastereoselectivity of the transformation leading to **2**.

Next, we envisioned that **3** could be derived from **1** by olefin cross metathesis. Treatment of **1** with methyl vinyl ketone and second-generation Grubbs catalyst (1,3-bis-(2,4,6-trimethylphenyl)-2-imidazolidinylidene)dichloro(*o*-isopropoxyphenylmethylene)ruthenium) [21] afforded **3** in 57% yield (Scheme 2). The ¹H-NMR and ¹³C-NMR spectra of synthetic **3** [22] were in good accordance with those reported for the natural product [10]. Hence, the absolute stereochemistry of pseurotin E (**3**) was defined. This metathesis was also applicable to the synthesis of other pseurotin E analogues (data not shown).

In summary, we have developed an efficient one-pot synthesis of synerazol and pseurotin E from readily available pseurotin A. This work also establishes the absolute stereochemistry of pseurotin E. We will report syntheses of pseurotin A, synerazol and various pseurotin E analogs, as well as their biological activities, in due course.

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References

- Bloch P, Tamm C, Bollinger P, Petcher TJ, Weber HP. Pseurotin, a new metabolite of *Pseudeurotium ovalis* Stolk having an unusual hetero-spirocyclic system. *Helv Chim Acta* 59: 133–137 (1976)
- Komagata D, Fujita S, Yamashita N, Saito S, Morino, T. Novel neurotogenic activities of pseurotin A and penicillic acid. *J Antibiot* 49: 958–959 (1996)
- Wenke J, Anke H, Sterner O. Pseurotin A and 8-*O*-demethylpseurotin A from *Aspergillus fumigatus* and their inhibitory activities on chitin synthase. *Biosci Biotech Biochem* 57: 961–964 (1993)
- Maebayashi Y, Horie Y, Satoh Y, Yamazaki M. Isolation of pseurotin A and a new pyrazine from *Pseudallescheria boydii*. *Mycotoxins* 22: 33–34 (1985)
- Wink J, Grabley S, Gareis M, Zeeck A, Phillips S. Biologically active pseurotin A and D, new metabolites from *Aspergillus fumigatus*, process for their preparation and their use as apomorphine antagonists. *Eur Pat Appl EP546475*, (1993)
- Weber HP, Petcher TJ, Bloch P, Tamm C. The crystal and molecular structure of 12,13-dibromopseurotin. *Helv Chim Acta* 59: 137–140 (1976)
- Wink J, Grabley S, Gareis M, Thiericke R, Kirsch R. Pseurotin F1/F2, new metabolites from *Aspergillus fumigatus*, process for their preparation and their use as apomorphine antagonists. *Eur Pat Appl EP546474*, (1993)
- (a) Igarashi Y, Yabuta Y, Sekine A, Fujii K, Harada K, Oikawa T, Sato M, Furumai T, Oki T. Directed biosynthesis of fluorinated pseurotin A, synerazol and gliotoxin. *J Antibiot* 57: 748–754 (2004)
(b) Ando O, Satake H, Nakajina M, Sato A, Nakamura T, Kinoshita T, Furuya K, Haneishi T. Synerazol, a new antifungal antibiotic. *J Antibiot* 44: 382–389 (1991)
- Asami Y, Kakeya H, Onose R, Yoshida A, Matsuzaki H, Osada H. Azaspirene: a novel angiogenesis inhibitor containing a 1-oxa-7-azaspiro[4.4]non-2-ene-4,6-dione skeleton produced by the fungus *Neosartorya* sp. *Org Lett* 4: 2845–2848 (2002)
- Breitenstein W, Chexal KK, Mohr P, Tamm C. Pseurotin B, C, D, and E. Further new metabolites of *Pseudeurotium ovalis* STOLK. *Helv Chim Acta* 64: 379–388 (1981)
- Hayashi Y, Shoji M, Yamaguchi S, Mukaiyama T, Yamaguchi J, Kakeya H, Osada H. Asymmetric total synthesis of pseurotin A. *Org Lett* 5: 2287–2290 (2003)
- (a) Aoki S, Oi T, Shimizu K, Shiraki R, Takao K, Tadano K. Total syntheses of natural pseurotins A and F₂ and azaspirene. *Heterocycles* 62: 161–166 (2004)
(b) Aoki S, Oi T, Shimizu K, Shiraki R, Takao K, Tadano K.

- Total syntheses of natural pseurotins A, F₂, and azaspirene. *Bull Chem Soc Jpn* 77: 1703–1716 (2004)
13. Hayashi Y, Shoji M, Mukaiyama T, Gotoh H, Yamaguchi S, Nakata M, Kakeya H, Osada H. First asymmetric total synthesis of synerazol, an antifungal antibiotic, and determination of its absolute stereochemistry. *J Org Chem* 70: 5643–5654 (2005)
 14. Hayashi Y, Shoji M, Yamaguchi J, Sato K, Yamaguchi S, Mukaiyama T, Sakai K, Asami Y, Kakeya H, Osada H. Asymmetric total synthesis of (–)-azaspirene, a novel angiogenesis inhibitor. *J Am Chem Soc* 124: 12078–12079 (2002)
 15. Mohr P, Tamm C. Biosynthesis of pseurotin A. *Tetrahedron* 37: 201–212 (1981)
 16. Igarashi Y, Yabuta Y, Furumai T. Determination of the absolute configuration of synerazol. *J Antibiot* 57: 537–540 (2004)
 17. Weissman SA, Rossen K, Reider PJ. Stereoselective synthesis of styrene oxides via a Mitsunobu cyclodehydration. *Org Lett* 3: 2513–2515 (2001)
 18. Optical rotation of synthetic **2**: $[\alpha]_{\text{D}}^{25} +24.0^{\circ}$ (*c* 0.15, CHCl₃), natural product [8b]: $[\alpha]_{\text{D}}^{25} +22.9^{\circ}$ (*c* 0.55, CHCl₃)
 19. Martinelli MJ, Nayyar NK, Moher ED, Dhokte UP, Pawlak JM, Vaidyanathan R. Dibutyltin Oxide Catalyzed Selective Sulfonylation of α -Chelatable Primary Alcohols. *Org Lett* 1: 447–450 (1999)
 20. ¹H-NMR (400 MHz, CDCl₃) δ 1.03 (3H, t, *J*=7.6 Hz), 1.82 (3H, s), 2.16–2.32 (2H, m), 3.39 (3H, s), 3.73 (1H, d, *J*=1.9 Hz), 4.02 (1H, ddd, *J*=0.7, 2.0, 7.8 Hz), 4.07 (1H, d, *J*=12.7 Hz), 4.63 (1H, d, *J*=12.5 Hz), 5.07 (1H, ddt, *J*=1.5, 9.0, 10.7 Hz), 5.85 (1H, br dt, *J*=11.0, 7.8 Hz), 7.31 (1H, br s), 7.50 (2H, br dd, *J*=7.3, 8.3 Hz), 7.65 (1H, br t, *J*=7.3 Hz), 8.29 (2H, dd, *J*=1.3, 8.3 Hz); ¹³C-NMR (100 MHz, CDCl₃) δ 5.1, 14.0, 21.3, 51.7, 53.1, 55.1, 74.0, 89.4, 91.8, 113.8, 123.6, 128.8, 130.5, 132.4, 134.7, 141.3, 164.9, 182.2, 194.2, 196.8; $[\alpha]_{\text{D}}^{26} -53.9^{\circ}$ (*c* 0.075, CHCl₃), HPLC (Inertsil ODS-2) retention time: 14.9 minutes (synerazol), 15.7 minutes (**4**).
 21. Chatterjee AK, Grubbs RH. Formal vinyl C–H activation and allylic oxidation by olefin metathesis. *Angew Chem Int Ed* 41: 3171–3174 (2002)
 22. ¹H-NMR (400 MHz, CDCl₃) δ 1.70 (3H, s), 2.24 (3H, s), 3.42 (3H, s), 3.62 (1H, br d, *J*=7.8 Hz), 3.92 (1H, br d, *J*=7.6 Hz), 4.20 (1H, d, *J*=12.2 Hz), 4.62–4.68 (1H, m), 4.68 (1H, d, *J*=12.2 Hz), 4.75 (1H, br t, *J*=6.1 Hz), 6.42 (1H, dd, *J*=1.7, 15.7 Hz), 6.88 (1H, dd, 4.4, *J*=15.8 Hz), 7.51 (2H, br dd, *J*=7.8, 8.0 Hz), 7.66 (1H, br t, *J*=7.4 Hz), 7.92 (1H, br s), 8.31 (2H, br d, *J*=8.0 Hz); ¹³C-NMR (100 MHz, acetone-*d*₆) δ 5.7, 27.2, 52.2, 72.2, 72.8, 75.4, 92.5, 92.8, 113.8, 129.3, 131.3, 131.5, 134.5, 134.7, 145.5, 167.3, 186.2, 196.2, 197.5, 198.0; $[\alpha]_{\text{D}}^{22} +77.4^{\circ}$ (*c* 0.31, CHCl₃).