

Fogacin, a Novel Cyclic Octaketide Produced by *Streptomyces* Strain Tü 6319[†]

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Abstract A new octaketide named fogacin (**1**) was isolated from *Streptomyces* sp. (strain Tü 6319). Furthermore two shunt metabolites, SEK4b (**2**) and anhydroSEK4b (**3**), were detected and identified as non-enzymatically cyclized products of polyketide intermediates built during the biosynthesis of actinorhodin. SEK4b (**2**) as well as anhydroSEK4b (**3**) were previously described as metabolites of genetically engineered strains.

Keywords polyketides, biosynthesis, shunt metabolites, SEK4b, HPLC screening

Freshly isolated strains from soils collected at various sites in Romania were included in our HPLC-diode array screening program to detect novel secondary metabolites. Strain Tü 6319 which was isolated from an industrial contaminated soil near Fogaras, Romania, drew our attention because of its characteristic metabolite pattern, which appears analyzing the culture filtrate extract. Three of the metabolites were identified by means of our HPLC-UV-Vis database [2] as SEK4b (**2**) [3], and germicidins A and B [4], respectively. Two further metabolites did not correspond to any of the 834 reference compounds stored in our database. One of them was identified as anhydroSEK4b (**3**) [5], the second was named fogacin (**1**).

Strain Tü 6319 was examined for a number of key properties known to be of value in streptomycete systematics. The presence of LL-diaminopimelic acid in the peptidoglycan together with its colonial characteristics

allowed its assignment to the genus *Streptomyces*. Strain Tü 6319 shared a high 16S rRNA gene sequence with the closely related type strains *Streptomyces coelestis* DSM 40421, *S. violaceolatus* DSM 40438 and *S. violaceoruber* DSM 40049.

Batch fermentation of strain Tü 6319 was carried out in 10-liter stirred tank fermenters (New Brunswick) in a medium that consisted of oat meal 2.0% in tap water (pH 7.3). The fermentation was conducted at 27°C for 168 hours with an aeration rate of 0.5 v/v/m and agitation of 200 rpm. Addition of 3% DMSO to the medium increased the production of **1** by 33%, reaching a maximum value of 10 mg/liter after 168 hours. Compound **1** was isolated from the culture filtrate (13 liters), loaded onto an Amberlite XAD-16 column (8×40 cm) and eluted by increasing concentrations of MeOH. The 40% MeOH fraction that contained **1** was concentrated *in vacuo*, adjusted to pH 2, and extracted with ethyl acetate. The raw material was separated on a LiChroprep Diol column (2.6×40 cm; Merck) applying a linear gradient CH₂Cl₂ to a content of 5% MeOH within 3 hours at a flow rate of 5 ml/minute. A further metabolite (**3**) was separated from **1**, and both compounds were purified by subsequent column chromatography on Sephadex LH-20 and Fractogel TSK HW-40 (each column 1.5×90 cm, flow rate 15 ml/hour) using MeOH as eluent. Pure **1** was obtained in an amount of 8 mg.

The antimicrobial activities of fogacin (**1**) and anhydro SEK4b (**3**) were examined by an agar plate diffusion assays. Compound **3** revealed a moderate inhibitory activity against *Bacillus subtilis* DSM 10 and

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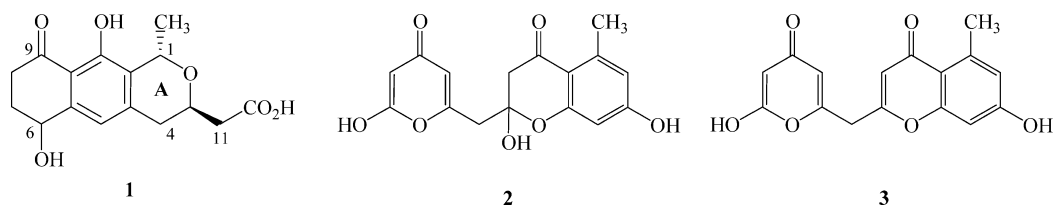


Fig. 1 Structures of fogacin (**1**) (relative stereochemistry of ring A shown), SEK4b (**2**) and anhydroSEK4b (**3**).

S. viridochromogenes Tü 57, showing inhibition zones of 11 mm and 16 mm, respectively, at concentrations of 1.0 mg/ml. Gram-negative bacteria, yeasts and filamentous fungi were not sensitive to **1** and **3**.

Fogacin (**1**) was isolated as colourless solid. The ESI-MS (positive mode) displayed a peak at m/z 307 ($M+H$)⁺, high resolution of it leads to the molecular formula C₁₆H₁₈O₆. The ¹³C NMR spectrum of **1** shows 16 signals among them a carbonyl group (δ_C 206.1) and a carboxylic acid (δ_C 176.0) besides six aromatic carbons. In the region of aliphatic carbon atoms three oxygen bound methine, four methylene groups and one methyl group can be observed. In accordance the ¹H-NMR spectrum of **1** shows besides four diastereotopic methylene and three highly coupled methine signals one methyl group as doublet and only one aromatic proton as singlet. ¹H-COSY and HMBC spectra as well as the values of the coupling constants of **1** prove the direct vicinity of 6-H, 7-H_{a,b} and 8-H_{a,b} with a quasi equatorial position of 6-OH. The connection between the rings is revealed by HMBC-couplings shown in Fig. 2. Additionally an 1D-NOE experiment exhibits a strong coupling between 3-H and the methyl group, while no coupling can be observed between 3-H and 1-H. Furthermore, 1-H shows no coupling to any proton except that of the methyl group. Thus, the relative stereochemistry of ring A of **1** is assigned to be *trans*. Compound **1** represents the framework of one half of actinorhodin (**4**), but differs in its structure from all known intermediates which were identified along the biosynthetic route of **4** [3].

Octaketides like **2** and **3** are known shunt metabolites of the actinorhodin biosynthetic pathway (Fig. 3) which are often found in experiments with streptomycetes mutants bearing manipulations in the cyclase functions of type II polyketide synthases (PKS) [6, 7] and are under ongoing investigations [8]. In these experiments **2** and **3** turned out to be important key metabolites for the elucidation of the biosynthetic pathway of **4**. They are built by a so called ‘minimal PKS’ and products of subsequent non-enzymatic cyclization of the linear polyketide. In contrast, substance **1** has to be a product of stereospecific cyclization and miscellaneous tailoring steps (*e.g.* reduction and hydroxylation) like actinorhodin and other aromatic

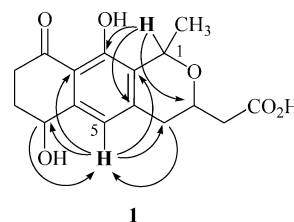


Fig. 2 Important HMBC-correlations of fogacin (**1**): ³J_{C,H} couplings reveal the regiochemistry of the ring systems.

polyketides [9]. Decaketides of similar ‘biosynthetic’ origin like **2** and **3** have been described not only from engineered biosynthesis but also from a wild-type strain [10]. Nevertheless **2** and **3** are the first shunt metabolites with an octaketide backbone isolated from a wild-type strain. The question why nature creates ‘shunt metabolites’ as those presented here remains open. Maybe this is part of evolutionary processes within the producing strain, which cover deletion or insertion of single genes (or gene functions). As a result **2** and **3** are related to substances isolated from man-made mutant strains.

Fogacin (**1**)

A colourless solid. $[\alpha]_D^{20} +7$ (c 1, MeOH); UV λ_{max}^{MeOH} nm (ϵ) 333 (900), 272 (2550), 222 (4260); $\lambda_{max}^{0.01 M HCl-MeOH}$ nm (ϵ) 331 (1050), 271 (2530), 221 (4390); $\lambda_{max}^{0.01 M NaOH-MeOH}$ nm (ϵ) 333 (1290), 271 (3120), 223 (4660); CD $\lambda_{extreme}^{MeOH}(\theta)$ deg·cm²·decimole⁻¹ 268 (−4190), 214 (8060); IR ν_{max} cm⁻¹ 3428 (br), 2937, 2354, 1707, 1625, 1370, 1264, 1155, 1085; ¹H NMR (300 MHz, CD₃OD) δ 6.86 (1H, s, 5-H), 5.06 (1H, q, $J_{1,3}$ =6.5 Hz, 1-H), 4.81 (1H, dd, $J_{7a,b}$ =8.0, 3.5 Hz, 6-H), 4.43 (1H, m, 3-H), 2.89 (1H, ddd, $J_{7a,b;8a}$ =17.0, 9.0, 4.5 Hz, 8-H_b), 2.85 (1H, m, 4-H_b), 2.68 (1H, m, 4-H_a), 2.65 (1H, ddd, $J_{7a,b;8b}$ =17.5, 9.0, 4.5 Hz, 8-H_a), 2.56 (2H, m, 11-H₂), 2.29 (1H, m, 7-H_b), 2.10 (1H, m, 7-H_a), 1.55 (3H, d, J_1 =6.5 Hz 13-H₃); ¹³C-NMR (75.5 MHz, CD₃OD) δ 206.1 (s, C-9), 176.0 (s, C-12), 159.7 (s, C-10), 145.9 (s, C-5a), 144.1 (s, C-4a), 127.8 (s, C-10a), 119.1 (d, C-5), 114.2 (s, C-9a), 69.3 (d, C-1), 68.1 (d, C-6), 65.4 (d, C-3), 42.9 (t, C-11), 35.8 (t, C-8), 35.4 (t, C-4), 32.5 (t, C-7), 19.3 (q, C-13); ESI-MS m/z 305

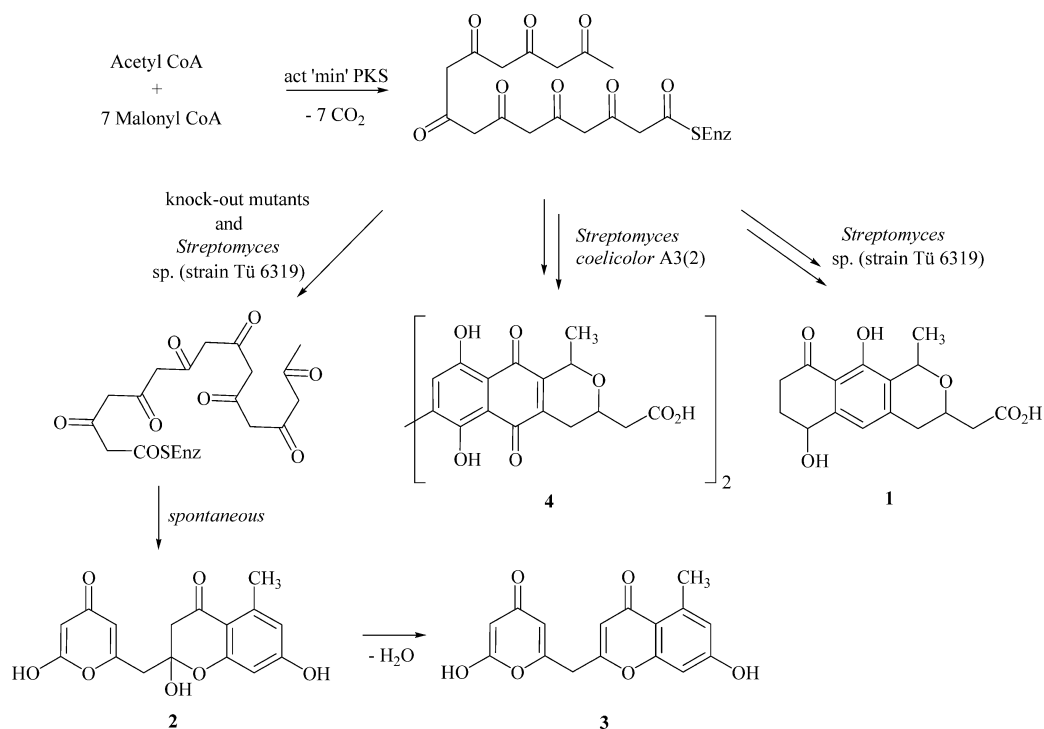


Fig. 3 Putative biosynthesis and folding pattern of fogacin (**1**), SEK4b (**2**) and anhydroSEK4b (**3**) in comparison to actinorhodin (**4**).

(M-H)⁻; HRESI-MS m/z 307.11763 ([M+H]⁺, C₁₆H₁₉O₆, δ_m 0.07 mmu).

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