

ORIGINAL ARTICLE

Strong effect of climate on ectomycorrhizal fungal composition: evidence from range overlap between two mountains

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Separating the effects of environmental factors and spatial distance on microbial composition is difficult when these factors covary. We examined the composition of ectomycorrhizal (EM) fungi along elevation gradients on geographically distant mountains to clarify the effect of climate at the regional scale. Soil cores were collected from various forest types along an elevation gradient in southwestern Japan. Fungal species were identified by the internal transcribed spacer regions of the rDNA using direct sequencing. The occurrence of fungal species in this study was compared with a previous study conducted on a mountain separated by ~550 km. In total, we recorded 454 EM fungi from 330 of 350 soil cores. Forty-seven fungal species (~20% of the total excluding singletons) were shared between two mountains, mostly between similar forest types on both mountains. Variation partitioning in redundancy analysis revealed that climate explained the largest variance in EM fungal composition. The similarity of forest tree composition, which is usually determined by climatic conditions, was positively correlated with the similarity of the EM fungal composition. However, the lack of large host effects implied that communities of forest trees and EM fungi may be determined independently by climate. Our data provide important insights that host plants and mutualistic fungi may respond to climate change idiosyncratically, potentially altering carbon and nutrient cycles in relation to the plant–fungus associations.

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Introduction

Ectomycorrhizal (EM) fungi establish obligate mutualistic relationships with many ecologically and economically important tree species, including Pinaceae, Fagaceae, Betulaceae and Dipterocarpaceae (Taylor and Alexander, 2005). EM fungi play a significant role in global nutrient and carbon cycles by enhancing the nutrient and water uptake by trees and absorbing carbon from their hosts (Smith and Read, 2008). Temperate and boreal forests, which cover ~14% of the land surface (FAO, 2012), harbor hundreds of taxonomically and functionally diverse EM fungi (Read and Perez-Moreno, 2003; Tedersoo *et al.*, 2010; Koide *et al.*, 2014). Recent environmental change may have altered fungal communities, which would subsequently affect the associated plant communities and ecosystem functioning (Parrent *et al.*, 2006;

Johnson *et al.*, 2013; Dickie *et al.*, 2014). Clarifying the drivers of the community structure of EM fungi is critical for predicting the effects of environmental change on forest ecosystems from local to global contexts (Lilleskov and Parrent, 2007).

Many studies have attempted to clarify the drivers of EM fungal composition at various scales and ecosystems. The host plant is regarded as one of the most important factors that influences EM fungal composition, especially at the stand scale, where hosts coexist in relatively narrow spatial and environmental ranges (Kennedy *et al.*, 2003; Richard *et al.*, 2005; Ishida *et al.*, 2007; Tedersoo *et al.*, 2008; Smith *et al.*, 2009; Murata *et al.*, 2013). However, isolating the host effect at larger spatial scales is difficult because the host composition changes with climatic conditions and geological history (Tedersoo *et al.*, 2012). In addition, EM fungal communities may be affected by climate (Bahram *et al.*, 2012) and soil (Cox *et al.*, 2010; Jarvis *et al.*, 2013; Roy *et al.*, 2013). Moreover, EM fungal communities often exhibit spatial structures (Lilleskov *et al.*, 2004; Bahram *et al.*, 2013a; Miyamoto *et al.*, 2014), which may be driven by autocorrelations in environmental factors

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(Toljander *et al.*, 2006; Tedersoo *et al.*, 2012) or independently by fungal internal factors (e.g., dispersal; Peay *et al.*, 2007). Thus, at larger regional and global scales, whether EM fungal communities are differentiated by hosts, environmental factors or geographical distance is less clear. Distinguishing the effects of these factors is critical to determine if fungal distributions are primarily constrained by contemporary environmental variations or by past dispersal events (Martiny *et al.*, 2006). Systematic sampling strategies are needed to address the relative importance of these factors at larger spatial scales.

The community structures of EM fungi are often examined by standard ordination and distance methods (Bahram *et al.*, 2012; Pöhlme *et al.*, 2013). However, unlike plant and animal community studies, the distribution and overlap of individual species of soil fungi between sites has rarely been reported, mainly because soil fungi are difficult to detect and quantify accurately in the field without applying molecular analyses. In addition, many EM fungi are observed rarely and sporadically because of the inherently limited sampling efforts relative to the entire soil volume (Taylor, 2002). Our previous study (Miyamoto *et al.*, 2014) demonstrated that intensive sampling (many samples from each community) is effective for recording a relatively large number of fungal taxa in a community and for identifying species overlaps among study sites. Consistent sampling across study sites also helps to remove the variations imposed by methodological discrepancies found in many meta-analyses (Nakagawa and Santos, 2012; Tedersoo *et al.*, 2014). A detailed examination of species overlaps among communities improves the chances of detecting distribution patterns, community structures and the underlying mechanisms of organisms (Leibold and Mikkelsen, 2002; Presley *et al.*, 2010; Thébault, 2013).

Here, we established study sites along an elevation gradient from a warm temperate to boreal climate on a mountain and examined the EM fungal

community at each site. The data collected were combined with the data from our previous study conducted on Mt. Fuji, which is located ~550 km away (Miyamoto *et al.*, 2014). Each elevation gradient encompassed a wide range in climate within a short geographical distance, and different types of forest were present at similar altitudes on both mountains. Specifically, we tested the following hypotheses: (1) fungal species occurring on the two mountains are distributed in the same environmental conditions; (2) compositions of EM fungi and trees are positively related; and (3) host phylogeny relatedness drives EM fungal composition. We then discuss the relative importance of environmental factors, host phylogeny, geographical distance or combination of these factors on EM fungal composition.

Materials and methods

Study sites and sampling

Field sampling was conducted on the south slope of Mt. Ishizuchi, western Japan, in 2012. This area has a temperate climate characterized by warm, wet summers and cool, moderately dry winters. We established three study sites in closed-canopy natural forests along an elevation gradient on the mountain (Table 1). Site I1 (850 m) was an evergreen broadleaf and conifer mixed forest dominated by *Tsuga sieboldii* (37% of the relative basal area), *Abies firma* (22%) and *Quercus salicina* (21%). Site I2 (1450 m) was a deciduous and conifer mixed forest dominated by *Fagus crenata* (49%) and *Abies homolepis* (16%). Site I3 (1850 m) was a subalpine conifer forest that was exclusively dominated by *Abies veitchii* (96%). The composition of tree species along the elevation represented a typical vegetation change in the southwest part of Japan. The highest sampling site (1850 m) was located just below the treeline.

Field sampling was conducted according to Miyamoto *et al.*, 2014. Briefly, 50 rectangular soil

Table 1 Description of study sites on Mt. Ishizuchi

Site	I1	I2	I3
Elevation (m)	850	1450	1850
Coordinates	N33°44' E133°07'	N33°44' E133°09'	N33°46' E133°07'
Temperature (°C) ^a	9.4	6.2	4.6
Precipitation (mm) ^a	2823	3137	2806
Tree density (stems per ha)	2363	2450	963
Tree basal area (m ² per ha)	46.9	39.1	33.2
Number of tree species	34	25	10
Number of host tree species	4	4	3
pH (H ₂ O) ^b	3.5–5.5 (4.1)	3.4–4.7 (3.8)	3.4–4.8 (3.8)
C/N ^b	11.9–36.9 (18.3)	9.0–20.8 (15.6)	6.8–21.2 (17.3)
Litter (cm) ^b	0–5.0 (2.4)	0.5–5.0 (1.9)	0.5–4.0 (1.7)

Abbreviation: C/N, carbon/nitrogen.

^aSite-specific mean annual temperature and mean annual precipitation obtained from the interpolated mesh data (1 × 1 km²; 30-year averages from 1981–2010) provided by The Japan Meteorological Agency, 2014.

^bValues are minimum–maximum with the mean in parentheses.

cores (5 × 5 cm to 10 cm depth) were collected from a 1-ha area at each site. A minimum distance of 5–10 m was maintained between soil-sampling locations to ensure sample independence (Lilleskov *et al.*, 2004; Pickles *et al.*, 2012). A vegetation survey was conducted at every other soil-sampling location. Tree species and diameter at breast height (1.3 m) were recorded for all living trees (>1.3 m tall) within a 5-m radius of the sampling point. Litter depth and geographical coordinates (Garmin 62S; Garmin International, Olathe, KS, USA) were recorded at each sampling point.

Molecular analyses

We followed Miyamoto *et al.*, 2014 for the molecular analyses. Briefly, all roots were collected from each soil sample, and EM root tips were classified by their morphological characteristics. Three healthy EM root tips were sampled from each morphotype of each core for molecular analyses. The cetyltrimethyl ammonium bromide method was used to extract fungal DNA from the root tips. A polymerase chain reaction (PCR) was performed to amplify the internal transcribed spacer (ITS) regions (ITS1, 5.8S and ITS2) of the rDNA using the forward primer ITS1F and reverse primers (i.e., ITS4, LR21 and LR22). Replicates of each morphotype were compared based on restriction fragment length polymorphism (RFLP) patterns using *Hinf*I and *Alu*I restriction enzymes, and unique RFLP types in a morphotype were subjected to direct sequencing (3730xl DNA analyzer; Applied Biosystems, Foster City, CA, USA). PCR products were purified and sequenced using primer ITS1. Primer ITS4 was used for poorly sequenced samples. All sequences were manually inspected using chromatograms. High-quality sequences with >350 bp were trimmed, aligned and clustered into molecular operational taxonomic units (hereafter referred to as 'species') at ≥97% similarity using the alignment software ATGC ver. 7 (Genetyx Corp., Tokyo, Japan). The sequences were compared with those deposited in the International Nucleotide Sequence Database (INSD). Sequences that showed a high degree of similarity with saprophytic and pathogenic fungi or nonfungal taxa were removed from the analyses. *Cenococcum geophilum* (hereafter referred to as 'Cg') was identified primarily based on its unique morphology as in previous studies (Twieg *et al.*, 2007; Murata *et al.*, 2013), while a few Cg operational taxonomic units were detected through molecular analyses. Host trees associated with EM fungi were identified to the genus level by amplifying the plastid *trnL* region of plant DNA using the primer pairs *trnC-trnD* and *trnE-trnF* (Taberlet *et al.*, 1991). RFLP patterns were then compared with those obtained from the leaves of host species recorded in the field. Direct sequencing was additionally applied to those samples with unclear RFLP patterns.

Soil data collection

Soil pH, total carbon (C), and total nitrogen (N) of each sample were measured according to Miyamoto *et al.*, 2014. Soil pH was measured from a 5-g air-dried soil sample in a 1:5 ratio with Milli-Q water (Millipore, Billerica, MA, USA) using a HM-25G glass electrode (DKK-TOA Corp., Tokyo, Japan). Total C and N were measured by dynamic flash combustion with a CN Analyzer (Flash EA 1112; AMCO Inc., Tokyo, Japan).

Statistical analyses

We combined the data from this study with those of a previous study conducted on Mt. Fuji (Miyamoto *et al.*, 2014) and analyzed the combined data to reveal community structures at a regional scale. Fungal species recorded on Mt. Ishizuchi and Mt. Fuji were matched based on a ≥97% sequence similarity. Sites on Mt. Fuji were reassigned as F1 (site 1, elevation 1100 m), F2 (site 2, 1550 m), F3 (site 3, 1900 m) and F4 (site 4, 2250 m). The forest types in the seven sites were defined based on the composition of all tree species (using a single-linkage agglomerative clustering of tree compositions; Supplementary Figure S1). Four forest types were identified: subalpine conifer (*Abies-Tsuga*-dominated; F3, F4, I3), cool temperate mixed (*Abies-Fagus*-dominated; F2, I2), warm temperate deciduous (*Quercus*-dominated; F1) and warm temperate mixed (*Abies-Quercus*-dominated; I1). The estimated richness (Chao 2 nonparametric estimators) was computed using EstimateS (version 8.20; Colwell *et al.*, 2012) with 5000 randomizations without replacement. R version 3.0.3 (R Development Core Team, 2013) was used for statistical analyses, with the significance level set to $P=0.05$ (unless otherwise noted). Singleton species that were recorded in one core across the entire data set were removed from community analyses.

The degree of species overlaps between EM fungal communities was calculated as the number of shared species divided by the total number of species recorded at each pair of sites. Fisher's exact tests were conducted to test whether the fungal species found on both mountains ('mountain-shared species') that were recorded at a particular site occurred randomly at sites on another mountain (e.g., the null hypothesis was that the mountain-shared species that were recorded at F1 on Mt. Fuji were randomly distributed at I1, I2 and I3 on Mt. Ishizuchi). The mountain-shared species were tested to determine whether the frequency of the species was biased toward particular forest types using Fisher's exact test. The Benjamini and Hochberg false discovery rate correction (Verhoeven *et al.*, 2005) was performed to adjust type I error for multiple comparisons using the *fmsb* package of R.

The occurrence of fungal species per site was treated as a sample unit ($n=7$) for community analyses. The relationship between EM fungal and tree species composition similarities was tested

using the Mantel test with Bray–Curtis distance matrices. Spatial autocorrelation among the fungal composition was tested using the Mantel test with fungal composition against the Euclidean distance of a geographical matrix. Significance was tested with 9999 permutations. Separation of EM fungal composition by location (Mt. Fuji vs Mt. Ishizuchi) and forest type was tested using Adonis (permutation-based multivariate analysis of variance) in the *vegan* package of R. Adonis partitions a distance matrix among categorical or continuous variables, and computes the strength and significance of the explanatory variables (Anderson, 2001). The significance was tested with 999 permutations. Community dissimilarities were visualized in two-dimensional space using nonmetric multidimensional scaling (NMDS) with 999 permutations. Bray–Curtis distances were calculated prior to Adonis and NMDS visualization. The effect of individual environmental variables on NMDS ordination was examined using environmental fitting tests in the *envfit* function in *vegan*, and the significance of vectors was tested with 999 permutations. The variables included were climate (mean annual temperature and mean annual precipitation), soil (C/N, pH and litter depth) and geographical distance. Soil variables were log transformed prior to analyses. Geographical distance (latitude and longitude) was transformed to the principal coordinates of neighbor matrices (PCNM) vectors that represented the geographical distances at various spatial scales (Borcard *et al.*, 2004; Dray *et al.*, 2006). PCNM vectors were calculated from the pairwise Euclidean distance of geographical coordinates between study sites.

Furthermore, the EM fungal community per host per site was treated as a sample unit ($n=19$) in analyses to evaluate the effect of host identity. Hosts represented by ≤ 5 cores were excluded from the analysis. The relative effects of climate, soil, host identity and geographical distance were analyzed using variation partitioning in redundancy analyses (the *varpart* function in *vegan*). Forward selection by redundancy analysis was used prior to variation partitioning to identify significant variables associated with EM fungal compositions based on 999 permutations (Borcard *et al.*, 2011). Only significant variables were used to compute the total variance of the EM fungal composition explained by these variables.

Host identity was measured as the phylogenetic distance among the seven host genera, computed according to Tedersoo *et al.*, 2013. Plastid *matK* and *trnL* gene sequences were downloaded from INSD or generated during our molecular host identification (Supplementary Table S1). These regions sufficiently distinguished host genus, whereas they were identical at the species level. Sequences of each region were aligned using MAFFT v.7.147 (<http://mafft.cbrc.jp/alignment/software/>) with the iterative refinement method (L-INS-i algorithm; Katoh and

Toh, 2008). A phylogenetic tree was constructed using *trnL-matK* by applying maximum likelihood algorithms with a general time reversible model and 500 bootstrap replicates using MEGA ver. 6.0 (Tamura *et al.*, 2013). Pairwise patristic distances (pairwise sum of the branch length connecting two terminal taxa) were then calculated using the *ape* package of R. The pairwise patristic distance matrix was converted to phylogenetic eigenvectors using principal coordinate analysis. Principal coordinate analysis vectors are orthogonal and represent phylogenetic distances of host trees. Significant eigenvectors were forward selected prior to analyses.

Results

General descriptions of EM fungi on Mt. Ishizuchi

In total, 1469 root tips were collected for molecular analyses; 1371 tips (93%) were successfully amplified and 1206 tips were identified to species by RFLP and sequencing analyses. Thirteen sequences did not belong to EM fungal taxa. In total, 465 of 514 cumulative morphotypes (90%) were successfully identified to species. We recorded 200 species on Mt. Ishizuchi, including 110 singletons (55.0%) and 39 doubletons (19.5%; Supplementary Table S2). The mean length of our sequences was 584 bp and 88% of the sequences (134 of 152 newly detected species) were longer than 500 bp. The most frequently observed lineages included */russula-lactarius* (49 species) and */tomentella-thelephora* (38 species). The most frequently observed species was *Cg* (76 of 150 soil cores; 50.7%), followed by *Clavulina castaneipes* (25 cores; 16.7%) and *Russula granulata* (19 cores; 12.7%). Seven species occurred at all sites. The observed species richness was 93 at site I1, 98 at site I2 and 55 at site I3. The rarefaction curves of Chao 2 estimated richness, reached an asymptote at site I1 but did not become asymptotic at sites I2 and I3 (Supplementary Figure S2). The Chao 2 estimated richness on Mt. Ishizuchi was 355 ± 40.6 . Exceptionally abundant *Cg*, which likely contained some cryptic species (Douhan and Rizzo, 2005), was removed from further analyses.

Species overlaps among communities

In total, 453 EM fungal species (excluding *Cg*) were recorded from the two mountains, including 225 singletons (49.7%) and 89 doubletons (19.6%). Pearson's linear correlation showed that EM fungal richness (Chao 2) was positively correlated with belowground host genus richness ($r=0.83$; $t_5=3.34$; $P=0.021$). Singleton species that were found in one core across the entire data set were removed to improve community analyses, leaving 228 species for further analyses.

A total of 47 species were shared between the mountains (Figure 1 and Table 2). Sites F2 and I2 (cool temperate mixed forest pairs) shared 20

species (18.4% of the total number of species recorded at these sites). Sites F3 and I3 (*Abies*-dominated subalpine conifer forest pairs) shared 14 species (14.1%). These shared fungi occurred across multiple host genera (Supplementary Figure S3). Fisher's exact test with false discovery rate correction showed that the mountain-shared species recorded in the subalpine conifer forests on Mt. Fuji (sites F3 and F4) occurred at significantly higher frequencies in the subalpine conifer forest (site I3) than in the other forest types (site I1 or I2) on Mt. Ishizuchi ($P < 0.043$). Similarly, the mountain-shared species recorded at site I3 on Mt. Ishizuchi occurred at a significantly higher frequency at site F3 on Mt. Fuji ($P = 0.019$). The mountain-shared species in the mixed forest on Mt. Ishizuchi (site I2) also occurred at higher frequency in the mixed forest on Mt. Fuji (site F2) at $P = 0.085$. Approximately, 66% (31 of 47) of mountain-shared species occurred in particular forest types, and Fisher's exact test showed significantly biased frequency of occurrence in some fungal species (Table 2).

Composition and underlying mechanisms

A Mantel test showed that the similarities in EM fungal composition were significantly correlated with those of forest tree composition ($r_M = 0.47$, $P = 0.042$), but not with geographical distance ($P = 0.146$) at the regional scale. A partial Mantel test revealed that geographical distance was significant when the effect of tree composition was

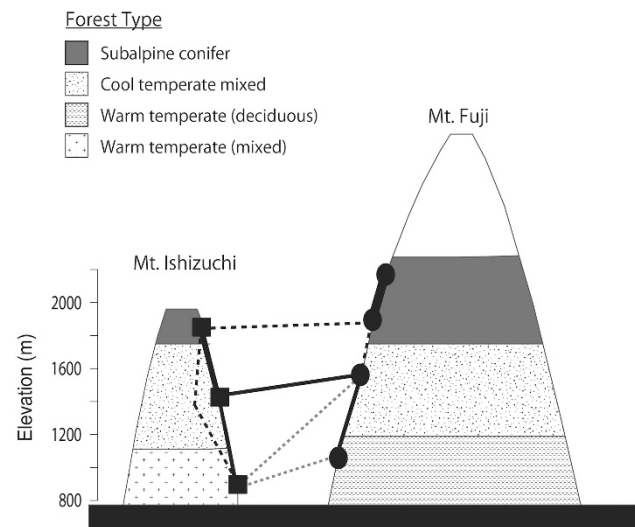


Figure 1 The degree of species overlaps among study sites. Circles indicate the study sites on Mt. Fuji, and squares indicate those on Mt. Ishizuchi along the elevation gradients. The thickness of the connecting line represents the pairwise proportion of shared fungal species between sites (i.e., the number of shared species divided by the total number of species at paired sites). Solid lines indicate a proportion of $>15\%$, dashed black lines indicate a proportion of 13–15% and dotted gray lines indicate a proportion of 10–13%. The proportion $<10\%$ is not shown for clarity.

excluded ($r_M = 0.45$, $P = 0.033$). Tree composition remained significant when the distance effect was excluded ($r_M = 0.55$, $P = 0.024$). An Adonis test showed that the larger variance was explained by forest type than location (Figure 2a and Table 3).

Furthermore, the EM fungal composition on *Abies*, which occurred at six of the seven studied sites, was analyzed separately to remove host effects. The results were similar to those of the entire community (Supplementary Figure S4). An Adonis test showed that forest type explained the larger variance in fungal composition ($F_{2,2} = 1.75$, $R^2 = 0.47$; $P = 0.02$) than location ($F_{1,2} = 1.96$, $R^2 = 0.26$; $P = 0.03$).

EM fungal communities on individual hosts per site were separated by forest type and environmental factors (Figure 2b). Variation partitioning showed that climate (temperature and precipitation) alone explained the largest variance ($R^2_{\text{adj}} = 0.13$) in the EM fungal composition, followed by soil (C/N and pH; $R^2_{\text{adj}} = 0.05$) and geographical distance ($R^2_{\text{adj}} = 0.04$; Figure 3). No significant effect of host identity alone was detected in variation partitioning. Temperature, precipitation, C/N and host identity were significantly correlated with NMDS vectors based on the environmental fitting test (Table 4).

Discussion

Differentiating the effects of host, environment and geographical distance on EM fungal communities is difficult when these factors covary at large spatial scales (Toljander *et al.*, 2006; Tedersoo *et al.*, 2012; Miyamoto *et al.*, 2014). We applied relatively intensive and consistent sampling methods and recorded 454 EM fungal species from 330 of 350 soil samples along two elevation gradients. This is among the highest richness reported from one region, which enabled us to show the degree of species overlaps between similar habitats on geographically distant mountains. We demonstrated that $\sim 20\%$ of the EM fungal species occurred on the two mountains and they tended to inhabit similar forest types (Figure 1 and Table 2). The species overlaps were particularly notable between the cool temperate mixed forests on both mountains, where the highest proportion (18.4%) of EM fungi overlapped despite the geographical distance of ~ 550 km. In contrast, only one and five EM fungal species were shared between nonadjacent sites within Mt. Fuji (F2 and F4) or Mt. Ishizuchi (I1 and I3), respectively, despite the close geographical distances of <10 km, indicating a weak effect of geographical distance *per se* (Miyamoto *et al.*, 2014). These results suggest that some EM fungi are widely distributed across the region and their presence is constrained by contemporary environmental factors. To support this, many mountain-shared species (31 of 47 species or 66.0%) displayed niche preferences to particular forest types (Table 2). Those EM

Table 2 List of mountain-shared species

Forest type	Species	Fuji				Ishizuchi			P-value
		F1	F2	F3	F4	I1	I2	I3	
Subalpine conifer	Clavulinaceae sp.5			●	●			■	*
	<i>Clavulina castaneipes</i>			●	●		■	■	*
	<i>Cortinarius scaurus</i> sp.1			●	●			■	*
	<i>Sebacina</i> sp.3			●	●			■	
	<i>Lactarius imperceptus</i>			●				■	*
	<i>Amanita</i> sp.1			●				■	
	<i>Piloderma</i> sp.3			●				■	
	<i>Sebacina</i> sp.29			●				■	
Cool temperate mixed	<i>Genea hispidula</i>		●				■		
	<i>Russula</i> sp.2		●				■		
	<i>Russula</i> sp.12		●				■		
	<i>Tomentella</i> sp.6		●				■		
	<i>Tomentella</i> sp.12		●				■		
Cool temperate mixed and warm temperate (deciduous and mixed)	<i>Tomentella</i> sp.50		●				■		
	<i>Cortinarius</i> sp.17		●			□	■		
	<i>Tuber</i> sp.2		●			□	■		
	<i>Inocybe</i> sp.15		●			□			
	<i>Tomentella</i> sp.11		●			□			
	<i>Tomentella stiposa</i> sp.1		●			□			
	<i>Entoloma</i> sp.1	○	●			□	■		*
	<i>Laccaria</i> sp.1	○	●			□	■		*
	<i>Sebacina</i> sp.4	○	●			□	■		*
	<i>Xerocomus</i> sp.2	○	●			□	■	■	*
	<i>Cortinarius anomalus</i>	○				□	■		*
	<i>Cortinarius</i> sp.14	○				□	■		
	<i>Cortinarius umbrinolens</i>	○				□	■		
	<i>Lactarius quietus</i>	○				□	■		
	Warm temperate (deciduous and mixed)	<i>Entoloma</i> sp.3	○				□		
<i>Tomentella</i> sp.41		○				□			
<i>Tomentella</i> sp.48		○				□			
<i>Xerocomus</i> sp.3		○				□			
Various forests	Clavulinaceae sp.1			●	●		■		
	<i>Inocybe</i> sp.11			●	●		■	■	
	<i>Laccaria</i> cf. <i>laccata</i> sp.2		●				■	■	
	<i>Russula peckii</i>		●	●			■	■	
	<i>Russula turci</i>				●	□		■	
	<i>Sebacina</i> sp.1		●	●	●		■	■	
	<i>Sebacina</i> sp.2		●	●	●	□		■	
	<i>Sebacina</i> sp.20		●	●	●		■	■	
	Thelephoraceae sp.2		●			□	■	■	
	<i>Tomentella</i> sp.5		●	●		□	■		
	<i>Tomentella</i> sp.7		●	●		□	■		
	<i>Tomentella</i> sp.8		●	●		□	■	■	
	<i>Tomentella</i> sp.18		●	●		□	■		
	<i>Tomentella</i> sp.21		●	●	●	□			
	<i>Tomentella</i> sp.31			●		□		■	
	<i>Tomentella subililacina</i>	○		●			■	■	

Symbols indicate the presence of a species. Circles represent sites on Mt. Fuji and squares represent sites on Mt. Ishizuchi. Forest types are subalpine conifer forest (black), cool temperate mixed forest (gray) and warm temperate mixed and deciduous forest (white). Asterisks indicate species whose frequency is significantly biased by forest type based on Fisher's exact tests after false discovery rate (FDR) correction.

fungal species restricted to particular habitats are likely more sensitive to environmental change than habitat generalists (Carignan and Villard, 2002), such as *Tomentella subililacina* and *Laccaria laccata*, which were recorded across wide environmental ranges.

Variation partitioning further indicated that environmental factors (i.e., climate and soil) had a stronger effect on the EM fungal composition than geographical distance or host identity (Figure 3). Previous studies have reported the importance of

climate (Bahram *et al.*, 2012; Tedersoo *et al.*, 2012) and soils (Cox *et al.*, 2010; Jarvis *et al.*, 2013) on EM fungal composition at regional to global scales. However, the potential effects of climate remain unclear because climate covaries with spatial distance in these studies. Alternatively, Põlme *et al.*, 2013 and Roy *et al.*, 2013 examined a single host genus (i.e., *Alnus*) and identified significant spatial structures in the EM fungal compositions, but environmental factors had a stronger influence on the community structures. *Alnus* is associated with

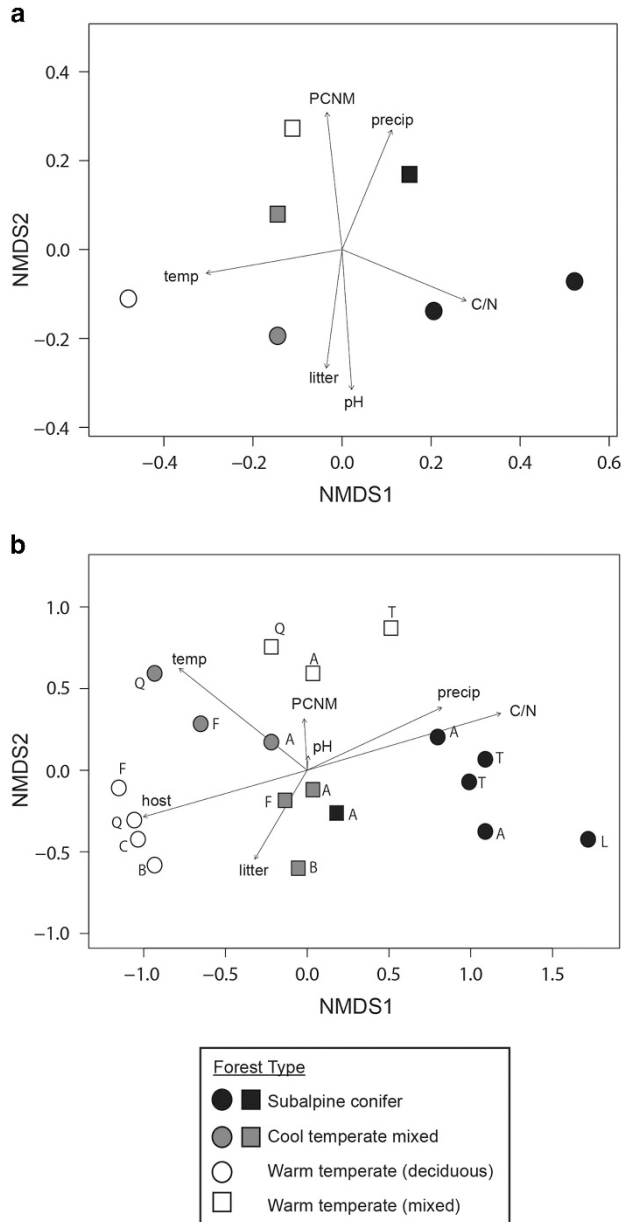


Figure 2 Nonmetric multidimensional scaling (NMDS) graphs of the ectomycorrhizal fungal communities. Environmental variables are fitted to the NMDS ordination. The abbreviations are temp (temperature), precip (precipitation), host (host phylogenetic eigenvector) and PCNM (spatial principal coordinates of neighbor matrices eigenvector). The significance of the vectors is shown in Table 4. Circles and squares indicate sites on Mt. Fuji and Mt. Ishizuchi, respectively. **(a)** Seven sites (stress = 0.08) and **(b)** 19 host communities (stress = 0.10). Host genera are *Fagus* (F), *Quercus* (Q), *Betula* (B), *Carpinus* (C), *Abies* (A), *Tsuga* (T) and *Larix* (L). Three-dimensional NMDS graphs were included in Supplementary Figure S5.

N-fixing bacteria and usually form unique EM fungal communities with low diversity (Kennedy and Hill, 2010). Our results indicated that a stronger effect of environmental factors than geographical distance could be applicable to the more species-rich EM fungal communities that are associated with many typical EM host genera. This was further

Table 3 Effect of forest type and location in separating the ectomycorrhizal fungal composition (Bray–Curtis distance) as revealed by the Adonis test

	DF	SS	MS	F-statistic	R ²	P-value
Forest type	3	1.41	0.47	2.22	0.59	0.003
Location	1	0.56	0.56	2.66	0.24	0.006
Residuals	2	0.42	0.42			
Total	6	2.4	2.39			

Abbreviations: DF, degree of freedom; MS, mean square; SS, sum of squares.

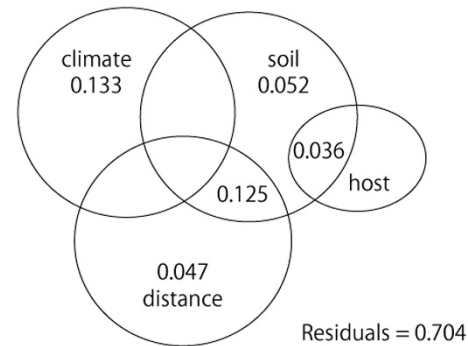


Figure 3 Individual and interaction effects of putative factors explaining 19 ectomycorrhizal fungal community compositions as revealed by variation partitioning in redundancy analyses. Values are adjusted R² (<0.01 are not shown). Factors include climate (mean annual temperature and mean annual precipitation), soil (C/N and pH), distance (spatial principal coordinates of neighbor matrices eigenvector) and host (host phylogenetic eigenvector).

Table 4 Effects of environmental variables on fungal composition fitted to the nonmetric multidimensional scaling ordination

Variables	R ²	P-value
<i>(a) 7 Sites</i>		
Annual temperature	0.795	0.043
Annual precipitation	0.703	0.057
PCNM vector	0.799	0.067
pH	0.828	0.055
C/N	0.754	0.069
Litter	0.601	0.144
<i>(b) 19 Host communities</i>		
Annual temperature	0.579	0.003
Annual precipitation	0.476	0.006
PCNM vector	0.058	0.640
pH	0.005	0.973
C/N	0.903	0.001
Litter	0.141	0.318
Host	0.628	0.002

Abbreviations: C/N, carbon/nitrogen; PCNM, principal coordinates of neighbor matrices. P-values in bold indicate the significance level <0.05.

confirmed in our analysis using a single host genus, *Abies* (Supplementary Figure S4).

The host family has been reported to influence EM fungal composition at the global scale

(Tedersoo *et al.*, 2012). However, whether EM fungal composition is driven by host identity *per se* or other confounding environmental factors is unknown because climate and geological history inherently affect host distributions at the global scale. In contrast, we clearly showed that host identity had a minor role in explaining EM fungal composition at the regional scale. First, different host genera coexisting at the same site tended to harbor similar EM fungi (Figure 2b). Second, although we detected significant host effects in the environmental fitting test (Table 4), the hosts alone were insignificant when environmental factors were excluded in the variation partitioning analysis (Figure 3). Finally, mountain-shared species tended to occur in similar forests on different mountains, but not strictly on the same hosts (Supplementary Figure S3). Thus, any host effect at the global scale would be a consequence of climate and geological history, and not derived from phylogenetic constraints between the symbiotic partners.

The lack of large host effects at the regional scale may be related to our studied system that was dominated by generalist EM partners. Our forests were composed of typical EM host trees (e.g., *Fagus*, *Quercus* and *Abies*) that harbored generalist EM fungal groups such as Russulaceae and Thelephoraceae (Smith *et al.*, 2009). In contrast, host effects would be more prominent for specialist symbionts, such as *Alnus–Alpova* and *Pinus–Rhizopogon* (Molina *et al.*, 1992; Kennedy and Hill, 2010). Indeed, we detected some host-specific fungal genera such as *Suillus* and *Tylospora*, which occurred only on conifer hosts. But their occurrence appeared to be too low to separate the communities among hosts. Moreover, the significance of host effects would depend on spatial scales and host composition within a stand. Our stand-scale analyses detected significant host effects at some sites, especially in gymnosperm–angiosperm mixed forests (Miyamoto, unpublished data; Supplementary Table S3), as in many previous studies (Ishida *et al.*, 2007; Tedersoo *et al.*, 2008; Smith *et al.*, 2009). However, most EM fungi are assumed to be host generalists (Bruns *et al.*, 2002), which is supported by numerous studies (Kennedy *et al.*, 2003; Roy *et al.*, 2008; Trocha *et al.*, 2012; Bahram *et al.*, 2013b). Therefore, although the host may influence the EM fungal community within a narrow range of climatic conditions at stand scales, the effect could be masked by more dominant determinants (e.g., climate) at larger spatial scales. Plant communities are well known to be primarily determined by climate, and to some extent by soil conditions, within a biogeographic region (Kira, 1991; Morin *et al.*, 2007). This is largely because climatic conditions (e.g., temperature and precipitation) and nutrients are the major growth-limiting resources for autotrophic plants. Both this work and previous studies (Bahram *et al.*, 2012) have indicated that temperature and precipitation could restrict EM fungal distributions.

We found a positive correlation between the compositional similarities of EM fungi and trees in the Mantel test, which agreed with previous studies on plant and fungal communities at regional scales (Edwards and Zak, 2010; Peay *et al.*, 2013). However, these correlations do not necessarily represent the direct links between plants and microorganisms, and the empirical evidence of direct links appears to be inconclusive (Waldrop *et al.*, 2006; Bryant *et al.*, 2008; Queloz *et al.*, 2011; van der Putten *et al.*, 2013). In fact, hosts *per se* had little predictive power with regard to the EM fungal composition in our analysis. Thus, EM fungi and trees may synchronously, but independently, respond to the same environmental factors, particularly climatic conditions. Global climate change is becoming a serious issue in forest ecosystems (Bonan, 2008; Allen *et al.*, 2010). Given the short generation turnover (Wadud *et al.*, 2014), EM fungi may exhibit a higher adaptive capacity to environmental change than trees. Our data provide important insights that host plants and mutualistic fungi may respond to climate change idiosyncratically, potentially altering carbon and nutrient cycles in relation to the plant–fungus associations.

We investigated seven sites using the intensive sampling approach combined with molecular identifications of both EM fungi and hosts. Although there is a trade-off between the intensity of sampling and number of sampling sites in microbial ecological studies (Miyamoto *et al.*, 2014), the number of sites in this study was sufficient to demonstrate the strong climatic effects on EM fungal community structures while excluding the effects of geographical distance and hosts at the regional scale. Extending the intensive sampling approach to other biogeographic regions in future studies would help to clarify the relative importance of biogeographic history and climate on distributions and community structures of EM fungi.

Conflict of Interest

The authors declare no conflict of interest.

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