

ORIGINAL ARTICLE

Thermodynamic constraints on methanogenic crude oil biodegradation

Jan Dolfig¹, Stephen R Larter² and Ian M Head¹

¹School of Civil Engineering and Geosciences, Newcastle University, Newcastle upon Tyne, UK and

²Petroleum Reservoir Group, Department of Geoscience and Alberta Ingenuity Center for In Situ Energy, University of Calgary, Calgary, Alberta, Canada

Methanogenic degradation of crude oil hydrocarbons is an important process in subsurface petroleum reservoirs and anoxic environments contaminated with petroleum. There are several possible routes whereby hydrocarbons may be converted to methane: (i) complete oxidation of alkanes to H₂ and CO₂, linked to methanogenesis from CO₂ reduction; (ii) oxidation of alkanes to acetate and H₂, linked to acetoclastic methanogenesis and CO₂ reduction; (iii) oxidation of alkanes to acetate and H₂, linked to syntrophic acetate oxidation and methanogenesis from CO₂ reduction; (iv) oxidation of alkanes to acetate alone, linked to acetoclastic methanogenesis and (v) oxidation of alkanes to acetate alone, linked to syntrophic acetate oxidation and methanogenesis from CO₂ reduction. We have developed the concept of a 'window of opportunity' to evaluate the range of conditions under which each route is thermodynamically feasible. On this basis the largest window of opportunity is presented by the oxidation of alkanes to acetate alone, linked to acetoclastic methanogenesis. This contradicts field-based evidence that indicates that in petroleum rich environments acetoclastic methanogenesis is inhibited and that methanogenic CO₂ reduction is the predominant methanogenic process. Our analysis demonstrates that under those biological constraints oxidation of alkanes to acetate and H₂, linked to syntrophic acetate oxidation and methanogenesis from CO₂ reduction offers a greater window of opportunity than complete oxidation of alkanes to H₂ and CO₂, linked to methanogenic CO₂ reduction, and hence is the process most likely to occur.

The ISME Journal (2008) 2, 442–452; doi:10.1038/ismej.2007.111; published online 13 December 2007

Subject Category: geomicrobiology and microbial contributions to geochemical cycles

Keywords: anaerobic oil degradation; methanogenesis; syntrophy; window of opportunity; syntrophic acetate oxidation; hydrocarbons

Introduction

The largest deposits of petroleum on Earth are not, as conventionally assumed, in the Middle East. The vast Saudi Arabian and Kuwaiti oilfields of Ghawar (2.6×10^{11} barrels (bbl) in place) and Burgan (7.0×10^{10} bbl in place) are dwarfed by the trillion bbl deposits of western Canada (Athabasca tar sands; 1.7×10^{12} bbl) and Venezuela (Orinoco heavy oil belt; 1.2×10^{12} bbl). These so-called super-giant heavy oil fields are the result of biodegradation of the lighter, more readily produced and valuable oil fractions over geological time. Biodegraded oil fields are more difficult to produce and the oils more difficult to refine than oil from conventional fields and are thus less economically attractive. On the

basis of known mechanisms of hydrocarbon degradation, conventional wisdom among petroleum geologists has for some time, been that biodegradation in oilfields was driven by oxygen delivered to petroleum reservoirs in meteoric water. This paradigm has been questioned in light of the discovery of a range of bacteria capable of coupling the oxidation of aliphatic or aromatic hydrocarbons to the reduction of nitrate, iron and sulphate (Widdel and Rabus, 2001) and microbial consortia capable of linking aliphatic hydrocarbon oxidation to methane generation (Zengler *et al.*, 1999; Anderson and Lovley, 2000; Townsend *et al.*, 2003).

Evidence is emerging to support the notion that in-reservoir petroleum biodegradation is caused by anaerobic hydrocarbon degrading bacteria. Reduced naphthoic acids, metabolites characteristic of anaerobic hydrocarbon degradation have been detected in biodegraded petroleum reservoirs, but not in non-degraded reservoirs (Aitken *et al.*, 2004). Geochemical and isotopic evidence also suggests that in many cases the end product of hydrocarbon degradation in petroleum reservoirs is methane (Scott *et al.*, 1994;

Correspondence: J Dolfig, School of Civil Engineering and Geosciences, Newcastle University, Cassie Building, Newcastle upon Tyne, NE1 7RU, UK.

E-mail: jan.dolfig@ncl.ac.uk

Received 23 August 2007; revised 8 November 2007; accepted 13 November 2007; published online 13 December 2007

Larter *et al.*, 1999; Sweeney and Taylor, 1999; Pallasser, 2000; Boreham *et al.*, 2001; Masterson *et al.*, 2001). Compositional gradients in oil columns towards the underlying water leg in biodegraded petroleum reservoirs suggests that the oil water transition zone is the primary site of biodegradation in petroleum reservoirs (Head *et al.*, 2003; Larter *et al.*, 2003).

On the basis of these and other data, a new conceptual model of in-reservoir petroleum biodegradation has been developed (Head *et al.*, 2003). In this model, anaerobic degradation of petroleum occurs most actively at the oil water transition zone. Electron donor, mainly hydrocarbons, is delivered to the oil water transition zone by diffusion from the oil column, with inorganic nutrients such as ammonium ions provided from the water leg (Head *et al.*, 2003; Manning and Hutcheon, 2004). This is consistent with reports from other deep subsurface environments that microbial activity is stimulated at geochemical interfaces (Parkes *et al.*, 2005). When the water leg contains low levels of sulphate, hydrocarbon degradation is driven by methanogenesis; indeed, many biodegraded petroleum reservoirs contain isotopically light methane indicative of a mixed secondary biogenic and thermogenic source (Scott *et al.*, 1994; Larter *et al.*, 1999; Sweeney and Taylor, 1999; Pallasser, 2000; Boreham *et al.*, 2001; Masterson *et al.*, 2001; Head *et al.*, 2003).

The significance of methanogenic crude oil degradation in petroleum reservoirs goes beyond its potential role in the biodegradation of petroleum reservoirs; it may ultimately be crucial for processes that can enhance the recovery of residual oil. Typically, over 60% of the oil in place in a petroleum reservoir remains unextractable following standard production procedures and the possibility that methanogenic degradation of this residual oil can re-pressurize a petroleum reservoir, has some potential for enhancing oil recovery. Furthermore, the volumetrics of gas recovery are far better than for oil (typically 70% of gas in place can be recovered) and methanogenic conversion of non-recoverable residual hydrocarbons to recoverable gas may be an economically viable way of extending the operational life of petroleum reservoirs (Parkes, 1999; Larter *et al.*, 1999; Head *et al.*, 2003). In addition, methanogenic hydrocarbon degradation may be a significant process in the attenuation of contaminated anoxic sediments and aquifers (Weiner and Lovley, 1998; Anderson and Lovley, 2000; Bekins *et al.*, 2005).

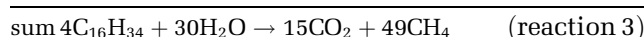
Because of the potential importance of methanogenic crude oil biodegradation and our limited knowledge of the organisms and mechanisms involved, it is important that we learn more about what governs the microbial conversion of oil to methane.

Quantitatively, the most important component of crude oil is the saturated hydrocarbon fraction and little is known about the methanogenic degradation of long-chain aliphatic hydrocarbons; only three

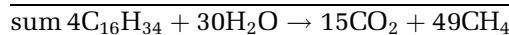
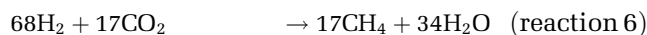
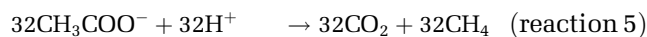
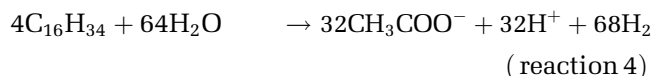
reports in the literature provide strong evidence of methanogenic degradation of aliphatic hydrocarbons or crude oil. Zengler *et al.* (1999) report degradation of pure hexadecane by an enrichment culture; Anderson and Lovley (2000) documented rapid mineralization of ¹⁴C-labelled hexadecane in sediments from a crude oil-contaminated aquifer; and Townsend *et al.* (2003) observed methanogenic transformation of crude oil in sediments from a gas condensate contaminated aquifer.

In this paper, we evaluate the thermodynamics of five possible routes of methanogenic hydrocarbon degradation, viz (with hexadecane as example):

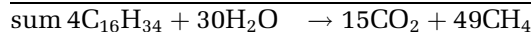
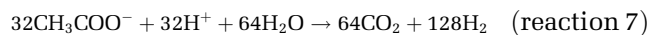
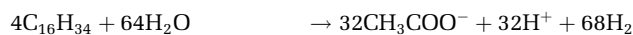
(i) complete oxidation of alkanes to H₂ and CO₂, linked to methanogenesis from CO₂ reduction:



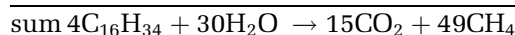
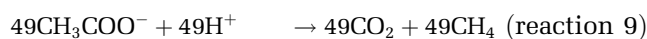
(ii) oxidation of alkanes to acetate and H₂, linked to acetoclastic methanogenesis and CO₂ reduction:



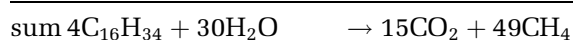
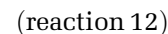
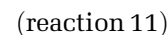
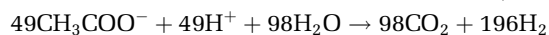
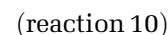
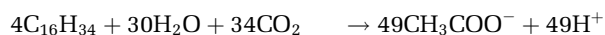
(iii) oxidation of alkanes to acetate and H₂, linked to syntrophic acetate oxidation and methanogenesis from CO₂ reduction:



(iv) oxidation of alkanes to acetate alone, linked to acetoclastic methanogenesis:



and (v) oxidation of alkanes to acetate alone, linked to syntrophic acetate oxidation and methanogenesis from CO₂ reduction:



The effects of temperature, pH, acetate and H₂ concentration on each of these processes is determined and conditions under which each process is likely to be most favourable are identified and related to conditions typical of petroleum reservoirs. We consider only the case of alkanes with even chain length. It is likely that similar patterns will be observed with odd chain alkanes, which will generate propionate in addition to acetate and H₂. This would require incorporation of syntrophic propionate oxidation as an intermediate reaction. Any effect of propionate will decrease with increasing alkane chain length.

Methods

Gibbs free energy calculations were made after Thauer *et al.* (1977) and Amend and Shock (2001).

Temperature corrections for ΔG° were made with the Gibbs–Helmholtz equation according to:

$$\Delta G_{\text{Tact}}^\circ = \Delta G_{\text{Tref}}^\circ \cdot (T_{\text{act}}/T_{\text{ref}}) + \Delta H_{\text{Tref}}^\circ \cdot (T_{\text{ref}} - T_{\text{act}})/T_{\text{ref}}$$

with T in K; T_{ref} = 298.15 K.

Gibbs free energies and enthalpies of formation data for alkanes in the liquid state were taken from (Helgeson *et al.*, 1998). For all other compounds the data were taken from Hanselmann (Hanselmann, 1991), with acetate in the aqueous phase and methane, hydrogen and carbon dioxide in the gaseous phase at partial pressures of 1 atm. Calculations were made for neutrality rather than for pH=7, with neutrality defined as the pH where activities of H⁺ and OH⁻ are equal. The pH representing neutrality varies with temperature (Harned and Owen, 1943).

Sample calculations

Table 1 lists change in Gibbs free energy values for various reactions of importance to anaerobic hexadecane degradation. The values shown are $\Delta G^{o'}$.

Table 1 Stoichiometry and change in Gibbs free energy values for reactions potentially involved in methanogenic hexadecane degradation

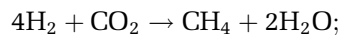
Substrates	Products	$\Delta G^{o'}$	
		kJ/reaction	kJ/mol
<i>Hexadecane degradation</i>			
4C ₁₆ H ₃₄ +128H ₂ O	64CO ₂ +196H ₂	4922.1	1230.5 ^a
4C ₁₆ H ₃₄ +64H ₂ O	32CH ₃ COO ⁻ +32H ⁺ +68H ₂	1883.1	470.8 ^a
4C ₁₆ H ₃₄ +30H ₂ O+34CO ₂	49CH ₃ COO ⁻ +49H ⁺	268.6	67.2 ^a
4C ₁₆ H ₃₄ +30H ₂ O	15CO ₂ +49CH ₄	-1487.1	-371.8 ^a
<i>Conversion of potential intermediates</i>			
CH ₃ COO ⁻ +H ⁺ +2H ₂ O	2CO ₂ +4H ₂	94.9	94.9 ^b
CH ₃ COO ⁻ +H ⁺	CO ₂ +CH ₄	-35.8	-35.8 ^c
4H ₂ +CO ₂	CH ₄ +2H ₂ O	-130.7	-130.7 ^c

^aPer mol hexadecane.

^bPer mol acetate.

^cPer mol methane.

From these data the threshold concentrations of products and reactants, which result in $\Delta G' < 0$ were calculated as follows. The example given is for the threshold H₂ concentration for hydrogenotrophic methanogenesis.



$$\Delta G^{o'} = -130.7 \text{ kJ/molCH}_4$$

Hence (for example Thauer *et al.*, 1977):
 $\Delta G' = -130.7 + RT \ln([\text{CH}_4]/[\text{CO}_2] \cdot [\text{H}_2]^4)$

(note that in biological systems $\ln[\text{H}_2\text{O}]$ is assumed to be 0)

Therefore, under otherwise standard conditions:
 $\Delta G' = -130.7 - 5.71 \log[\text{H}_2]^4$ (where 5.71 logx equals R.T_{298.15} ln x).

Since the threshold value is the value where $\Delta G' = 0$ it follows that

$$-130.7 - 5.71 \log[\text{H}_2]^4 = 0$$

Thus $4 \log[\text{H}_2] = 130.7 / -5.71$, and hence

$$[\text{H}_2] = 10^{(-130.7/22.84)} = 10^{-5.72}$$

Thus $[\text{H}_2]_{\text{crit}} = 1.89 \times 10^{-6}$ atm.

Results

We have examined the effect of alkane chain length on the free energy yield of methanogenic alkane degradation. Hexadecane was chosen as an exemplar to determine the effect of temperature, H₂ concentration, acetate concentration and pH on the thermodynamics of alkane degradation via routes (i) to (v) outlined above. By determining threshold conditions at which the different component reactions for each route become exergonic we have defined 'windows of opportunity' under which different individual reactions can be linked, and thus what conditions are permissive for each of the five routes. The windows of opportunity determined, are discussed in the context of

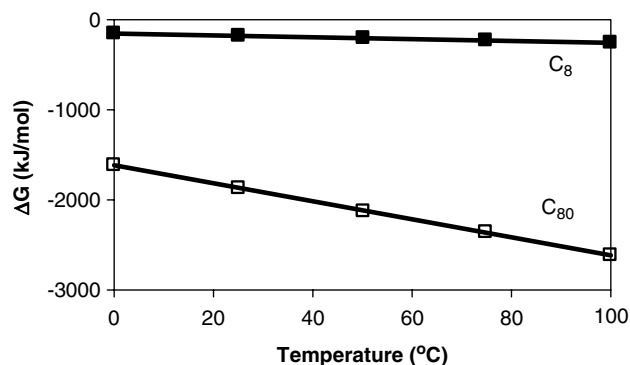


Figure 1 Effect of temperature on change in Gibbs free energy for methanogenesis from alkanes; closed symbols C₈, open symbols C₈₀.

empirical data on crude oil degradation in petroleum reservoirs and used to assess the relative importance of thermodynamics relative to biological factors in determining the routes of methanogenic crude oil degradation, observed in nature.

Effect of chain length on the thermodynamics of anaerobic alkane degradation

The effect of chain length on the free energy yield of methanogenic degradation of alkanes was investigated to evaluate if inferences made using a single model alkane would be generally applicable across all alkanes.

Thermodynamic calculations for all alkanes in the range C₈–C₈₀, demonstrated that conversion to methane was exergonic. ΔG° varied between -185 and -1870 kJ per mol for C₈ to C₈₀ alkanes respectively (Figure 1). In all cases, the free energy yield increased with increasing temperature and the increase in energy yield per degree Kelvin was greater for longer chain hydrocarbons (-0.93 kJ mol⁻¹ K⁻¹ for C₈ and -10.0 kJ mol⁻¹ K⁻¹ for C₈₀). Similar trends were observed for complete oxidation of alkanes to H₂ and CO₂ and incomplete oxidation of alkanes to H₂ and acetate (reactions 1 and 4). In contrast, oxidation of alkanes to acetate alone (reaction 8) became less energetically favourable with increasing temperature, especially for longer chain alkanes ($+0.73$ kJ mol⁻¹ K⁻¹ for C₈ and $+6.0$ kJ mol⁻¹ K⁻¹ for C₈₀; see Supplementary Information for data). When normalized for the number of carbon atoms, there is a slightly greater energy yield per carbon from longer chain length alkanes (for C₈ $\Delta G^\circ = -23.1$ kJ per mol C, for C₈₀ $\Delta G^\circ = -23.4$ kJ per mol C).

These results relate to standard conditions, where the initial oxidation reactions (reactions 1, 4 and 8) are all endergonic. When the effect of H₂ on the complete oxidation pathway is taken into account, it is clear that H₂ concentration has a more marked effect on long chain alkanes than short chain alkanes (Figure 2). However, it is interesting to note that the

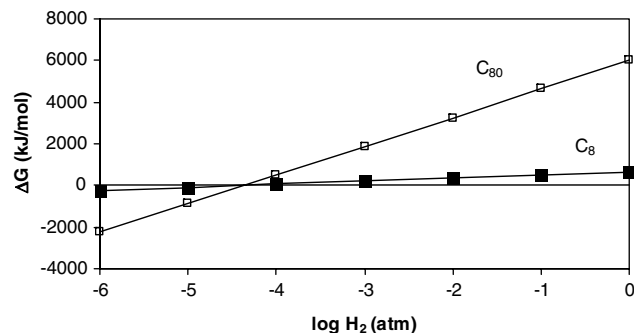


Figure 2 Effect of hydrogen partial pressure on the change in Gibbs free energy for oxidation of alkanes to H₂ plus CO₂; closed symbols C₈, open symbols C₈₀.

threshold H₂ concentration at which alkane oxidation to H₂ and CO₂ becomes energetically favourable under otherwise standard conditions, increases merely from 3.7×10^{-5} atm for *n*-octane (C₈) to 4.3×10^{-5} atm for *n*-octacontane (C₈₀), that is, is essentially independent of chain length (Figure 2).

Overall the same trends are observed, independent of alkane chain length and for simplicity all subsequent discussion is restricted to analysis of hexadecane oxidation, which we have used as an example to illustrate the effect of hydrogen concentration, acetate concentration, temperature and pH on different pathways of methanogenic alkane degradation.

Complete oxidation of hexadecane to H₂ and CO₂ linked to methanogenic CO₂ reduction

While hexadecane oxidation to H₂ and CO₂ becomes more thermodynamically favourable with increasing temperature, methanogenesis by CO₂ reduction becomes less thermodynamically favourable with increasing temperature (Figure 3). However, because the energy yield from hexadecane oxidation increases more steeply than the reduction in energy yield for the methanogenic reaction, the net result is an increase in the overall energy yield from methanogenic hexadecane degradation with increasing temperature (Figure 3).

When the H₂ concentration is taken into account, the energy yield from complete oxidation of hexadecane decreases with increasing H₂ concentration (Figure 4) whereas methanogenic CO₂ reduction becomes more exergonic with increasing H₂ concentration. Provided the overall reaction is exergonic, there is a 'window of opportunity' defined by the H₂ concentrations where both processes are exergonic (Figure 4). It is particularly interesting to note that this window of opportunity increases with increasing temperature and thus this pathway of methanogenic hexadecane degradation becomes less constrained with increasing temperature (that is the window of opportunity becomes larger. Note that the y axis is on a log scale (Figure 5)).

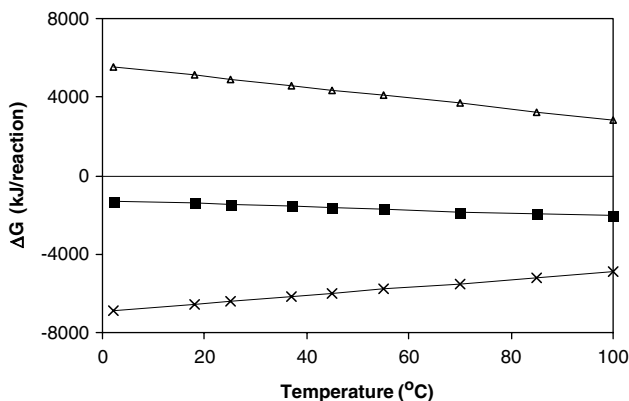


Figure 3 Effect of temperature on the change in Gibbs free energy for complete oxidation of hexadecane to H_2 and CO_2 (open symbols), for stoichiometric methanogenesis of the hydrogen produced (+) and for the sum of the aforementioned reactions, that is, methanogenic degradation of hexadecane (closed symbols). Reactions considered: $4C_{16}H_{34} + 128H_2O \rightarrow 196H_2 + 64CO_2$; $196H_2 + 49CO_2 \rightarrow 49CH_4 + 98H_2O$; $4C_{16}H_{34} + 30H_2O \rightarrow 15CO_2 + 49CH_4$.

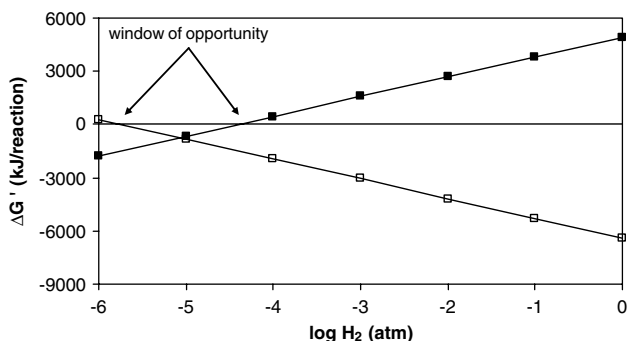


Figure 4 Effect of hydrogen partial pressure on the change in Gibbs free energy for oxidation of hexadecane to H_2 and CO_2 (open symbols) and for stoichiometric methanogenesis of the hydrogen produced. The arrows delineate the 'window of opportunity' where both reactions are exergonic. Reactions considered: $4C_{16}H_{34} + 128H_2O \rightarrow 196H_2 + 64CO_2$; $196H_2 + 49CO_2 \rightarrow 49CH_4 + 98H_2O$.

Incomplete oxidation of hexadecane to acetate and H_2 linked to acetoclastic methanogenesis and methanogenic CO_2 reduction

As with complete oxidation of hexadecane, the free energy yield of incomplete oxidation of hexadecane to acetate and H_2 is highly dependent on H_2 concentration (Figure 6). The range of H_2 concentrations where oxidation of hexadecane to acetate and H_2 is exergonic is also dependent on the acetate concentration (Figure 7). As acetate concentration decreases, the range of H_2 concentrations that are permissive for incomplete oxidation of hexadecane linked to methanogenic CO_2 reduction increases. Furthermore, the higher the temperature the more permissive incomplete hexadecane oxidation becomes with respect to H_2 concentration (Figure 7). The effect of acetate concentration on incomplete oxidation of hexadecane is therefore considerable, but is dependent upon H_2 concentration.

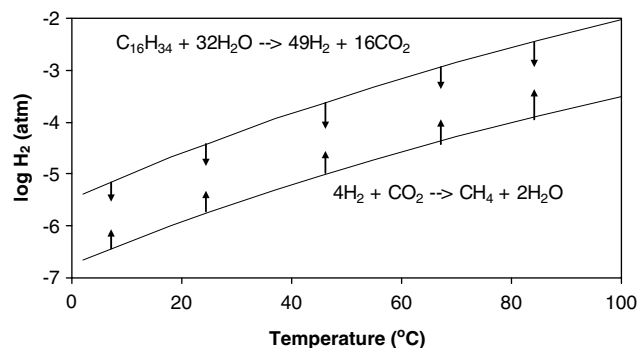


Figure 5 Effect of temperature on the range of H_2 partial pressures where both hexadecane oxidation and methanogenesis from H_2/CO_2 are exergonic. The lines represent the threshold at which the free energy change for each process is equal to zero and the arrows indicate conditions under which the processes become increasingly exergonic.

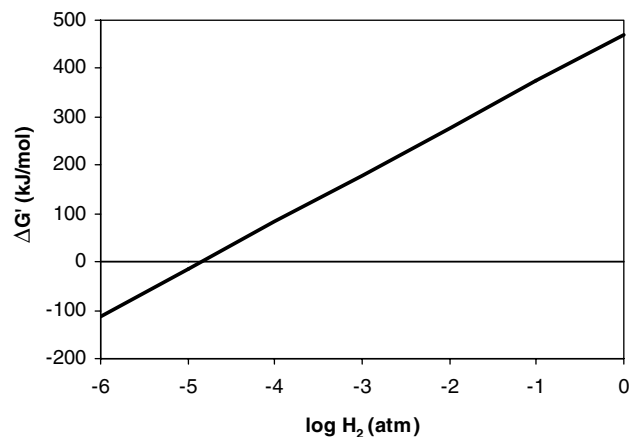


Figure 6 Effect of hydrogen partial pressure on the change in Gibbs free energy for incomplete oxidation of hexadecane to acetate and H_2 , under otherwise standard conditions, that is acetate 1 M, hexadecane as liquid and pH = 7.

Oxidation of hexadecane to acetate and H_2 linked to syntrophic acetate oxidation and methanogenesis from CO_2 reduction

A variant of incomplete hexadecane oxidation linked to acetoclastic methanogenesis and methanogenic CO_2 reduction could involve syntrophic acetate oxidation linked exclusively to methanogenic CO_2 reduction, with no involvement of acetoclastic methanogenesis. Crude oil has been shown to have an adverse effect on acetoclastic methanogenesis (Warren *et al.*, 2004); under such circumstances it is possible that syntrophic acetate oxidation might permit complete conversion of hexadecane to methane and CO_2 via methanogenic CO_2 reduction. Interestingly, the range of H_2 concentrations that permit hexadecane oxidation linked to methanogenic CO_2 reduction is least for incomplete oxidation and greatest for syntrophic acetate oxidation, which is exergonic at H_2 partial pressures

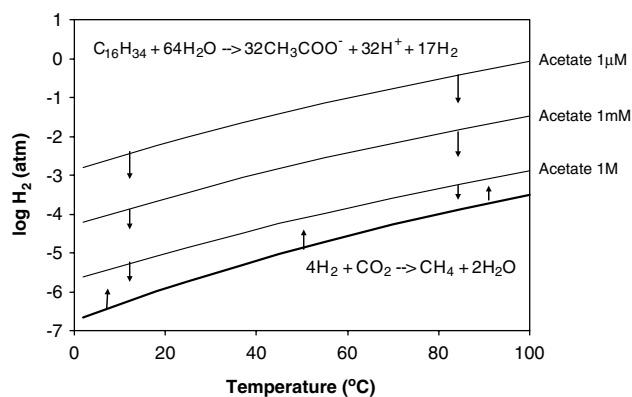


Figure 7 Effect of temperature on the range of H_2 partial pressures where both incomplete oxidation of hexadecane to acetate and H_2 and methanogenic CO_2 reduction are exergonic. The thin lines demarcate the hydrogen thresholds where the free energy yield equals zero at acetate concentrations of 1 μM , 1 mM and 1 M respectively. The thick line gives the threshold for methanogenic CO_2 reduction. The arrows indicate conditions under which the processes become increasingly exergonic.

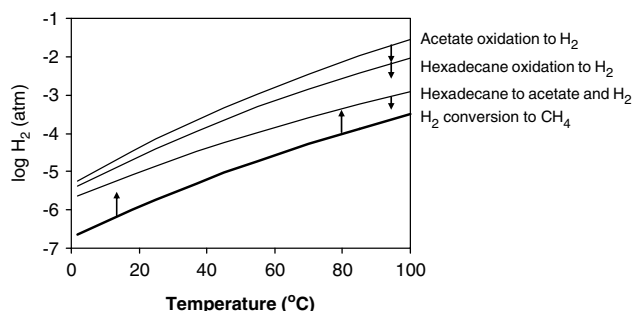


Figure 8 Effect of temperature on the range of H_2 partial pressures where complete oxidation of acetate to H_2 , complete oxidation of hexadecane to H_2 and CO_2 and incomplete oxidation of hexadecane to acetate and H_2 are exergonic and can be coupled to methanogenic CO_2 reduction. The thin lines demarcate the hydrogen thresholds for acetate oxidation and for hexadecane oxidation; the thick line gives the threshold for methanogenic CO_2 reduction. Standard states are: acetate 1 M, hexadecane as liquid and CO_2 and CH_4 at 1 atm.

slightly greater than required to give a negative ΔG for complete oxidation of hexadecane to H_2 and CO_2 (Figure 8). However, as acetate concentrations decrease (less than 0.1 M acetate), syntrophic acetate oxidizers require lower H_2 concentrations than either complete or incomplete hexadecane oxidizers (Figure 9; sector II and III compared to sector IV) and the feasibility of this route is therefore dictated by the sensitivity of syntrophic acetate oxidation to H_2 concentration.

Oxidation of hexadecane to acetate alone, linked to acetoclastic methanogenesis

A further route for the primary oxidation of alkanes is their conversion to acetate alone, which may then

be linked to acetoclastic methanogenesis. This pathway becomes thermodynamically less favourable with increasing temperature (Figure 10). Like incomplete hexadecane oxidation, this mechanism of primary oxidation of hexadecane, is dependent on the acetate concentration. Under otherwise standard conditions, this route of hexadecane oxidation becomes exergonic at concentrations less than 0.1 M acetate (Figure 9). This contrasts with incomplete hexadecane oxidation which is exergonic at higher acetate concentrations provided that the H_2 partial pressure is maintained at less than 1.4×10^{-5} atm (Figure 9, sector V; see also section 3.4).

Oxidation of hexadecane to acetate alone linked to syntrophic acetate oxidation and methanogenic CO_2 reduction

A variation on complete acetate oxidation linked to methanogenesis involves syntrophic acetate oxidation and methanogenic CO_2 reduction as the terminal reaction. This metabolic route has the effect of reducing the domain under which conversion of hexadecane to methane is feasible (Sector I through VI in Figure 9, for hexadecane conversion to acetate linked to acetoclastic methanogenesis, compared to Sector I through IV in Figure 9 if syntrophic acetate oxidation was involved). This is because acetoclastic methanogenesis is independent of H_2 concentration, but methanogenic CO_2 reduction is prevented at low H_2 concentrations, and in addition oxidation of hexadecane to acetate alone is prevented at acetate concentrations greater than 0.1 M (Figure 9).

Effect of pH on incomplete oxidation and acetogenic oxidation of hexadecane

Both incomplete oxidation of hexadecane and conversion of hexadecane to acetate are pH dependent. Both processes become increasingly energetically favourable with increasing pH. In line with the stoichiometry of acetate production, the effect is more pronounced for complete conversion of hexadecane to acetate (Figure 11). Higher pH values also increase the range of acetate concentrations where the reaction remains exergonic (Figure 12).

Discussion

The methanogenic alkane degradation landscape

Microbial degradation of hydrocarbons under anoxic conditions is of global significance. It is responsible for the in-reservoir degradation of oil that has led to vast heavy oil and tar sand deposits that constitute the bulk of the world's petroleum inventory (Roadifer, 1987; Head *et al.*, 2003; Aitken *et al.*, 2004). Despite the importance of the process, relatively little is known about the

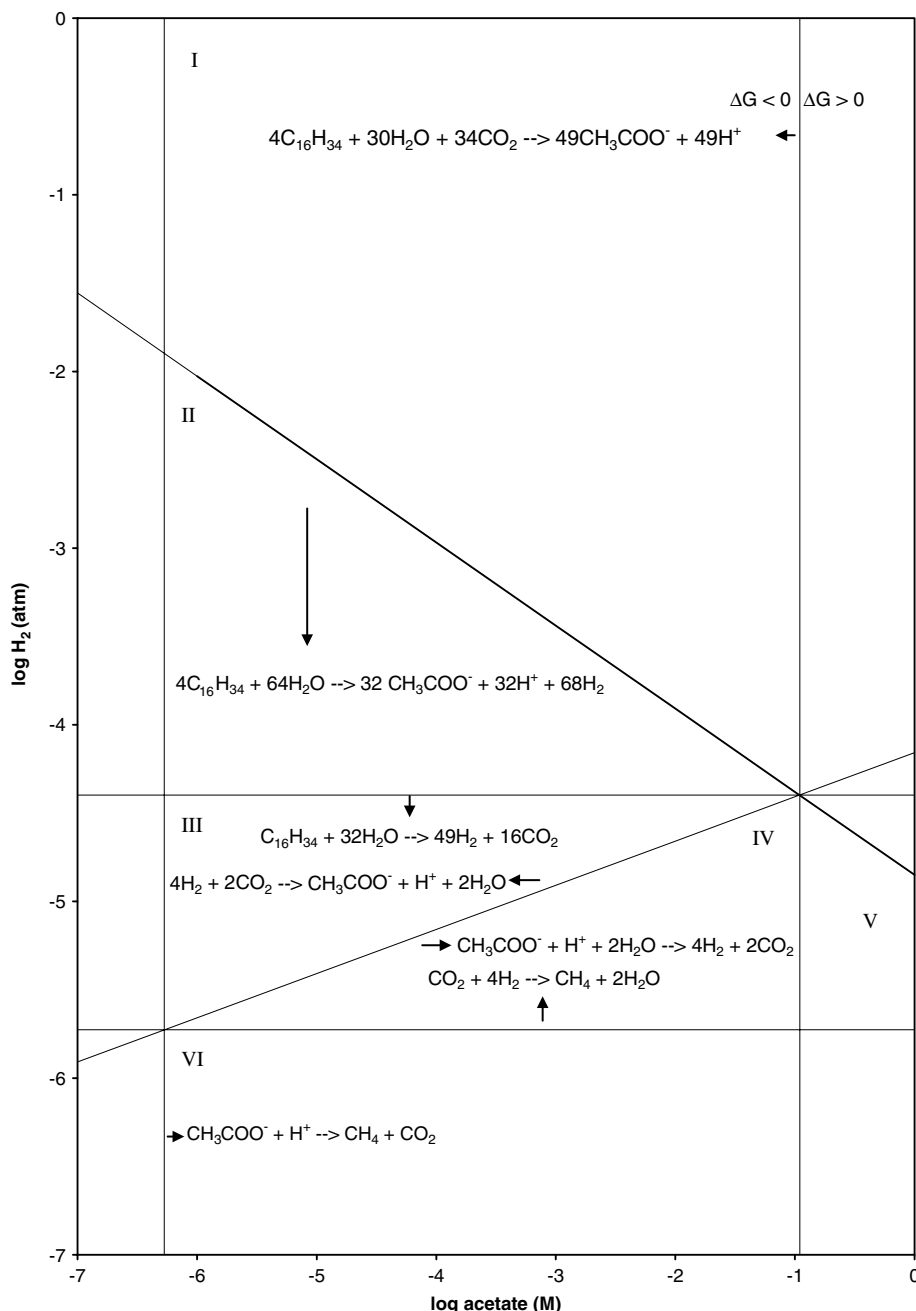


Figure 9 Hydrogen and acetate as thermodynamic constraints on methanogenic hexadecane degradation.

factors which dictate the occurrence of in-reservoir biodegradation. The geothermal history of petroleum basins has been shown to exert a broad control on the occurrence of biodegraded petroleum reservoirs and has led to the concept of palaeopasteurization or palaeosterilization, which prevents the biodegradation of crude oil in subterranean formations (Wilhelms *et al.*, 2001). The limited availability of nutrients in aquifers associated with petroleum reservoirs and geological constraints on the physical interaction of water and oil legs have also been proposed as factors which may limit in-reservoir oil biodegradation (Head *et al.*, 2003;

Larter *et al.*, 2006). As we have seen, there are several possible routes that lead to the conversion of crude oil hydrocarbons to methane, and the interplay between physical and chemical conditions in the reservoir and reservoir microbiology likely have a role in controlling in-reservoir oil biodegradation. In addition to these broad scale geological controls, thermodynamic factors may also have a bearing on crude oil biodegradation in petroleum reservoirs.

Thermodynamic calculations demonstrate that methanogenic alkane degradation becomes more energetically favourable at higher temperatures and

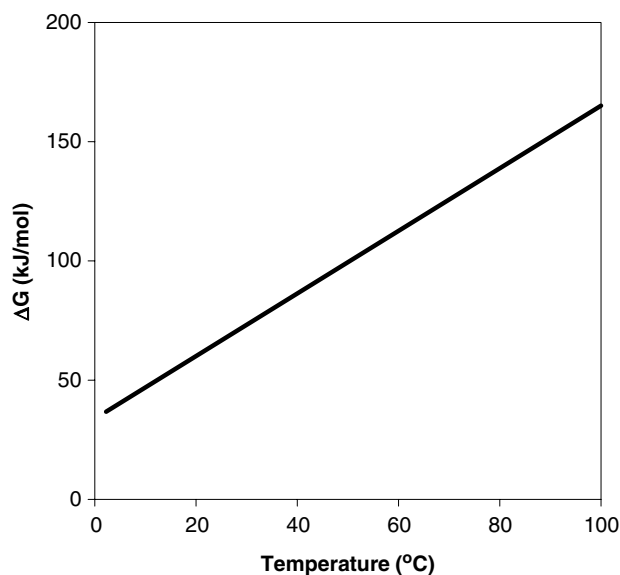


Figure 10 Effect of temperature on change in Gibbs free energy for oxidation of hexadecane to acetate alone.

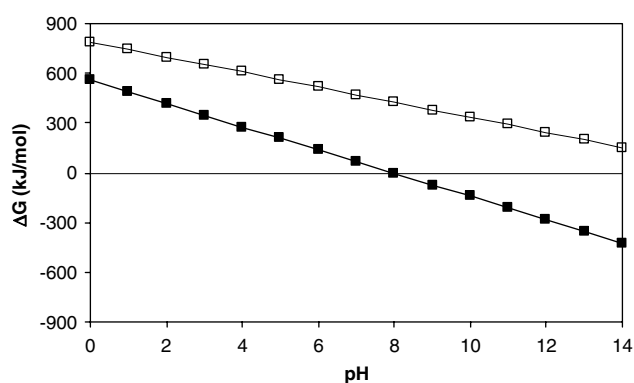


Figure 11 Effect of pH on change in Gibbs free energy for complete oxidation of hexadecane to acetate (closed symbols) and for incomplete oxidation to H_2 and acetate (open symbols).

although the three potential initial oxidation reactions examined here are all endergonic under standard conditions, at low concentrations of H_2 and/or acetate they are all thermodynamically feasible. Acetate concentrations ranging from less than $10 \mu M$ to over 17 mM have been measured in oil-field formation waters (Barth and Riis, 1992) with the majority (85 of 121 samples analyzed) having acetate concentrations below 5 mM . Such concentrations are well within the permissive range for the hydrocarbon oxidation reactions investigated here (Figure 9). There are very few data in the literature on hydrogen gas concentrations in petroleum reservoirs. Hydrogen however, seems to be present below detection limits in most oilfield waters examined and is unlikely to represent more than $1 \text{ mol}\%$ (equivalent to 10^{-2} atm) in associated gases (Hill *et al.*, 2007). This again is compatible with the

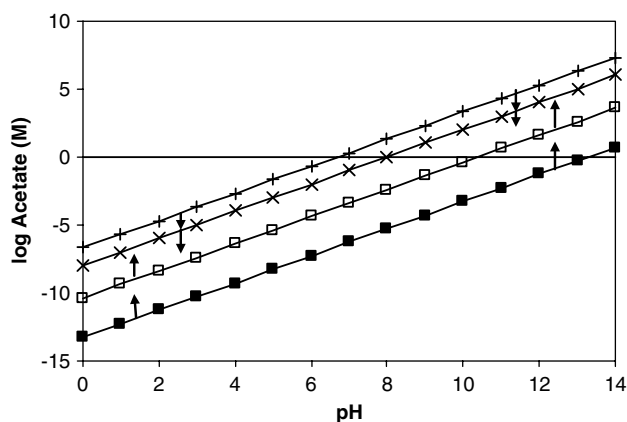


Figure 12 Effect of pH on the range of acetate concentrations where hexadecane conversion to acetate or acetate plus hydrogen are exergonic and can be coupled to methanogenesis from acetate at a hydrogen partial pressure of 10^{-5} atm ; symbols: +, incomplete oxidation of hexadecane to acetate and H_2 ; x, oxidation of hexadecane to acetate; open squares, oxidation of acetate to H_2 and CO_2 ; closed squares, conversion of acetate to CH_4 . The arrows indicate conditions under which the processes become increasingly exergonic.

hydrocarbon oxidation pathways examined here (Figure 9). However, it should be borne in mind that the gas data were obtained from well-head samples and may not reflect the true concentrations present in deep petroleum reservoirs.

To evaluate how the different initial oxidation processes can be linked to conventional methanogenic pathways we have introduced the concept of a ‘window of opportunity’, which defines the conditions under which both the initial oxidation process and the terminal methanogenic pathways are energetically favourable. This demonstrates that as temperature increases, so does the window of opportunity with respect to H_2 and acetate. Clearly this is only part of the story as it is well known that at temperatures in excess of $80\text{--}90^\circ\text{C}$, in-reservoir petroleum biodegradation apparently ceases (Connan, 1984; Head *et al.*, 2003), illustrating that above these temperatures biological factors are more important than thermodynamic factors in controlling methanogenic hydrocarbon degradation. The windows of opportunity with respect to acetate and H_2 , the central intermediates in methanogenic alkane degradation, have been summarized for the range of processes that are feasibly involved in methanogenic alkane degradation. Figure 9 represents the situation at 25°C and $\text{pH } 7$; however, the activity domains identified from this analysis remain similar for other temperatures and pH values, with the proviso that they are shifted towards higher H_2 and lower acetate concentrations at higher temperatures (see Supplementary Information). The value of this analysis is that it identifies clear zones where different methanogenic alkane degradation pathways can occur. For example the window of opportunity for linking complete

conversion of hexadecane to acetate with acetoclastic methanogenesis (domain I to IV and VI in Figure 9) is much larger than the equivalent window for linking incomplete oxidation of hexadecane to both acetoclastic methanogenesis and methanogenic CO₂ reduction (domain II, III, IV and VI in Figure 9), or complete oxidation of hexadecane to methanogenic CO₂ reduction (domain III, IV and V in Figure 9). This might suggest that acetoclastic methanogenesis might predominate in methanogenic oil-degrading systems. However, several lines of evidence suggest that methanogenic CO₂ reduction, may be more prevalent in petroleum systems. The majority of methanogens identified from oil field waters are CO₂-reducing methanogens, and acetoclastic methanogens are apparently rare (Magot *et al.*, 2000; Orphan *et al.*, 2000, 2003; Grabowski *et al.*, 2005). Furthermore, experimental measurements of methanogenic pathways in oil field waters also suggest that CO₂ reduction to methane may be more important than acetoclastic methanogenesis in petroleum reservoirs. A synthesis of data from 138 measurements made across 6 different oilfields (Nazina *et al.*, 1995a,b; Rozanova *et al.*, 1995, 2001; Bonch-Osmolovskaya *et al.*, 2003) indicates that in 67% of the cases, the predominant methanogenic pathway was CO₂ reduction, whereas in 33% acetoclastic methanogenesis predominated. This is supported by field data which demonstrate that acetate is present at high concentrations in some oil field waters (Barth and Riis, 1992), and by modelling of gas isotope composition during in-reservoir biodegradation and data from methanogenic oil-degrading microcosms (Jones *et al.*, 2008 accepted for publication). Studies of methanogenesis in a crude oil contaminated aquifer also suggest that crude oil suppresses acetoclastic methanogenesis (Warren *et al.*, 2004). Interestingly predominance of acetoclastic methanogens has been observed in hydrocarbon contaminated aquifers (Dojka *et al.*, 1998; Struchtemeyer *et al.*, 2005). However, in these instances, refined petroleum products (jet fuel) and a gas condensate were the contaminants, not crude oil. Taken together these suggest that there are biological controls that override thermodynamic considerations and lead to a subordinate role for processes of crude oil alkane degradation that require a major role for acetoclastic methanogenesis. The thermodynamic analysis; however, demonstrates that there is a wide range of conditions under which alkane degradation can be supported by methanogenic CO₂ reduction linked either to the complete oxidation of alkanes (domain III to V), or incomplete oxidation of alkanes where the acetate generated can be oxidized to H₂ and CO₂ by syntrophic acetate oxidizing organisms (domain II to V in Figure 9). The window of opportunity, which is permissive for the pathway involving incomplete alkane oxidation coupled to syntrophic acetate oxidation and methanogenic CO₂ reduction is larger than the corresponding window for complete alkane

oxidation coupled to methanogenic CO₂ reduction. Furthermore, if one considers the line relating ΔG to hydrogen concentration for complete and incomplete alkane oxidation, it is apparent that incomplete oxidation is more favourable at higher hydrogen concentrations and the slope is much steeper for complete oxidation (see Supplementary Figure S6). The second of these observations indicates that complete oxidation of alkanes is likely be more sensitive to small fluctuations in hydrogen concentrations around the point that the process shifts from being exergonic to endergonic. Thus under conditions where acetoclastic methanogenesis is inhibited one would expect a pathway via incomplete oxidation of alkanes linked to syntrophic acetate oxidation and methanogenic CO₂ reduction, rather than a pathway via complete oxidation to H₂/CO₂ linked to methanogenic CO₂ reduction. Interestingly, putative thermophilic acetate-oxidizing bacteria have been detected in a petroleum reservoir (Nazina *et al.*, 2006). Coupled with the inability to detect acetoclastic methanogens in formation waters or enrichment cultures, even when high rates of methanogenesis from acetate were measured, this has led to the suggestion that syntrophic acetate oxidation might be important for driving methanogenesis in a high temperature oil reservoir (Nazina *et al.*, 2006). Therefore it is feasible that the principal pathways leading to methanogenic oil degradation in petroleum reservoirs involve either complete oxidation of hydrocarbons to H₂ and CO₂ followed by methanogenic CO₂ reduction or incomplete hydrocarbon oxidation coupled with syntrophic acetate oxidation and methanogenic CO₂ reduction with the latter of these likely to occur more widely.

Acknowledgements

JD and IMH acknowledge funding from the European Commission, which supported this work through ECO-SERV, a Marie Curie Excellence Grant (EXT 023469) and the Natural Environment Research Council (Grant No NE/E01657X/1). SRL acknowledges support from Alberta Ingenuity, NSERC and CFI.

References

- Aitken CM, Jones DM, Larter SR. (2004). Anaerobic hydrocarbon biodegradation in deep subsurface oil reservoirs. *Nature* **431**: 291–294.
- Amend JP, Shock EL. (2001). Energetics of overall metabolic reactions of thermophilic and hyperthermophilic archaea and bacteria. *FEMS Microbiol Rev* **25**: 175–243.
- Anderson RT, Lovley DR. (2000). Hexadecane decay by methanogenesis. *Nature* **404**: 722–723.
- Barth T, Riis M. (1992). Interactions between organic acid anions in formation waters and reservoir mineral phases. *Org Geochem* **19**: 455–482.

- Bekins BA, Hostettler FD, Herkelrath WN, Delin GN, Warren E, Essaid HI. (2005). Progression of methanogenic degradation of crude oil in the subsurface. *Environ Geosci* **12**: 139–152.
- Bonch-Osmolovskaya EA, Miroshnichenko ML, Lebedinsky AV, Chernyh NA, Nazina TN, Ivoilov VS *et al*. (2003). Radioisotopic, culture-based and oligonucleotide microchip analyses of thermophilic microbial communities in a continental high-temperature petroleum reservoir. *Appl Environ Microbiol* **69**: 6143–6151.
- Boreham CJ, Hope JM, Hartung-Kagi B. (2001). Understanding source, distribution and preservation of Australian natural gas: a geochemical perspective. *APPEA J* **41**: 523–547.
- Connan J. (1984). Biodegradation of crude oils in reservoirs. In: Brooks J, Welte DH (eds). *Advances in Petroleum Geochemistry*, vol. 1. Academic Press: London, pp 299–335.
- Dojka MA, Hugenholtz P, Haack SK, Pace NR. (1998). Microbial diversity in a hydrocarbon- and chlorinated-solvent-contaminated aquifer undergoing intrinsic bioremediation. *Appl Environ Microbiol* **64**: 3869–3877.
- Grabowski A, Nercessian O, Fayolle F, Blanchet D, Jeanthon C. (2005). Microbial diversity in production waters of a low-temperature biodegraded oil reservoir. *FEMS Microbiol Ecol* **54**: 427–443.
- Hanselmann KW. (1991). Microbial energetics applied to waste repositories. *Experientia* **47**: 645–687.
- Harned HS, Owen BB. (1943). *The Physical Chemistry of Electrolytic Solutions*. Reinhold Publishing Corporation: New York.
- Head IM, Jones DM, Larter SR. (2003). Biological activity in the deep subsurface and the origin of heavy oil. *Nature* **426**: 344–352.
- Helgeson HC, Owens CE, Knox AM, Richard L. (1998). Calculation of the standard molal thermodynamic properties of crystalline, liquid, and gas organic molecules at high temperatures and pressures. *Geochim Cosmochim Acta* **62**: 985–1081.
- Hill RJ, Jarvie DM, Zumberge J, Henry M, Pollastro RM. (2007). Oil and gas geochemistry and petroleum systems of the Fort Worth Basin. *AAPG Bull* **91**: 445–473.
- Jones DM, Head IM, Gray ND, Adams JJ, Rowan AK, Aitken CM *et al*. (2008). Crude oil biodegradation via methanogenesis in subsurface petroleum reservoirs. *Nature*, doi:10.1038/nature06484.
- Larter S, Hockey A, Aplin A, Telnaes N, Wilhelms A, Horstad I *et al*. (1999). When biodegradation preserves petroleum! Petroleum geochemistry of N. Sea Oil Rimmed Gas Accumulations (ORGAs). *Proceedings AAPG Hedberg Research Conference on 'Natural Gas Formation and Occurrence'*. American Association of Petroleum Geochemists: Tulsa, Oklahoma, pp 3–5.
- Larter S, Huang H, Adams J, Bennett B, Jokanolaa F, Oldenburgh T *et al*. (2006). The controls on the composition of biodegraded oils in the deep subsurface: (part II). Geological controls on subsurface biodegradation fluxes and constraints on reservoir fluid property prediction. *AAPG Bull* **90**: 921–938.
- Larter S, Wilhelms A, Head I, Koopmans M, Aplin A, Di Primio R *et al*. (2003). The controls on the composition of biodegraded oils in the deep subsurface—part 1: biodegradation rates in petroleum reservoirs. *Org Geochem* **34**: 601–613.
- Magot M, Ollivier B, Patel BKC. (2000). Microbiology of petroleum reservoirs. *Antonie van Leeuwenhoek* **77**: 103–116.
- Manning DAC, Hutcheon IE. (2004). Distribution and mineralogical controls on ammonium in deep groundwaters. *Appl Geochem* **19**: 1495–1503.
- Masterson WD, Dzou LIP, Holba AG, Fincannon AL, Ellis L. (2001). Evidence for biodegradation and evaporative fractionation in West Sak, Kuparuk and Prudhoe Bay field areas, North Slope, Alaska. *Org Geochem* **32**: 411–441.
- Nazina TM, Ivanova AE, Borzenkov IA, Belyaev SS, Ivanov MV. (1995a). Occurrence and geochemical activity of microorganisms in high-temperature, water-flooded oil fields of Kazakhstan and Western Siberia. *Geomicrobiol J* **13**: 181–192.
- Nazina TN, Ivanova AE, Golubeva OV, Ibatullin RR, Belyaev SS, Ivanov MV. (1995b). Occurrence of sulfate- and iron-reducing bacteria in stratal waters of the Romashkinskoe oil field. *Mikrobiologiya (Eng Trans)* **64**: 203–208.
- Nazina TN, Shestakova NM, Grigor'yan AA, Mikhailova EM, Tourova TP, Poltarus AB *et al*. (2006). Phylogenetic diversity and activity of anaerobic microorganisms of high-temperature horizons of the Dagang oil field (P R China). *Microbiology* **75**: 55–65.
- Orphan VJ, Goffredi SK, DeLong EF, Boles JR. (2003). Geochemical influence on diversity and microbial processes in high temperature oil reservoirs. *Geomicrobiol J* **20**: 295–311.
- Orphan VJ, Taylor LT, Hafenbradl D, DeLong EF. (2000). Culture-dependent and culture-independent characterization of microbial assemblages associated with high-temperature petroleum reservoirs. *Appl Environ Microbiol* **66**: 700–711.
- Pallasser RJ. (2000). Recognising biodegradation in gas/oil accumulations through the $\delta^{13}\text{C}$ compositions of gas components. *Org Geochem* **31**: 1363–1373.
- Parkes J. (1999). Cracking anaerobic bacteria. *Nature* **401**: 217–218.
- Parkes RJ, Webster G, Cragg BA, Weightman AJ, Newberry CJ, Ferdelman TG *et al*. (2005). Deep sub-seafloor prokaryotes stimulated at interfaces over geological time. *Nature* **436**: 390–394.
- Roadifer RE. (1987). Size distributions of the World's largest known oil and tar accumulations. In: Meyer RF (ed). *Exploration for Heavy Crude Oil and Natural Bitumen*: AAPG Studies in Geology 25. American Association of Petroleum Geologists: Tulsa, pp 3–23.
- Rozanova EP, Borzenkov IA, Tarasov AL, Suntsova LA, Dong CL, Belyaev SS *et al*. (2001). Microbiological processes in a high-temperature oil field. *Microbiology* **70**: 102–110.
- Rozanova EP, Savvichev AS, Karavaiko SG, Miller YM. (1995). Microbial processes in the Savuiskoe oil field in the Ob' region. *Mikrobiologiya (Eng Trans)* **64**: 85–90.
- Scott AR, Kaiser WR, Ayers WBJ. (1994). Thermogenic and secondary biogenic gases, San-Juan Basin, Colorado and New-Mexico—implications for coalbed gas producibility. *AAPG Bull* **78**: 1186–1209.
- Struchtemeyer CG, Elshahed MS, Duncan KE, McInerney MJ. (2005). Evidence for acetoclastic methanogenesis in the presence of sulfate in a gas condensate-contaminated aquifer. *Appl Environ Microbiol* **71**: 5348–5353.
- Sweeney RE, Taylor P. (1999). Biogenic methane derived from biodegradation of petroleum under environmental conditions and in oil & gas reservoirs. In: Schoell M, Claypool GE (eds). *Proceedings AAPG Hedberg*

- Research Conference on Natural Gas Formation and Occurrence*. American Association of Petroleum Geochemists: Tulsa, Oklahoma, pp 6–10.
- Thauer RK, Jungermann K, Decker K. (1977). Energy conservation in chemotrophic anaerobic bacteria. *Bacteriol Rev* **41**: 100–180.
- Townsend GT, Prince RC, Suflita JM. (2003). Anaerobic oxidation of crude oil hydrocarbons by the resident microorganisms of a contaminated anoxic aquifer. *Environ Sci Technol* **37**: 5213–5218.
- Warren E, Bekins BA, Godsy EM, Smith VK. (2004). Inhibition of acetoclastic methanogenesis in crude oil- and creosote-contaminated groundwater. *Bioremediation J* **8**: 1–11.
- Weiner J, Lovley DR. (1998). Rapid benzene degradation in methanogenic sediments from a petroleum-contaminated aquifer. *Appl Environ Microbiol* **64**: 1937–1939.
- Widdel F, Rabus R. (2001). Anaerobic biodegradation of saturated and aromatic hydrocarbons. *Curr Opin Biotechnol* **12**: 259–276.
- Wilhelms A, Larter SR, Head I, Farrimond P, Di-Primio R, Zwach C. (2001). Biodegradation of oil in uplifted basins prevented by deep-burial sterilization. *Nature* **411**: 1034–1037.
- Zengler K, Richnow HH, Rosselló-Mora R, Michaelis W, Widdel F. (1999). Methane formation from long-chain alkanes by anaerobic microorganisms. *Nature* **401**: 266–269.

Supplementary Information accompanies the paper on The ISME Journal website (<http://www.nature.com/ismej>)