

## DATA REPORT

The first Japanese patient with mandibular hypoplasia, deafness, progeroid features and lipodystrophy diagnosed via *POLD1* mutation detectionAsami Okada<sup>1,5</sup>, Tomohiro Kohmoto<sup>2,5</sup>, Takuya Naruto<sup>2</sup>, Ichiro Yokota<sup>1,3</sup>, Yumiko Kotani<sup>1</sup>, Aki Shimada<sup>3,4</sup>, Yoko Miyamoto<sup>2</sup>, Rizu Takahashi<sup>2</sup>, Aya Goji<sup>1</sup>, Kiyoshi Masuda<sup>2</sup>, Shoji Kagami<sup>1</sup> and Issei Imoto<sup>2</sup>

Mandibular hypoplasia, deafness, progeroid features and lipodystrophy (MDPL) syndrome is a rare autosomal dominant disorder caused by heterozygous *POLD1* mutations. To date, 13 patients affected by *POLD1* mutation-caused MDPL have been described. We report a clinically undiagnosed 11-year-old male who noted joint contractures at 6 years of age. Targeted exome sequencing identified a known *POLD1* mutation [NM\_002691.3:c.1812\_1814del, p.(Ser605del)] that diagnosed him as the first Japanese/East Asian MDPL case.

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Mandibular hypoplasia, deafness, progeroid features and lipodystrophy (MDPL, MIM#615381) syndrome is a rare autosomal dominant systemic disorder resulting from heterozygous mutations in *POLD1* (MIM#174761).<sup>1–3</sup> MDPL is clinically characterized by prominent loss of subcutaneous fat, characteristic facial appearance, metabolic abnormalities involving insulin resistance and diabetes mellitus, and sensorineural deafness occurring late in the first or second decades of life. To date, 18 patients affected by this syndrome have been described, including 1 Indian, 2 Hispanic and 15 Caucasians.<sup>1,2,4–6</sup> Among the 13 *POLD1* mutation-caused MDPL cases, all have been caused by one of two different *POLD1* mutations: an in-frame deletion (Ser605del, 11 cases) and a missense mutation (R507C, 2 cases).<sup>2,4–6</sup> In most of these cases, the mutation occurred *de novo*.<sup>2,4–6</sup> Although these *POLD1* mutations appear to occur in genic hotspots regardless of race/ethnicity, no East Asian cases (including Japanese) have been reported.

We herein report the first Japanese/East Asian case of MDPL in an 11-year-old male with characteristics of MDPL. We used targeted exome sequencing (TES) as a genome-first approach in a clinically undiagnosed Japanese patient and determined that he was carrying a known heterozygous *POLD1* mutation.

The patient was an 11-year-old, first-born male child of healthy, nonconsanguineous Japanese parents with unremarkable family history (Figure 1a). He was born through normal vaginal delivery at full term with birth weight of 2.692 kg (−0.8 s.d.), body length of 46.6 cm (−1.1 s.d.) and occipitofrontal circumference of 33.2 cm (−0.1 s.d.). His early developmental milestones were normal, but his parents noticed poor height and weight gain when he was 3 years of age. At 6 years of age, school teachers noticed that he was unable to either perform kicking motions while swimming or sit on his heels, and an orthopedist pointed out the presence of joint contractures. At 7 years of age, he was referred to a pediatric

endocrinology department due to his short stature. He presented with prominent eyes, beaked nose, mandibular hypoplasia, crowded teeth, small mouth and testicular hypoplasia. He was also diagnosed with moderate sensorineural bilateral hearing loss and he started to wear hearing aids. Growth hormone (GH) levels in a GH stimulation test were observed to be normal. Standard karyotyping using peripheral blood revealed no abnormalities (46, XY). No results consistent with known metabolic syndromes were obtained from metabolic surveys, including serum amino acids and urine organic acids. At 9 years of age, tight skin around his cheeks, hepatic steatosis and mild liver dysfunction were noted. At the age of 11, due to his short stature (weight; 24.6 kg (−1.7 s.d.), height; 125.2 cm (−2.9 s.d.) and body mass index; 15.7), facial features, deafness, primary hypogonadism, joint contractures and thin arms and legs with a wide trunk, the patient visited a division of clinical genetics to consider any genetic diseases (with informed consent from his parents). Although the patient's clinical features were retrospectively consistent with the recently proposed clinical spectrum of MDPL caused by *POLD1* mutations (Table 1),<sup>1,2,4–6</sup> the patient remained undiagnosed. As a result, TES was considered using a panel of multiple potential disease-causing genes.

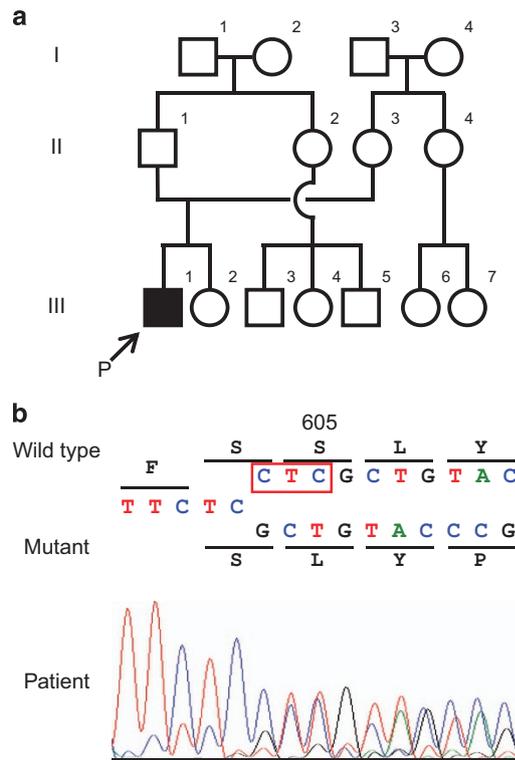
After informed consent was obtained from the parents, molecular diagnosis was performed using genomic DNA extracted from the patient's blood sample. The study was approved by the ethics committees of Tokushima University. To screen known disease-associated genes for molecular diagnosis, we used a TruSight One Sequencing Panel (Illumina, San Diego, CA, USA) with a MiSeq sequencer (Illumina), followed by our pipeline for next generation sequencing (NGS) data analysis as previously described,<sup>7,8</sup> with a minor modification due to a software update specific for a bioinformatics pipeline.<sup>8</sup> To identify presumably pathogenic single-nucleotide variants, we excluded sequence

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**Figure 1.** (a) Family pedigree; arrow shows the proband (P). (b) Partial sequence chromatograms around codon 605 on exon 15 of *POLD1* in the patient. The red box denotes the deleted bases. The DNA and corresponding amino acid sequences of the wild-type and mutant *POLD1* alleles are also shown.

variants with low-allele frequencies, that is, >0.01 included in the 1000 Genomes Project database (<http://www.1000genomes.org>), National Heart, Lung, and Blood Institute Grand Opportunity Exome Sequencing Project (ESP6500, <http://evs.gs.washington.edu/EVS>), Human Genetic Variation Database (<http://www.genome.med.kyoto-u.ac.jp/SnpDB>) and integrative Japanese Genome Variation Database (<https://ijgvd.megabank.tohoku.ac.jp>). Copy-number variations analysis using TES data was also performed as described elsewhere.<sup>8,9</sup> These analyses detected an in-frame heterozygous deletion in exon 15 of *POLD1*, NM\_002691.3 (*POLD1*\_v001):c.1812\_1814del, affecting the polymerase-active site, NM\_002691.3(*POLD1*\_i001):p.(Ser605del), which was confirmed by Sanger sequencing (Figure 1b). This mutation has been shown to cause most cases of MDPL.<sup>2,6</sup> No other variants or gross deletions were detected in the coding regions of other progeroid-related genes (data not shown). As a result of this molecular diagnosis and the re-evaluation of the affected patient's clinical features, together with the clinical spectrum of patients harboring *POLD1* mutations (Table 1),<sup>1,2,4-6</sup> the patient was diagnosed with MDPL caused by a known frameshift deletion in *POLD1*. Because parental DNA was not available, we were unable to determine if the mutation occurred *de novo*.

To the best of our knowledge, the patient described herein is the 19th MDPL and the 14th *POLD1* mutation-caused MDPL case reported worldwide. Notably, this patient is the first Japanese or East Asian case with MDPL, which is caused by the most common *POLD1* in-frame deletion mutation (c.1812\_1814del). Our case supports the hypothesis that *POLD1* mutations causing MDPL, at least this in-frame deletion mutation, commonly occur at hotspots irrespective of race/ethnicity. The CTCCT motif occurs within two CTC triples, one of which is deleted in most MDPL cases, and corresponds to the complement of the mirror image of the 'deletion hotspot consensus sequences' TG(A/G) (A/G) (G/T) (A/C).<sup>10,11</sup> In addition, (A/T)GGAG is

**Table 1.** Clinical characteristics of the *POLD1* mutation-caused MDPL patient presented here compared to previously described subjects

Clinical features	Study patient	Previous studies <sup>a</sup> (n = 13)
Age (years; range, median)	11	10–62, 25
Sex	Male	6 males, 7 females
Birth weight (kg; range, mean)	2.692	2.4–4.2, 3.23 (n = 9)
Height (cm)	125.2	—
Weight (kg)	24.58	—
BMI (kg/m <sup>2</sup> ; range, mean)	15.7	13.8–26.8, 17.5
<b>Metabolic profile</b>		
Diabetes mellitus	N	5/13
Hepatic steatosis	Y	4/6
ALT (U/l)	52 (5–40)	Abnormal LFT 5/8
Total cholesterol (mg/dl)	141 (130–220)	High 8/10
Triglycerides (mg/dl)	125 (35–150)	High 9/11
Leptin (ng/ml)	10.9	4.4–8.2, 5.6 (n = 4)
<b>Morphology</b>		
Short stature	Y	8/13
Tight skin around cheeks and small nasal bones	Y	13/13
Mandibular underdevelopment	Y	12/13
Dental overcrowding/irregular teeth	Y	10/13
Telangiectasia	N	9/13
Thin arms and legs with wide trunk	Y	13/13
High pitched voice	Y	9/12
Hearing impairment	Y	10/13
<b>Musculoskeletal</b>		
Joint contractures	Y	5/13
Muscle wasting	Y	11/13
Kyphosis/scoliosis	N	4/5
Hypogonadism	Y	4/5 males
Abnormal cognitive function	N	1/12

Abbreviations: ALT, alanine aminotransferase; BMI, body mass index; LFT, liver function test; MDPL, mandibular hypoplasia, deafness, progeroid features and lipodystrophy; N, no; Y, yes.

<sup>a</sup>Previously reported cases with *POLD1* mutation-caused MDPL.

one of the specific native DNA sequences known to arrest DNA synthesis by DNA polymerase  $\alpha$ .<sup>12</sup> Therefore, the arrest of DNA synthesis at this DNA polymerase pause site may increase the possibility that a slipped mispairing was mediated by direct repeats and/or secondary structure formation promoted by symmetric elements and might cause the commonly observed c.1812\_1814del mutation in *POLD1* in MDPL patients.<sup>5,11</sup>

Although clinical characteristics of the patient, including his phenotype, history of the disease and metabolic profile showed features fully and retrospectively compliant with the diagnosis of MDPL (Table 1),<sup>1,2,4-6</sup> clinical diagnosis could not be evoked because of the rarity of this disease. Indeed, reported MDPL cases are clinically and/or genetically diagnosed at a relatively higher age (median age >20 years).<sup>1,2,4-6</sup> By facilitating differential diagnosis of this syndrome and related diseases in a cost-effective manner, molecular diagnosis by a genome-first approach may be crucially important for providing appropriate therapeutic options and optimized health care in patients with unclassified segmental progeroid syndromes.<sup>5</sup> In addition, correct diagnosis through this approach also can be useful in establishing recurrence risks and for providing appropriate genetic counseling to the family.

#### HGV DATABASE

The relevant data from this Data Report are hosted at the Human Genome Variation Database at <http://dx.doi.org/10.6084/m9.figshare.hgv.1393>.

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## COMPETING INTERESTS

The authors declare no conflict of interest.

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