

## ORIGINAL ARTICLE

# Patterns of hybridization and asymmetrical gene flow in hybrid zones of the rare *Eucalyptus aggregata* and common *E. rubida*

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The patterns of hybridization and asymmetrical gene flow among species are important for understanding the processes that maintain distinct species. We examined the potential for asymmetrical gene flow in sympatric populations of *Eucalyptus aggregata* and *Eucalyptus rubida*, both long-lived trees of southern Australia. A total of 421 adults from three hybrid zones were genotyped with six microsatellite markers. We used genealogical assignments, admixture analysis and analyses of spatial genetic structure and spatial distribution of individuals, to assess patterns of interspecific gene flow within populations. A high number of admixed individuals were detected (13.9–40% of individuals), with hybrid populations consisting of F<sub>1</sub> and F<sub>2</sub> hybrids and backcrosses in both parental directions. Across the three sites, admixture proportions were skewed towards the *E. aggregata* genetic cluster ( $x=0.56–0.65$ ), indicating that backcrossing towards *E. aggregata* is more frequent. Estimates

of long-term migration rates also indicate asymmetric gene flow, with higher migration rates from *E. aggregata* to hybrids compared with *E. rubida*. Taken together, these results indicate a greater genetic input from *E. aggregata* into the hybrid populations. This asymmetry probably reflects differences in style lengths (*E. rubida*: ~7 mm, *E. aggregata*: ~4 mm), which can prevent pollen tubes of smaller-flowered species from fertilizing larger-flowered species. However, analyses of fine-scale genetic structure suggest that localized seed dispersal (<40 m) and greater clustering between hybrid and *E. aggregata* individuals may also contribute to directional gene flow. Our study highlights that floral traits and the spatial distributions of individuals can be useful predictors of the directionality of interspecific gene flow in plant populations.

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## Introduction

Natural interspecific hybridization occurs commonly in flowering plants (Mallet, 2005), and is thought to have an important role in their evolutionary history (Grant, 1981). In many cases, however, hybridization is asymmetrical, and one species is more often the maternal parent of hybrid offspring (Tiffin *et al.*, 2001). In some cases, repeated backcrossing between hybrids and one of the parental species can lead to unidirectional introgression (Bacilieri *et al.*, 1996). In particular, the directional bias in gene flow and infiltration of genes from one species into another can be important in determining the direction of evolutionary change and species succession (Bacilieri *et al.*, 1996; Petit *et al.*, 2004). Asymmetrical hybridization is also important in a conservation context, especially when hybrids exhibit high fitness and rare species exist in small populations. Under these conditions, frequent hybridization followed by unidirectional backcrossing

towards the rare species, can further exacerbate their decline through a combination of pollen swamping and genetic assimilation (Rhymer and Simberloff, 1996). Determining which reproductive barriers influence asymmetrical gene flow is therefore important for understanding both the processes that facilitate speciation and maintain species identities.

Given the importance of asymmetrical hybridization and introgression, there has been a long standing interest in understanding the factors which drive these patterns in plants. Some of these underlying factors include: mating system variation (SI × SE rule; Lewis and Crowe, 1958), the relative proportions of parent species (Burgess *et al.*, 2005) and differences in the fitness of reciprocal crosses (Tiffin *et al.*, 2001). Flower size can also be a major structural barrier to hybridization that is often unilateral (Williams and Rouse, 1988; Gore *et al.*, 1990). This trait is especially important because flower size often differs among hybridizing species. The genus *Eucalyptus* (Myrtaceae), which forms a dominant vegetative component of the Australian continent, provides a good model system for investigating hybridization dynamics in relation to flower size. On the basis of morphological patterns, interspecific hybridization is considered widespread in *Eucalyptus* (~50% of species reportedly hybridize; Griffin *et al.*, 1988)

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and several molecular studies have detected broad-scale introgression of maternally inherited markers among numerous sympatric species (McKinnon *et al.*, 2001, 2010). In this genus, smaller flowered species are often unable to cross-fertilize with larger flowered species. This is because the pollen tubes of small flowered species are unable to grow the full length of the styles of larger flowered species (Gore *et al.*, 1990). This potential difference in fertilization success provides a testable prediction regarding the expected direction of hybridization and introgression in natural hybrid zones consisting of small and large flowered *Eucalyptus* species.

The availability of highly polymorphic genetic markers (for example, microsatellites, amplified fragment length polymorphisms) and powerful statistical analyses (for example, Bayesian clustering) has greatly improved the detection of first generation and later generation hybrids. Apart from some notable exceptions (for example, Chung *et al.*, 2005), few studies have utilized these statistical procedures to examine hybrid populations in the spatial context of their parental species. In particular, for plants with limited seed dispersal, this information can be important for inferring the direction of gene flow because early generation hybrids are expected to be located near their maternal parent. Moreover, greater spatial clustering of hybrids around one of the parental species may be important for reinforcing directional backcrossing and can contribute to subsequent asymmetrical introgression. When assessing patterns of hybridization and introgression, the spatial analysis of reproductive adult genotypes has several advantages over direct methods including reciprocal pollinations and assessments of introgression in open pollinated progeny arrays. First, for tall trees with a long generation time and time to first flowering such as *Eucalyptus*, crossing experiments are often impractical. Second, both direct techniques do not provide information regarding the establishment of backcrossed hybrids, a necessary pre-requisite for potential introgression between species.

Here, we investigate the potential for asymmetrical gene flow and introgression in putative hybrid zones of a common and a rare member of the genus *Eucalyptus*. In southeastern Australia, the uncommon forest tree *Eucalyptus aggregata* is restricted to the central and southern tablelands of New South Wales and a single locality in Victoria. This species occurs in small populations ( $n < 1000$ ; Field, 2008) in combination with more widespread species that belong to the same subgenus (*Symphyomyrtus*) and section (*Maidenaria*). *Eucalyptus aggregata* often co-occurs with *E. rubida* which is common across New South Wales, Victoria and Tasmania. These two species are well differentiated morphologically (Field *et al.*, 2009) with *E. aggregata* assigned to series *Faveolatae* and *E. rubida* to series *Viminales* (Brooker, 2000). Moreover, *E. aggregata* and *E. rubida* are well differentiated genetically on the basis of allozyme markers ( $F_{ST} = 0.59$ ; Field *et al.*, 2009). There are also strong differences in habitat preference with *E. aggregata* found on poorly drained flats and hollows, whereas *E. rubida* is found on rocky and well drained skeletal soils of loams on clay subsoils (Cayzer, 1993). Where these species co-occur, putative adult hybrids are often observed, particularly along transition zones between the parental habitats. Both species are pollinated by generalists and

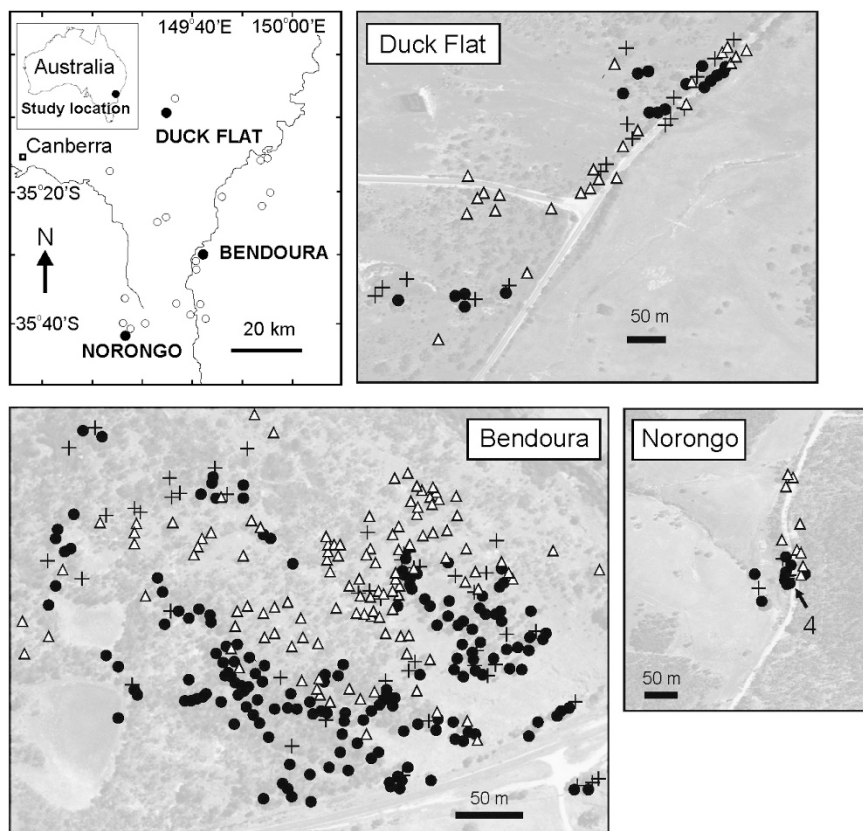
substantial overlap in flowering times (October–January for *E. rubida*, December–February for *E. aggregata*) likely provides ample opportunities for interspecific pollen flow. Morphological and molecular evidence has identified substantial hybrid frequencies in the seed arrays of *E. aggregata* where the two species occur in sympatry (Mean 8.9%; Field *et al.*, 2009), and the presence of both  $F_1$  and later generation hybrids. As the more common *E. rubida* has larger flowers (style length  $\sim 7$  mm) than *E. aggregata* (style length  $\sim 4$  mm), and both species are pollinated by generalist insects (Field *et al.*, 2008), floral morphology may be an important driver of asymmetrical gene flow. Despite the widespread occurrence of hybridization in *Eucalyptus*, little is known about the fine-scale patterns of hybridization and the factors driving asymmetrical gene flow in natural hybrid zones.

To assess the extent and direction of interspecific gene flow we use molecular markers and Bayesian analyses to examine the genomic composition of three hybrid zones consisting of *E. rubida*, *E. aggregata* and putative hybrids. A prevalence of  $F_1$  hybrids would indicate little opportunity for gene exchange, whereas a high frequency of backcross hybrids would suggest a high potential for introgression. Moreover, a skewed frequency of hybrid backcrosses in one parental direction would indicate the directionality of introgression. We also used coalescent-based methods to estimate long-term historical levels of gene flow between parental and hybrid populations. Given the differences in flower size, we predict that hybrid populations will exhibit directional gene flow towards *E. aggregata*. From these predictions we asked the following questions: (i) what is the frequency of introgression between the two species? and (ii) is asymmetrical gene flow in the direction of the smaller-flowered *E. aggregata*? In addition, we used spatial analyses and spatial autocorrelation methods to assess both the degree of spatial clustering between parental species and hybrids and the scale of seed dispersal. Given that seed dispersal is locally restricted in *Eucalyptus* (House, 1997) greater clustering between hybrids and one of the parental species could indicate which species is more often the maternal parent. Moreover, considering plant mating is more frequent among near neighbors, stronger spatial clustering between hybrids and one particular parental species may also contribute to asymmetrical gene flow. If restricted seed dispersal and spatial clustering is contributing to asymmetrical hybridization, we predict that spatial patterns should match the direction of asymmetrical gene flow. From this prediction, we ask the following questions: (iii) are hybrids spatially clustered around *E. aggregata* rather than *E. rubida*? and (iv) what is the scale of seed dispersal in *E. aggregata*, *E. rubida* and hybrid populations as indicated by fine-scale spatial genetic structure?

## Methods

### Study species and sampling

Three sites containing reproductive populations of *E. aggregata*, *E. rubida* and their putative hybrids were selected for this study on the southern tablelands of New South Wales, Australia (Figure 1). These included: Bendoura ( $35^{\circ}30'$  S,  $142^{\circ}42'$  E), a relatively large and



**Figure 1** Map showing the location of three study sites, each consisting of populations of *E. aggregata*, *E. rubida* and their putative hybrids. Individual maps of each site indicate the location of adult *E. aggregata* (closed circles), *E. rubida* (open triangles), and hybrids (plus symbol). Arrow on Norongo map indicates the location of four hybrids.

intact open woodland ( $N = 337$  adults); Duck Flat ( $33^{\circ}28' S$ ,  $150^{\circ}02' E$ ), a medium sized remnant site ( $N = 59$ ); and Norongo ( $32^{\circ}42' S$ ,  $149^{\circ}25' E$ ), a small and cluster of trees along a road side ( $N = 25$ ). At each of these sites, individual *E. aggregata* and *E. rubida* were easily distinguished on the basis of morphology. *E. aggregata* displays green elliptical-ovate-shaped leaves in alternating stem arrangements in the juvenile phase, and smaller lanceolate shaped leaves in the adult phase (12 cm long  $\times$  2.5 cm wide), brown and rough furrowed bark that is persistent into the upper branches, and has smaller flower buds (0.4  $\times$  0.3 cm) in clusters of seven. In contrast, *E. rubida* displays round glaucous juvenile leaves in opposite arrangement on the stem in the juvenile phase, has larger lanceolate-shaped leaves in the adult phase (15 cm long  $\times$  2.4 cm wide), smooth white to reddish bark and larger flowers (0.7  $\times$  0.4 cm) instead in groups of three (Brooker and Kleinig, 1999). Putative hybrids were identified on the basis of intermediate morphology between the parental species for each of the distinguishing characteristics including bark persistence, leaf size and flower size; however, hybrids always exhibited flowers in groups of seven. Adult populations were mapped using high-resolution aerial photos (1:1838; taken 2004, Lands and Property Information NSW) and by measuring the relative location of the individually-tagged trees on the ground.

To examine the genomic composition of the adult populations, mature leaves were collected for DNA and subsequent genotyping. A total of 421 trees were sampled,

which included all observed reproductive adults of each parental species and putative hybrids displaying intermediate morphology from Duck Flat ( $N = 59$ ) and Norongo ( $N = 25$ ). At the Bendoura site, a 400  $\times$  500 m area of the site containing the majority of reproductive trees was selected for sampling. Within this area, all reproductive *E. aggregata* and *E. rubida*, and all putative hybrids were sampled ( $N = 337$ ). The majority of the remaining trees at each site was the distantly related *E. pauciflora*, which is not reported to hybridize with either *E. aggregata* or *E. rubida* (Griffin et al., 1988).

#### Genetic analysis

Adult leaves were stored at  $-80^{\circ}C$  until freeze-dried for five days and then ground to powder using tungsten beads in a Mixer Mill MM 300 (Qiagen, Doncaster, Australia). DNA was extracted from up to 10 mg of the ground leaves using DNeasy 96 Plant extraction kits (Qiagen) as per the manufacturer's protocol. To reduce phenolic contaminants, 1% polyvinylpyrrolidone (molecular weight 40 000) and 5% sodium D-Isoascorbate was added to the AP1 extraction buffer.

We initially screened a panel of 40 microsatellite primer pairs developed for *Eucalyptus*. Many microsatellites have been developed for *Eucalyptus* and cross-species amplification is often successful (for example, Byrne et al., 1996). We assayed adults for six microsatellite markers selected from the panel, including four markers developed for *E. globulus*: Eg65, Eg91, Eg96



(G Moran, unpublished) and Emrc8 (Steane *et al.*, 2001), and two markers developed for *E. grandis* and *E. urophylla*: Embra10 (Brondani *et al.*, 1998), Fmrsal (van der Nest *et al.*, 2000). All of these primers were directly labeled with fluorescent dyes (FAM, NED, VIC) and the SSR fragments were amplified by PCR in a mixture of 10  $\mu$ l containing: 4.44  $\mu$ l H<sub>2</sub>O, 1  $\times$  Hotstar PCR buffer (1.5 mM MgCl<sub>2</sub>, Tris-Cl, KCl, (NH<sub>4</sub>)<sub>2</sub> SO<sub>4</sub>; Qiagen), 1.65 mM MgCl<sub>2</sub>, 0.1  $\mu$ M of the forward (fluorescently labelled) and reverse primer, 0.2  $\mu$ M of each dNTP, 10 mg ml<sup>-1</sup> bovine serum albumin, 0.05 U of Hotstar Taq (Qiagen), and 1  $\mu$ l (2–15 ng) of DNA template. Amplification of SSR fragments were performed in 384-well plates (Applied Biosystems, Mulgrave, Australia) on a Hybaid express thermocycler with a step-down PCR program consisting of 94 °C for 4 min; 15 cycles at 94 °C for 30 s, 65 °C for 30 s, 72 °C for 1 min 20 s; 30 cycles at 94 °C for 30 s, 50 °C for 30 s, 72 °C for 45 s. The final product was evaporated and 5  $\mu$ l of combined loading dye (Hi-Di Formamide: Applied Biosystems) and an internal lane standard was added, followed by denaturing at 94 °C for 5 min. The final multi-mixed PCR products were separated by capillary electrophoresis on an ABI 3130 Genetic Analyser (Applied Biosystems) and their size determined in relation to an internal lane standard (GS-500 LIZ; Applied Biosystems) using Genemapper Version 4.0 (Applied Biosystems). Individuals with low or missing peaks were amplified and genotyped a second time. No evidence of null alleles was detected at Duck Flat or Norongo using the software MICROCHECKER (van Oosterhout *et al.*, 2004). At Bendoura, there was a low probability (0.05) of a null allele at locus Fmrsal. However, we included all loci as removing the Fmrsal locus had minimal effect on the following genetic analyses.

#### Microsatellite diversity

Genetic diversity parameters were assessed separately at each site for *E. aggregata*, *E. rubida* and hybrids classified using Bayesian analyses (see below). Observed number of alleles ( $A$ ), unbiased genetic diversity ( $H_E$ ), and observed heterozygosity ( $H_O$ ) were calculated using the software GenAlEx version 6 (Peakall and Smouse, 2005). The fixation index ( $F_{IS}$ ) and departures from random mating were calculated using the software FSTAT version 2.9.3.2 (Goudet, 1995). To measure the average pair-wise level of genetic differentiation ( $F_{ST}$ ; Wright (1965)) between populations of each species and the hybrids, according to the multi-locus methods implemented in FSTAT. Each pair-wise estimate and the 95% bootstrap confidence intervals (CIs) were estimated with 1000 replicates.

#### Patterns of hybridization

We used the Bayesian methods implemented in the program STRUCTURE version 2.2 (Pritchard *et al.*, 2000; Falush *et al.*, 2003) to distinguish hybrids from pure-breds. This method uses a Markov chain Monte Carlo (MCMC) algorithm to assign individuals (as represented by their multi-locus genotypes) to  $K$  genetic clusters by minimizing within group linkage-disequilibrium and simultaneously assuming within group Hardy–Weinberg Equilibrium. Using this approach, individuals can be assigned largely to a single genetic cluster based on

admixture values (for example,  $Q_1 = 0.99$ ,  $Q_2 = 0.01$ ) or jointly to two or more if their multi-locus genotype indicates admixture due to hybridization between clusters (for example,  $Q_1 = 0.5$ ,  $Q_2 = 0.5$ ). For all analyses we used STRUCTURE with the admixture model, no previous population information, correlated allele frequencies and a burn-in period of 20 000 and 200 000 MCMC. First we conducted four independent runs at each  $K$  from two to eight, to determine the most likely number of genetic clusters and test the stability of admixture values ( $r > 0.98$ ). For the final analysis we assumed  $K = 2$  ( $Q_1 = E. aggregata$ ,  $Q_2 = E. rubida$ ) as this agrees with the presence of two species and was the most likely number of genetic clusters identified (data not shown) using the post-hoc methods of Evanno *et al.* (2005).

#### Estimates of recent introgression

In order to assess the rate and direction of recent gene flow among populations at each site, we classified individuals further into genealogical classes using the methods implemented in the NEWHYBRIDS program (Anderson and Thompson, 2002). This analysis is a probability-based model, which computes through MCMC, the posterior probability,  $q_i$ , of individuals belonging to distinct genealogical classes. We ran the software with the following parameters: a burn-in period of 20 000 generations and 200 000 MCMC, no previous population information, 'Jeffery's like priors', and the default six genealogical classes that correspond to: (i) Pure *E. aggregata*, (ii) Pure *E. rubida*, (iii) F<sub>1</sub> hybrid, (iv) F<sub>2</sub> hybrid, (v) Bx *aggregata* (backcross towards *E. aggregata*), (vi) Bx *rubida* (backcross towards *E. rubida*). As suggested by Vähä and Primmer (2006), a posterior probability of >50% was used as a threshold for assigning an individual to a specific class. For broader classification of individuals into each of the three groups: *E. rubida*, *E. aggregata* and hybrids; the posterior probabilities of all hybrid classes were summed for an individual.

We used simulations to assess the efficiency (proportion of individuals in a group that were correctly identified) and accuracy (proportion of an identified group that truly belong to that category), of STRUCTURE for identifying hybrids and NEWHYBRIDS for delineating the six genealogical classes. Simulated *E. aggregata*, *E. rubida*, first-generation (F<sub>1</sub>) and second-generation (F<sub>2</sub>) hybrids and backcrosses with each parental were obtained using the microsatellite genotypes of real adults and the software HYBRIDLAB (Nielsen *et al.*, 2006). As suggested by Vähä and Primmer (2006) we used reference parental genotypes of *E. aggregata* and *E. rubida* to obtain simulated genotypes. We used 40 *E. rubida* individual genotypes sampled from an allopatric population (Kings Hwy, Table 1 in Field *et al.*, 2009) and 40 *E. aggregata* individuals from sites where *E. aggregata* was numerically dominant and hybridization was not detected (Field *et al.*, 2009). Reference individuals were designated pure parentals only if initial runs with STRUCTURE indicated  $Q > 0.95$ . Simulated *E. aggregata* ( $N = 1000$ ) and *E. rubida* ( $N = 1000$ ), and each hybrid generation described above ( $N = 240$ ) were analyzed using STRUCTURE and NEWHYBRIDS under the same conditions as the real dataset.

**Table 1** Summary of genetic diversity measures and mean fixation ( $F_{IS}$ ) estimates in adult populations of *Eucalyptus aggregata*, *E. rubida* and their hybrids at each of three sites (Bendoura, Duck Flat, Norongo)

Species/Site	N	A	$H_E$ (SE)	$H_O$ (SE)	$F_{IS}$ (95% CI)
<i>E. aggregata</i>					
Bendoura	171	9.33	0.709 (0.019)	0.702 (0.019)	0.009 (−0.046, 0.064)
Duck Flat	18	4.66	0.648 (0.024)	0.711 (0.056)	−0.093 (−0.218, 0.032)
Norongo	10	5.00	0.648 (0.049)	0.733 (0.042)	−0.139 (−0.190, −0.087)
<i>Hybrids</i>					
Bendoura	47	14.2	0.855 (0.010)	0.823 (0.040)	0.038 (−0.048, 0.125)
Duck Flat	17	9.66	0.817 (0.018)	0.862 (0.060)	−0.055 (−0.191, 0.082)
Norongo	6	5.67	0.769 (0.023)	0.889 (0.070)	−0.153 (−0.304, −0.002)
<i>E. rubida</i>					
Bendoura	119	12.8	0.794 (0.027)	0.738 (0.057)	0.072 (−0.048, 0.188)
Duck Flat	24	9.50	0.750 (0.048)	0.736 (0.068)	0.025 (−0.049, 0.100)
Norongo	9	6.67	0.778 (0.018)	0.907 (0.053)	−0.169 (−0.307, −0.030)

Abbreviations: A, mean number of alleles per locus; CI, confidence interval;  $H_E$ , Hardy–Weinberg mean expected heterozygosity;  $H_O$ , observed heterozygosity; N, sample size; SE, standard error.

### Estimates of long-term gene flow

To estimate migration rates and assess the potential for asymmetry in gene flow between the two species and the hybrid populations, we used a coalescent-based approach implemented in the program MIGRATE-N version 3.0 (Beerli and Felsenstein, 1999, 2001). This analysis uses maximum-likelihood or Bayesian inference to estimate the parameter  $M$  ( $M = m/u$ , where  $m$  is the migration rate, and  $u$  is the mutation rate), which describes the mutation-scaled long-term migration rate between populations. MIGRATE-N also jointly estimates  $\Theta$  ( $\Theta = 4N_e u$ ), the mutation-scaled effective population size. Unlike  $F$ -statistics, this method allows for the possibility of asymmetrical gene flow between population pairs and differences in the effective population size.

For the purposes of assessing the rates of historical gene flow, individuals were assigned to one of three populations on the basis of admixture values implemented in STRUCTURE: *E. aggregata* ( $Q_1 > 0.9$ ), *E. rubida* ( $Q_1 < 0.1$ ) and hybrids ( $0.1 \leq Q_1 \leq 0.9$ ). For each site (Bendoura, Duck Flat and Norongo), we estimated  $\Theta$  for each population and  $M$  between all pair-wise populations ( $3 \times 3$ ) using the approaches implemented in MIGRATE-N. We conducted four identical maximum-likelihood analyses under both the step-wise mutational model with the Brownian motion model and the infinite-allele model. Search parameters consisted of 10 short chains with an increment of 20 and sampling of 500; five long chains with an increment of 20 and sampling of 10000; and a burn-in of 5000. With each run we changed the random number seed and the starting values of  $\Theta$  and  $M$ . The first run estimated  $\Theta$  and  $M$  from  $F_{ST}$  values, and subsequent runs used the maximum-likelihood estimates of  $\Theta$  and  $M$  from the previous run as starting parameters. We omitted locus Fmrsa1 from all analyses because of a high frequency of single base-pair mutations that do not adhere to the strict step-wise mutational model assumed by MIGRATE-N. Alleles occurring at moderate frequencies (>10%) at other loci also violated the strict step-wise mutational model and therefore we only report analysis using the infinite-allele model.

### Spatial clustering

We used  $O$ -ring statistics to assess the spatial patterns among hybrids and parental species at Bendoura and

Duck Flat. The Norongo site was excluded because of small sample size ( $N = 25$ ). We used Wiegand–Moloney's  $O$ -ring statistic (Wiegand and Moloney, 2004) to assess the spatial relationships of adults from three groups (*E. aggregata*,  $F_1$  hybrids and *E. rubida*) in two separate pair-wise analyses (*E. aggregata* vs hybrids, *E. rubida* vs hybrids). For each analysis, we calculated the bivariate  $O$ -ring statistic,  $O_{ij}(r)$ , from the number of individuals of group  $i$ , found in concentric rings of radius ( $r = 3.5$  m) about plants of group  $j$ . Values of  $O_{ij}(r) = 0$ ,  $O_{ij}(r) > 0$ ,  $O_{ij}(r) < 0$  correspond to spatial independence, spatial clustering and spatial repulsion, respectively. Tests for significance of spatial clustering against the null model (hybrids randomly dispersed around parent species) were performed by comparing the observed  $O_{ij}(r)$  statistic with 95% confidence envelopes from multiple Monte Carlo simulations ( $n = 1000$ ) at each radius. All calculations and simulations were conducted using the software PROGRAMITA (Wiegand and Moloney, 2004).

### Fine-scale spatial genetic structure (SGS)

SGS refers to the decrease of pair-wise relatedness among individuals with distance (that is, isolation-by-distance Wright, 1943). This can provide information regarding the scale of dispersal within populations and the potential for interspecific gene dispersal between sympatric species. To characterize SGS for *E. aggregata*, *E. rubida* and the hybrid populations at Bendoura and Duck Flat, we used the spatial autocorrelation procedures of Loiselle et al. (1995) and the software SPAGeDi version 1.2 (Hardy and Vekemans, 2002). Given the small sample size at Norongo ( $N = 25$ ), this site was excluded from the analysis. The average kinship coefficients ( $F_{ij}$ ) between pairs of individuals  $i$  and  $j$  were calculated for intervals of 15 m for distances between zero and 100 m, and intervals of 25 m for distances between 100 m and 450 m. Given the fragmented distribution of individuals at Duck Flat, here  $F_{ij}$  was estimated to a maximum of 150 m. The SGS pattern was visualized separately for each group by plotting the mean  $F_{ij}$  against distance. The significance of SGS at each distance interval was tested by permuting the spatial location of individuals

10 000 times to obtain 95% CIs about the null hypothesis of no SGS ( $F_{ij}=0$ ). We also calculated approximate standard errors (s.e.) for the mean  $F_{ij}$  at each distance interval by jackknifing over loci.

## Results

### Microsatellite diversity

Across all adults ( $N=421$ ) we detected a total of 105 alleles at six polymorphic loci. Among the adult populations, the widespread *E. rubida* tended to have the greater allelic diversity. A total of 77 alleles were recorded in *E. rubida* populations compared with 58 in the uncommon *E. aggregata*. The mean number of alleles per locus ( $A$ ) was 12.8 at Bendoura, 9.5 at Duck Flat and 6.67 at Norongo for *E. rubida*, compared with 9.33 at Bendoura, 4.66 at Duck Flat and 5.0 at Norongo for *E. aggregata* (Table 1). Despite the lower allelic diversity in *E. aggregata*, each adult had a unique multi-locus genotype. Genetic diversity estimates ( $H_E$ ) were also consistently higher in *E. rubida* (Bendoura:  $H_E=0.794$ , Duck Flat:  $H_E=0.750$ , Norongo:  $H_E=0.778$ ) than *E. aggregata* (Bendoura:  $H_E=0.709$ , Duck Flat:  $H_E=0.648$ , Norongo:  $H_E=0.648$ ). The hybrids displayed the highest levels of genetic diversity at the two larger sites but not at Norongo given the small sample size (Bendoura:  $H_E=0.855$ , Duck Flat:  $H_E=0.817$ , Norongo:  $H_E=0.769$ ) (Table 1).

### Wright's F-statistics

Mean fixation indices ( $F_{IS}$ ) were not significantly different from zero (random mating) for the *E. aggregata*, *E. rubida* or hybrid populations at the two largest sites Bendoura and Duck Flat. In contrast, significant departures from random mating were present for all three groups at the small remnant site Norongo, as  $F_{IS}$  was significantly  $<0$ . Therefore, at Norongo heterozygosity was higher than expected among the *E. aggregata* ( $F_{IS}=-0.139$ ), *E. rubida* ( $F_{IS}=-0.153$ ) and hybrid populations ( $F_{IS}=-0.169$ ) (Table 1).

Genetic differentiation between *E. aggregata* and *E. rubida* populations was moderate and significant as shown by Wright's  $F_{ST}$  at Bendoura ( $F_{ST}=0.209$ ), Duck Flat ( $F_{ST}=0.262$ ) and Norongo ( $F_{ST}=0.171$ ) (Table 2). This level of differentiation is consistent with the fact that allele frequencies at each locus were highly skewed towards either *E. aggregata* or *E. rubida*, rather than exhibiting fixed differences (data not shown). In contrast to interspecific genetic differentiation, the level of differentiation between *E. rubida* populations was very low (range  $F_{ST}$ ; 0.035–0.059). For the uncommon *E. aggregata*, genetic differentiation between populations was higher than between populations of the widespread *E. rubida* (range  $F_{ST}$ ; 0.067–0.119) (Table 2). Genetic differentiation between hybrids and *E. aggregata* ( $F_{ST}$ ; 0.019–0.065) was lower at each site compared with differentiation between hybrids and *E. rubida* ( $F_{ST}$ ; 0.082–0.119), but this was not significant as the 95% CIs overlapped (Table 2).

### Simulations

The admixture analysis performed with STRUCTURE had maximum accuracy when assigning individuals broadly to each of the three classes: *E. aggregata*,

**Table 2** Pair-wise tests for genetic differentiation (Wright's  $F_{ST}$ ) among adult populations of *E. aggregata*, *E. rubida* and their hybrids at three sites (Bendoura, Duck Flat, Norongo)

Pair-wise comparison	$F_{ST}$ (95% CI)
<i>Interspecific</i>	
agg (BT) vs rub (BT)	0.209 (0.170, 0.251)
agg (DF) vs rub (DF)	0.262 (0.209, 0.312)
agg (NG) vs rub (NG)	0.171 (0.117, 0.222)
<i>Intraspecific</i>	
agg (BT) vs agg (DF)	0.067 (0.032, 0.103)
agg (BT) vs agg (NG)	0.112 (0.080, 0.145)
agg (DF) vs agg (NG)	0.119 (0.070, 0.169)
rub (BT) vs rub (DF)	0.035 (0.022, 0.046)
rub (BT) vs rub (NG)	0.038 (0.013, 0.069)
rub (DF) vs rub (NG)	0.059 (0.034, 0.082)
<i>Hybrids</i>	
agg (BT) vs hyb (BT)	0.051 (0.033, 0.070)
agg (DF) vs hyb (DF)	0.065 (0.031, 0.110)
agg (NG) vs hyb (NG)	0.019 (0.016, 0.051)
rub (BT) vs hyb (BT)	0.082 (0.057, 0.107)
rub (DF) vs hyb (DF)	0.105 (0.062, 0.153)
rub (NG) vs hyb (NG)	0.119 (0.070, 0.169)

Abbreviations: agg, *Eucalyptus aggregata*; rub, *E. rubida*; hyb, hybrids; BT, site Bendoura; DF, site Duck Flat; NG, site Norongo; CI, Confidence interval.

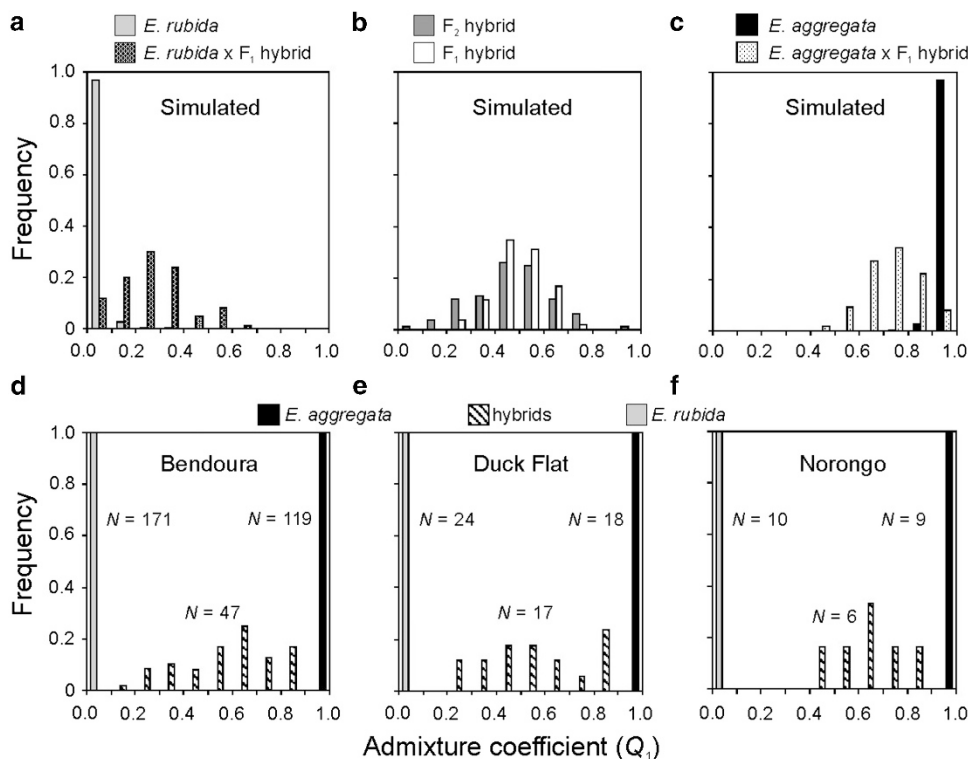
*E. rubida* and hybrids. With these classes, a threshold of  $Q=0.9$  had the highest efficiency and accuracy for distinguishing hybrids from purebreds (Supplementary Figure S1). At this threshold, the number of apparent hybrids in the simulated population (Mean 7.1%) was the closest to the actual number of hybrids (7.4%). Most misidentified hybrids were backcrosses, with  $\sim 10\%$  of this group assigned incorrectly to a purebred class because of substantial overlap in admixture values (Figures 2a–c).

The assignment of simulated individuals using NEWHYBRIDS was mostly congruent with those obtained from STRUCTURE. All simulated purebred individuals were assigned with high posterior probability ( $q_i > 95\%$ ) to the same purebred category as STRUCTURE (Figure 3a). This analysis was most successful in identifying early generation hybrids. Using an assignment threshold of  $q_i > 50\%$ , 93% of  $F_1$  hybrids were assigned to the correct class. NEWHYBRIDS performed poorly with the assignment of more advanced generation hybrids. A threshold of  $q_i > 50\%$  had the highest efficiency and accuracy for assignment of simulated backcrosses with 75% of Bx *aggregata* and 77% of Bx *rubida* classes correctly assigned (Supplementary Figure S2). The majority of the misidentified backcrosses were mostly assigned as the parent species involved in the backcross.

### Frequency of hybridization

Analyses of admixture values calculated in STRUCTURE indicate that hybrids between *E. aggregata* and *E. rubida* are common, with 69 (16%) individuals displaying hybrid ancestry (that is,  $0.1 \leq Q_1 \leq 0.9$ ). The highest number of hybrids detected was at the fragmented site Duck Flat, where 17 (28.8%) of the trees were determined to be hybrids (Figure 2e). In comparison, 47 (13.9%) and 6 (24.0%) of the trees were identified as hybrids at





**Figure 2** Frequency distribution of admixture coefficients ( $Q_1$ ) of multilocus genotypes of simulated parental and hybrid individuals (a–c), and adult plants at each of three sites: (d) Bendoura, (e) Duck Flat and (f) Noronogo. Adult plant assignment based on the following thresholds:  $Q_1 > 0.9$  as *E. aggregata*,  $Q_1 < 0.1$  as *E. rubida* and  $0.1 \leq Q_1 \leq 0.9$  as hybrids.

Bendoura and Noronogo, respectively (Figures 2d and f). When restricted to three classes (*E. aggregata*, *E. rubida*, hybrids), the NEWHYBRIDS analyses assigned individuals to the same classes at Bendoura and Duck Flat as the STRUCTURE analysis. In contrast, results from NEWHYBRIDS indicate that 10 (40%) of the trees at Noronogo were hybrids. This was because four individuals assigned with STRUCTURE to *E. rubida* ( $Q_1 < 0.1$ ) exhibited above 80% probability of hybrid assignment when summed across all hybrid classes (Figure 3d). The discrepancy between the techniques for the four individuals at this site is probably because of the lower genetic differentiation between parental species at Noronogo (Table 2) and the subsequent reduced power in distinguishing backcrosses from parentals at this site.

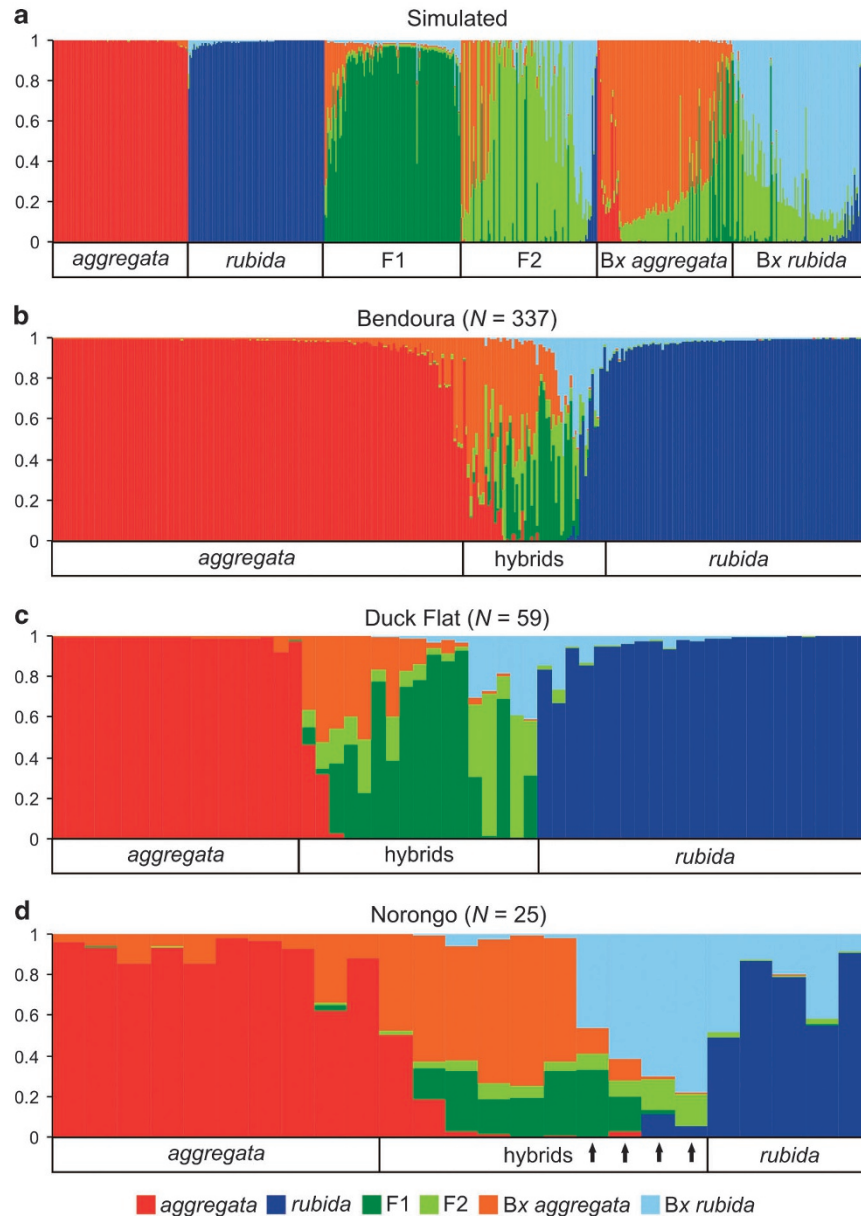
#### Nature of introgression

Assignment probabilities for genealogical classes (NEWHYBRIDS) indicate admixed individuals belong to a range of hybrid generations; however, the number of introgressed individuals is likely to be underestimated with these markers (see simulations above). Among the hybrid individuals, 20 of 47 (42.6%) at Bendoura (Figure 3b), 3 of 17 (17.6%) at Duck Flat (Figure 3c) and 6 of 10 (60%) at Noronogo (Figure 3d) displayed high probabilities (>50%) of belonging to a backcrossed generation. A skewed distribution in the admixture coefficients towards *E. aggregata* (STRUCTURE) suggests a greater genetic input from *E. aggregata* in the hybrid populations. We found 50 (72%) of the hybrids showed admixture coefficients closer to the *E. aggregata* cluster

( $0.5 < Q_1 < 0.9$ ). This bias among the hybrid populations was evident at all three sites, with 35 (74%) at Bendoura (Figure 2d), 10 (58%) at Duck Flat (Figure 2e), and 5 (83%) at Noronogo (Figure 2f) exhibiting admixture coefficients between 0.5 and 0.9. Similarly, the mean admixture values of hybrid populations ( $0.1 < Q_1 < 0.9$ ) were skewed towards the *E. aggregata* cluster at Bendoura ( $Q_1 \bar{x} = 0.59 \pm 0.03$ ), Duck Flat ( $Q_1 \bar{x} = 0.56 \pm 0.05$ ) and Noronogo ( $Q_1 \bar{x} = 0.65 \pm 0.05$ ).

#### Long-term estimates of gene flow

The maximum likelihood estimates of long-term gene flow using MIGRATE-N indicated low levels of migration between the two species. The migration rate  $M$  was  $< 0.8$  (Figure 4), suggesting low levels of historic gene exchange. There was also no evidence of asymmetrical gene flow between the two species (Figure 4) because of overlapping CIs for estimates of  $M$  in both directions at all three sites (for example, Bendoura,  $M_{E. aggregata \rightarrow E. rubida} = 0.68$ , 95% CIs = (0.49, 0.93);  $M_{E. rubida \rightarrow E. aggregata} = 0.37$  (0.24, 0.53)). However, there was evidence that *E. aggregata* has a greater genetic input into the hybrid populations, with more gene flow detected from the *E. aggregata* to hybrid populations compared with *E. rubida* to the hybrid populations. Higher rates of gene flow from the *E. aggregata* into the hybrid population was most evident at Bendoura ( $M_{E. aggregata \rightarrow hybrids} = 7.86$ , (6.96, 8.83);  $M_{E. rubida \rightarrow hybrids} = 4.65$  (3.95, 5.42)) (Figure 4a), and is consistent with the prediction of greater introgression towards the smaller-flowered species. Whereas higher maximum-



**Figure 3** Bayesian assignment of multilocus genotypes of simulated purebred and hybrid individuals (a) and real adults ( $N = 421$ ) from three sympatric populations (b–d) using the software NEWHYBRIDS. Each line represents an individual's posterior probability of assignment ( $q_i$ ) to each of six genealogical classes including: purebred *E. aggregata*, purebred *E. rubida* and four hybrid classes ( $F_1$ ,  $F_2$ , backcrosses *E. rubida* towards each parent (Bx *aggregata*, Bx *rubida*)). Simulated individuals for each of the six classes were generated using reference genotypes and HYBRIDLAB. Arrows indicate individuals assigned as purebred *E. rubida* using the software STRUCTURE.

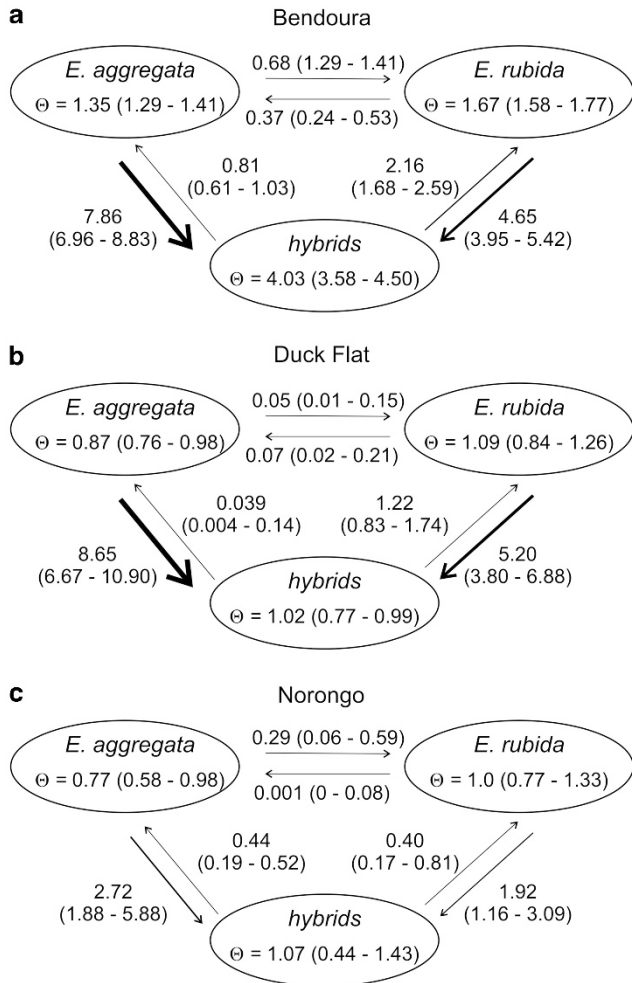
likelihood estimates of gene flow from *E. aggregata* into the hybrid population was evident at the remaining sites, the 95% CIs marginally overlapped at both Duck Flat ( $M_{E. aggregata \rightarrow hybrids} = 8.65, (6.67, 10.90)$ ;  $M_{E. rubida \rightarrow hybrids} = 5.20 (3.80, 6.88)$ ) (Figure 4b) and Norongo ( $M_{E. aggregata \rightarrow hybrids} = 2.72, (1.88, 5.88)$ ;  $M_{E. rubida \rightarrow hybrids} = 1.92 (1.16, 3.09)$ ) (Figure 4c).

#### Spatial clustering

Bivariate *O*-ring analyses of the spatial distributions of individuals indicated significant spatial clustering between hybrids and both parental species at short distance

intervals between 5 and 20 m (Figure 5). At greater distance intervals, there was little spatial clustering between these groups because the  $O_{12}(r)$  statistic was within the simulated 95% CIs (randomly dispersed). At short distance intervals, higher levels of the  $O_{12}(r)$  for *E. aggregata* vs hybrids comparison suggest hybrids are more spatially clustered with *E. aggregata* than *E. rubida*. At Bendoura, the higher spatial clustering between hybrids and *E. aggregata* was marginal and evident only at the smallest distance interval of 5 m (Figure 5). However, at Duck Flat, higher levels of clustering between hybrids and *E. aggregata* compared with hybrids and *E. rubida* persisted from 5 to 20 m (Figure 5).





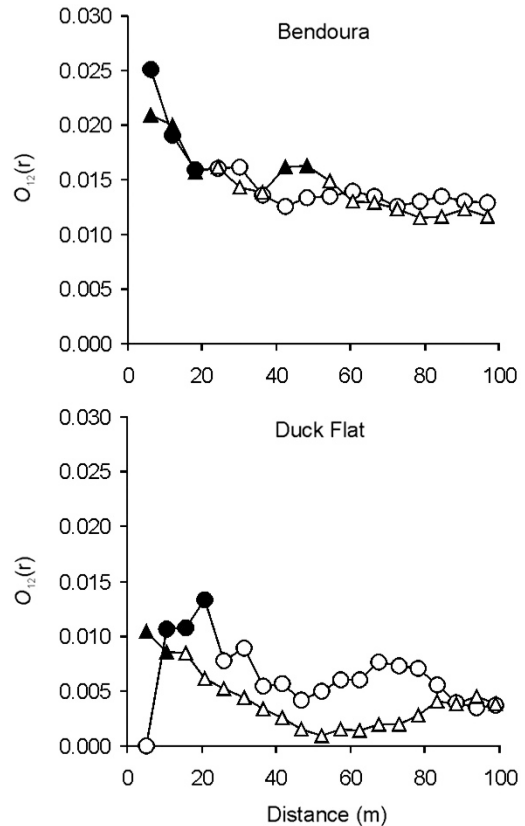
**Figure 4** Maximum-likelihood estimates of the long-term migration rates ( $M$ ) and the mutation-scaled effective population size ( $\Theta$ ) in each of three *Eucalyptus* hybrid zones (a–c) using the methods of MIGRATE-N. Thickness of the arrows indicates the directional migration rate estimates ( $M = m/u$ , where  $m$  is the migration rate, and  $u$  is the mutation rate). Values in parenthesis indicate the 95% CIs.

### Fine-scale SGS

Fine-scale genetic structure was low to moderate in *E. aggregata* and *E. rubida* populations, with the mean kinship coefficient ( $F_{ij}$ ) significantly different from zero at several of the short distance classes (Figure 6). The  $F_{ij}$  estimates were significant and positive at smaller distance intervals of <45 m for *E. aggregata* at Bendoura and Duck Flat and *E. rubida* at Bendoura. In comparison, the range of SGS was less for *E. rubida* at Duck Flat, because  $F_{ij}$  was only significant and positive at distances <30 m. For the hybrid populations, significant SGS was only detected at Bendoura from 15 to 30 m ( $F_{ij} = 0.058$ , 95% CIs = (0.033, 0.085)) (Figure 6).

### Discussion

The patterns of interspecific gene flow among species are important for understanding the processes that maintain distinct species. In this study, molecular evidence in each of the three hybrid zones of *E. rubida* and *E. aggregata* indicate that hybridization is frequent (14–40% of the

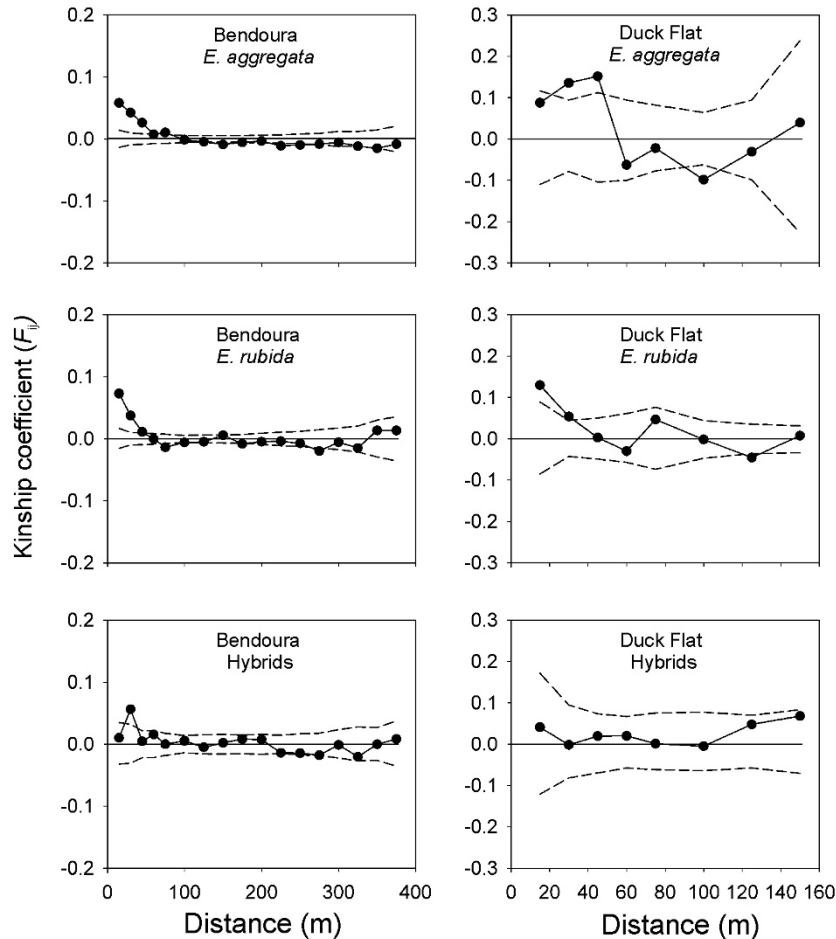


**Figure 5** Wiegand and Moloney's  $O_{12}(r)$  statistics for bivariate analyses of adult populations of *E. aggregata*, *E. rubida* and hybrids at the continuous site Bendoura and remnant site Duck Flat. Circles and triangles represent bivariate analysis for *E. aggregata* vs hybrids and *E. rubida* vs hybrid individuals, respectively. Filled symbols indicate values significantly different from the null hypothesis as they lie outside the 95% CIs. The 95% CIs were generated from 1000 Monte Carlo simulations of hybrid individuals randomly distributed around fixed *E. aggregata* and *E. rubida* distributions.

adults), and introgression has occurred (5.1–24% of adults) with backcrosses in both directions. These results support previous molecular and morphological evidence of both  $F_1$  and backcross hybrids among progeny arrays collected from *E. aggregata* parents (Field et al., 2009). In addition, further genealogical assessments and measures of historical gene flow revealed greater genetic input from *E. aggregata* into the hybrid populations. These results fit our prediction that interspecific gene flow would be asymmetrical and directed towards the smaller flowered *E. aggregata*.

### Extent of introgression

Detection of backcross hybrids and evidence of historic introgression suggest hybrid zones between *E. aggregata* and *E. rubida* are complex, and are not limited to the first generation. Hybridization beyond the  $F_1$  generation is commonly reported in plants, with many examples of advanced generation hybrids outnumbering early generations in hybrid populations (see review; Rieseberg and Carney, 1998). This is because bridging the initial gap between two species through the formation of  $F_1$  hybrids is generally considered the more difficult step (Manel et al., 2005). In *Eucalyptus*, asynchronous flowering



**Figure 6** Spatial autocorrelation analyses of pair-wise co-ancestry ( $F_{ij}$ ) estimated from multi-locus genotypes of *Eucalyptus aggregata*, *E. rubida* and hybrids at each site (Bendoura and Duck Flat). Dashed lines indicate upper and lower 95% CIs around the null hypothesis of  $F_{ij}=0$ .

is a strong reproductive barrier, with limited reports of hybridization among species with completely divergent flowering times (Potts *et al.*, 2003). In our system,  $F_1$  hybrids produce viable seed and exhibit intermediate flowering times between the parental species (Field, 2008). Given the greater flowering synchrony between established  $F_1$  hybrids and parentals, early-generation hybrids may provide a bridge for the movement of genes between *E. aggregata* and *E. rubida*.

Frequent hybridization and introgression in adult populations (present study) and in seedling cohorts (Field *et al.*, 2008) suggest that *E. aggregata* and *E. rubida* have maintained distinct morphological traits and habitat associations despite only partial reproductive isolation. This could reflect recent adaptive radiation of these species in the presence of ongoing gene flow, considering the low to moderate level of genetic differentiation between *E. aggregata* and *E. rubida* ( $F_{ST}=0.17-0.26$ ). There is a growing body of evidence of widespread introgression of cytoplasmic and nuclear DNA among distinct plant species (Mallet, 2005), suggesting that the speciation process can occur despite frequent and ongoing interspecific-gene flow (Lexer and Widmer, 2008). Fine-scale estimates of the frequency of introgressed individuals within mixed populations have varied widely among species pairs, from cases of low

frequencies (0.01%) of advanced generation recombinants in *Liparis* (Chung *et al.*, 2005) to moderate frequencies (14%) between the rare *E. cordata* and the widespread *E. globulus* (McKinnon *et al.*, 2010). In the latter study, McKinnon *et al.*, 2010 also found low levels of genetic differentiation between the rare and common species in mixed populations ( $F_{ST}=0.208$ ) despite distinct morphological differences, habitat preferences and assignment to different series within section *Maide-naria* (series *Obiculares* and *Globulares*, respectively). Evidence of historical introgression among *Eucalyptus* species (McKinnon *et al.*, 2001) and recent introgression in mixed populations of *E. aggregata* and *E. rubida* suggest frequent opportunities exist for gene exchange between distinct *Eucalyptus* species.

An important aspect in detecting introgression using nuclear DNA markers is being able to distinguish early-generation hybrids from backcrosses, with only the latter potentially resulting in the transfer of genetic material between species. In our study, simulations indicate that six microsatellite markers provide low accuracy (75–77%) in distinguishing backcross hybrids. This highlights an important limitation of using low numbers of codominant markers, as the power to accurately distinguish hybrid classes is dependent on both the number of loci and the level of genetic differentiation

between the parentals. Vähä and Primmer (2006) found that a relatively high degree of accuracy (95%) in detecting backcross hybrids might require >48 codominant loci and high genetic divergence ( $F_{ST} = 0.21$ ). These requirements are often not practical in population studies of natural hybrid zones and are much higher than the number of loci commonly reported with codominant markers such as microsatellites (for example, Curtu *et al.*, 2009; Valbuena-Carabana *et al.*, 2007). Larger numbers of loci typically obtained using alternative techniques such as amplified fragment length polymorphism, may be the best strategy for detecting advanced hybrid generations with a high degree of confidence.

### Direction of Introgression

Given the limitations in accurately detecting hybrid classes, what can we infer about the direction of introgression between *E. aggregata* and *E. rubida*? Several characteristics of the genomic composition of the hybrid populations and long-term estimates of gene flow suggest a higher contribution of genetic material from *E. aggregata*. First, lower genetic differentiation between hybrids and the uncommon *E. aggregata*, despite broad CIs, indicates gene flow between these populations is more frequent than between hybrids and the widespread *E. rubida*. This finding is consistent with long-term gene flow estimates from MIGRATE-N. Despite an overall low level of migration ( $M$ ) between the purebred populations ( $M < 1.0$ ), we found evidence of asymmetrical gene exchange, with higher levels of migration between the hybrid populations and *E. aggregata* compared with *E. rubida* (Bendoura; Figure 4a). Although migration rates were not significantly asymmetrical at the remaining sites, broad CIs because of smaller sample sizes may have obscured the same underlying trend (Figures 4b and c). Contemporary approaches using NEWHYBRIDS analyses found a similar pattern, with more individuals assigned to the Bx aggregata class in each of the hybrid populations (Figure 3). This trend is also consistent with the results from STRUCTURE, with admixture values skewed towards *E. aggregata* and few hybrids exhibiting admixture values close to the *E. rubida* genetic cluster ( $Q_1$  0.05 to 0.375). Although this pattern could be a sampling artefact, the consistent trend across all three sites using contemporary approaches suggests backcrossing is probably more frequent in the direction of *E. aggregata*. This is in comparison with a dominance of  $F_1$  hybrids, which would suggest limited introgression. The complex genomic composition of the hybrid populations suggests a high potential for gene movement between *E. aggregata* and *E. rubida* through the hybrid populations.

### Causes of asymmetric introgression

The greater degree of backcrossing towards *E. aggregata* observed within the three sympatric populations could be explained by several pre-zygotic and post-zygotic mechanisms. In natural plant populations, selection remains the primary mechanism implicated in determining patterns of hybridization and introgression (Lexer *et al.*, 2005; Whitney *et al.*, 2006). In this case, *E. aggregata*-like hybrids may possess a selective advantage over *E. rubida*-like hybrids in these mixed habitats. Alternatively, asymmetric

introgression could reflect demographic processes related to species range expansion, which has been implicated as a major determinant of gene flow patterns in a wide array of plants and animals (Currat *et al.*, 2008). Using simulations, Currat *et al.* (2008) demonstrated that range expansion can result in asymmetric introgression of neutral alleles from the local to the invading species. Under this model, expansion of the widespread *E. rubida* into *E. aggregata* habitat is predicted to result in more introgression of local *E. aggregata* alleles into the invading *E. rubida* and subsequent hybrid population, a pattern consistent with the higher frequency of introgression detected between *E. aggregata* and the hybrid populations at Bendoura. Although selection and demographic processes may have a role in determining the patterns observed in this system, the presence of unilateral pre-zygotic barriers may also contribute towards limiting the success of backcrossing in the direction of *E. rubida*. The most likely mechanism is a difference in flower size between the parental species. In *Eucalyptus*, this has been found to impede pollen tubes of small-flowered species from reaching the ovule of large-flowered species (Gore *et al.*, 1990). This structural barrier may be reduced in  $F_1$  hybrids as they tend to exhibit intermediate flower size (Field, 2008). In this case, the presence of  $F_1$  hybrids may therefore reduce the strength of asymmetrical barriers to gene flow.

Interspecific gene flow between hybrids and *E. rubida* may also be impeded by the non-random distribution of individuals. At Bendoura and Duck Flat, O-ring analyses indicated that hybrids are more spatially clustered with *E. aggregata* compared with *E. rubida*. This spatial pattern could be because *E. aggregata* is more often the maternal parent of hybrid offspring combined with localized seed dispersal and recruitment of hybrids. Consistent with this prediction, our analyses of fine-scale SGS indicate seed dispersal and recruitment is restricted to <40 m around maternal plants. *Eucalyptus* seed are small (*E. aggregata*, 1–1.2 mm) and not adapted to wind dispersal, resulting in generally low dispersal distances (House, 1997). The seed dispersal radius inferred from the SGS analyses (<40 m), is consistent with direct measures of seed dispersal in similarly sized *Eucalyptus* (*E. marginata*; Cremer, 1977). Considering that plant mating is typically more frequent among nearest neighbors (for example, García *et al.*, 2005; Stacy *et al.*, 1996), this spatial pattern may further enhance positive assortative mating among hybrids and *E. aggregata* plants. Our data demonstrates that an initial bias in the maternal parent involved in  $F_1$ -hybrid formation, coupled with localised pollen and seed dispersal, may be important contributing factors in determining the direction of backcrossing. This indicates that the local distribution of hybrids with respect to the parental species may be an important and under-appreciated process for reinforcing asymmetrical introgression in plant populations.

The complex genetic composition of hybrids and their persistence through a range of age cohorts (seed to adult populations) (Field *et al.*, 2008), indicates that hybridization and introgression is a dynamic and ongoing process between *E. aggregata* and *E. rubida*. In this system, there are no differences in early hybrid fitness relative to purebreds (Field, 2008), but in general *Eucalyptus* hybrids tend to exhibit poorer fitness at later life-history stages



(Lopez *et al.*, 2000). Given that these species are associated with different habitats, selection may be strong against intermediate-hybrid genotypes that are less well adapted than the parental species in either local environment. It is therefore likely that selection, demography and floral architecture are contributing factors to the patterns of hybridization and introgression in this system.

## Conflict of interest

The authors declare no conflict of interest.

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## References

- Anderson EC, Thompson EA (2002). A model-based method for identifying species hybrids using multilocus genetic data. *Genetics* **160**: 1217–1229.
- Bacilieri R, Ducouso A, Petit R, Kremer A (1996). Mating system and asymmetric hybridization in a mixed stand of European oaks. *Evolution* **50**: 900–908.
- Beehler P, Felsenstein J (1999). Maximum-likelihood estimation of migration rates and effective population numbers in two populations using a coalescent approach. *Genetics* **152**: 763–773.
- Beehler P, Felsenstein J (2001). Maximum likelihood estimation of a migration matrix and effective population sizes in *n* subpopulations by using a coalescent approach. *Proc Natl Acad Sci USA* **98**: 4563–4568.
- Brondani RPV, Brondani C, Tarchini R, Grattapaglia D (1998). Development, characterization and mapping of microsatellite markers in *Eucalyptus grandis* and *E urophylla*. *Theor Appl Genet* **97**: 816–827.
- Brooker MIH (2000). A new classification of the genus *Eucalyptus* L'Hér. (Myrtaceae). *Aust Syst Bot* **13**: 79–148.
- Brooker MIH, Kleinig DA (1999). *Field Guide to Eucalyptus South-Eastern Australia*, 2nd edn. Bloomings Books: Hawthorn.
- Burgess KS, Morgan M, Deverno L, Husband BC (2005). Asymmetrical introgression between two *Morus* species (*M. alba*, *M. rubra*) that differ in abundance. *Mol Ecol* **14**: 3471–3483.
- Byrne M, Marquez-Garcia MI, Uren T, Smith DS, Moran GF (1996). Conservation and genetic diversity of microsatellite loci in the genus *Eucalyptus*. *Aust J Bot* **44**: 331–341.
- Cayzer LW (1993). The investigation of phenotypic variation within and between three Eucalypt taxa occurring in the ACT. Honours Thesis, Australian National University.
- Chung MY, Nason JD, Chung MG (2005). Patterns of hybridization and population genetic structure in the terrestrial orchids *Liparis kumokiri* and *Liparis makinoana* (Orchidaceae) in sympatric populations. *Mol Ecol* **14**: 4389–4402.
- Cremer KW (1977). Distance of seed dispersal in eucalypts estimated from seed weights. *Aust Forest Res* **7**: 225–228.
- Curat M, Ruedi M, Petit RJ, Excoffier L (2008). The hidden side of invasions: massive introgression by local genes. *Evolution* **62**: 1908–1920.
- Curtu AL, Gailing O, Finkeldey R (2009). Patterns of contemporary hybridization inferred from paternity analysis in a four-oak-species forest. *BMC Evol Biol* **9**: 9.
- Evanno G, Regnaut S, Goudet J (2005). Detecting the number of clusters of individuals using the software structure: a simulation study. *Mol Ecol* **14**: 2611–2620.
- Falush D, Stephens M, Pritchard JK (2003). Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics* **164**: 1567–1587.
- Field DL (2008). The importance of ecological factors in determining the pattern of interspecific hybridisation in fragmented landscapes of *Eucalyptus aggregata*, PhD Thesis, University of Wollongong.
- Field DL, Ayre DJ, Whelan RJ, Young AG (2008). Relative frequency of sympatric species influences rates of interspecific hybridization, seed production, and seedling performance in the uncommon *Eucalyptus aggregata*. *J Ecol* **96**: 1198–1210.
- Field DL, Ayre DJ, Whelan RJ, Young AG (2009). Molecular and morphological evidence of natural interspecific hybridization between the uncommon *Eucalyptus aggregata* and the widespread *E rubida* and *E viminalis*. *Conserv Genet* **10**: 881–896.
- García C, Arroy JM, Godoy JA, Jordano P (2005). Mating patterns, pollen dispersal, and the ecological maternal neighbourhood in a *Prunus mahaleb* L population. *Mol Ecol* **14**: 1821–1830.
- Gore P, Potts B, Volker P, Megalos J (1990). Unilateral cross-incompatibility in *Eucalyptus*: the case of hybridisation between *E globulus* and *E. nitens*. *Aust J Bot* **38**: 383–394.
- Goudet J (1995). FSTAT (Version 1.2): A computer program to calculate F-statistics. *J Hered* **86**: 485–486.
- Grant V (1981). *Plant Speciation*. Columbia University Press: New York.
- Griffin AR, Burgess IP, Wolf L (1988). Patterns of natural and manipulated hybridization in the genus *Eucalyptus*. *Aust J Bot* **36**: 41–66.
- Hardy OJ, Vekemans X (2002). SPAGEDi: a versatile computer program to analyse spatial genetic structure at the individual or population levels. *Mol Ecol Notes* **2**: 618–620.
- House SM (1997). Reproductive biology of eucalypts. In: Williams J, Woinarski J (eds) *Eucalypt Ecology: Individuals to Ecosystems*. Cambridge University Press: Cambridge, pp 30–55.
- Lewis D, Crowe LK (1958). Unilateral interspecific incompatibility in flowering plants. *Heredity* **12**: 233–256.
- Lexer C, Fay MF, Joseph JA, Nica MS, Heinze B (2005). Barrier to gene flow between two ecologically divergent *Populus* species, *P. alba* (white poplar) and *P. tremula* (European aspen): the role of ecology and life history in gene introgression. *Mol Ecol* **14**: 1045–1057.
- Lexer C, Widmer A (2008). The genic view of plant speciation: recent progress and emerging questions. *Philos Trans R Soc B Biol Sci* **363**: 3023–3036.
- Loiselle BA, Sork VL, Nason J, Graham C (1995). Spatial genetic structure of a tropical understory shrub, *Psychotria officinalis* (Rubiaceae). *Am J Bot* **82**: 1420–1425.
- Lopez GA, Potts BM, Tilyard PA (2000). F<sub>1</sub> hybrid inviability in *Eucalyptus*: the case of *E ovata* x *E. globulus*. *Heredity* **85**: 242–250.
- Mallet J (2005). Hybridization as an invasion of the genome. *Trends Ecol Evol* **20**: 229–237.
- Manel S, Oscar EG, Waples RS (2005). Assignment methods: matching biological questions with appropriate techniques. *Trends Ecol Evol* **20**: 136–142.
- McKinnon GE, Smith JJ, Potts BM (2010). Recurrent nuclear DNA introgression accompanies chloroplast DNA exchange between two eucalypt species. *Mol Ecol* **19**: 1367–1380.
- McKinnon GE, Vaillancourt RE, Jackson HD, Potts BM (2001). Chloroplast sharing in the tasmanian *Eucalyptus*. *Evolution* **55**: 703–711.
- Nielsen EE, Bach LA, Kotlicki P (2006). HYBRIDLAB (version 1.0): a program for generating simulated hybrids from population samples. *Mol Ecol Notes* **6**: 971–973.
- Peakall R, Smouse PE (2005). *GenAlEx 6: Genetic Analysis in Excel. Population Genetic Software for Teaching and Research*. Australian National University: Canberra, Australia.

- Petit RJ, Bodénès C, Ducousso A, Roussel G, Kremer A (2004). Hybridization as a mechanism of invasion in oaks. *New Phytol* **161**: 151–164.
- Potts BM, Barbour RC, Hingston AB, Vaillancourt RE (2003). Genetic pollution of native eucalypt gene pools—identifying the risks. *Aust J Bot* **51**: 1–25.
- Pritchard JK, Stephens M, Donnelly P (2000). Inference of population structure using multilocus genotype data. *Genetics* **155**: 945–959.
- Rhymer JM, Simberloff D (1996). Extinction by hybridization and introgression. *Annu Rev Ecol Syst* **27**: 83–109.
- Rieseberg LH, Carney SE (1998). Plant hybridization. *New Phytol* **140**: 599–624.
- Stacy EA, Hamrick JL, Nason JD, Hubbell SP, Foster RB, Condit R (1996). Pollen dispersal in low-density populations of three neotropical tree species. *Am Naturalist* **148**: 275–298.
- Steane DA, Vaillancourt RE, Russel J, Powell W, Marshall D, Potts BM (2001). Development and characterisation of microsatellite loci in *Eucalyptus globulus* (Myrtaceae). *Silv Genet* **50**: 89–91.
- Tiffin P, Olson MS, Moyle MC (2001). Asymmetrical crossing barriers in angiosperms. *Proc R Soc B* **268**: 861–867.
- Vähä JP, Primmer CR (2006). Efficiency of model-based Bayesian methods for detecting hybrid individuals under different hybridization scenarios and with different numbers of loci. *Mol Ecol* **15**: 63–72.
- Valbuena-Carabana M, Gonzalez-Martinez SC, Hardy OJ, Gil L (2007). Fine-scale spatial genetic structure in mixed oak stands with different levels of hybridization. *Mol Ecol* **16**: 1207–1219.
- van der Nest MA, Steenkamp ET, Wingfield BD, Wingfield MJ (2000). Development of simple sequence repeat (SSR) markers in *Eucalyptus* from amplified inter-simple sequence repeats (ISSR). *Plant Breeding* **119**: 433–436.
- van Oosterhout C, Hutchinson WF, Wills DPM, Shipley P (2004). micro-checker: software for identifying and correcting genotyping errors in microsatellite data. *Mol Ecol Notes* **4**: 535–538.
- Whitney KD, Randell RA, Rieseberg LH (2006). Adaptive introgression of herbivore resistance traits in the weedy sunflower *Helianthus annuus*. *Am Naturalist* **167**: 794–807.
- Wiegand T, Moloney KA (2004). Rings, circles, and null-models for point pattern analysis in ecology. *Oikos* **104**: 209–229.
- Williams EG, Rouse JL (1988). Disparate style lengths contribute to isolation of species in *Rhododendron*. *Aust J Botany* **36**: 183–191.
- Wright S (1943). Isolation by distance. *Genetics* **28**: 114–138.
- Wright S (1965). The interpretation of population structure by *F*-statistics with special regard to systems of mating. *Evolution* **19**: 395–420.

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