

Tetrasomic inheritance and isozyme variation in *Turnera ulmifolia* vars. *elegans* Urb. and *intermedia* Urb. (Turneraceae)

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Tetrasomic inheritance of three isozyme loci is demonstrated in *Turnera ulmifolia* vars. *elegans* and *intermedia*. The data support the occurrence of an autopolyploid origin for tetraploids of both taxonomic varieties. The extent of isozyme variation at 14 loci was determined for diploid and tetraploid populations. Tetraploid *Turnera ulmifolia* var. *intermedia* showed the lowest levels of isozyme variation, perhaps a result of founder effect, upon island colonization. In contrast, *Turnera ulmifolia* var. *elegans* showed the greatest levels of isozyme variation, suggesting that hybridization among locally differentiated diploid or tetraploid populations might have occurred. The number of independent origins of autotetraploidy in *Turnera ulmifolia* is unknown.

Keywords: isozyme variation, tetrasomic inheritance.

Introduction

Turnera ulmifolia is a species complex of perennial weeds native to the Neotropics. The complex contains diploid, tetraploid, hexaploid and octoploid cytotypes and has a base number of $x=5$ (Raman & Kesavan, 1964; Barrett, 1978; Arbo & Fernandez, 1983; Shore & Barrett, 1985a; Barrett & Shore, 1987; Fernandez, 1987; Fernandez & Arbo, 1989). Diploid and tetraploid populations are distylous and individuals are strongly self-incompatible and obligately outcross, while hexaploids of three taxonomic varieties are homostylous and self-compatible (Barrett, 1978; Shore & Barrett, 1985b; Barrett & Shore, 1987).

Turnera ulmifolia vars. *elegans* and *intermedia* are highly interfertile members of the species complex (Shore & Barrett, 1985a; Arbo & Fernandez, 1987; Shore, 1991) and diploid hybrids show regular bivalent formation at meiosis (Fernandez & Arbo, 1989). The varieties are most easily distinguished by the presence of a purple petal spot in the cream-coloured corollas of *T. ulmifolia* var. *elegans* while var. *intermedia* has yellow flowers that lack the petal spot. Diploid populations of *T. ulmifolia* var. *intermedia* occur throughout Central America and in portions of South America, while tetraploid populations are apparently restricted to Puerto Rico and Hispaniola (Barrett & Shore, 1987). Native populations of diploid and tetraploid *T.*

ulmifolia var. *elegans* occur in Brazil. A tetraploid population that shares characteristics of both varieties occurs in Columbia (Barrett, 1978; Shore & Barrett, 1985a), but insufficient numbers of plants were available for study.

Three lines of evidence suggest that tetraploid populations of *T. ulmifolia* vars. *elegans* and *intermedia* are autopolyploids. Cytological observations of tetraploids of both varieties have revealed the occurrence of quadrivalents at diakinesis and the first metaphase of meiosis, as well as reduced pollen fertility (Raman & Kesavan, 1964; Arbo & Fernandez, 1983; Fernandez, 1987; Shore, 1991). Finally, frequencies of meiotic configurations fit the autotetraploid pairing models of Jackson & Hauber (1982) for four of six populations investigated (Shore, 1991).

In this paper, the hypothesis that *T. ulmifolia* vars. *elegans* and *intermedia* have had autotetraploid origins is tested and the genetic consequences of autotetraploidy is investigated. Genetic evidence is provided for autopolyploidy in these taxonomic varieties by demonstrating tetrasomic inheritance at three isozyme loci. To investigate the genetic consequences of autotetraploidy, levels of isozyme variation in diploid and tetraploid populations of *T. ulmifolia* were compared. Finally, the evidence bearing on the evolutionary origin of the tetraploid varieties was examined.

Materials and methods

Bulk seed samples were collected from populations found throughout much of the native range of *T. ulmifolia* vars. *intermedia* and *elegans*. Seeds were germinated and population samples maintained in a glasshouse following the methods of Shore & Barrett (1985a). The varietal status, sample size and ploidal level of populations sampled is provided in Table 1.

Seven populations of tetraploid *T. ulmifolia* var. *elegans* were sampled from Brazil. For *T. ulmifolia* var. *intermedia*, seven diploid populations were sampled from Central and South America and 16 tetraploid populations were sampled from Puerto Rico and the

Table 1 Varietal status, population code, locality, ploidal level and sample size (*n*) of *Turnera ulmifolia* populations used

Population	Locality	Ploidal level	<i>n</i>
I1	Barreirinhas, Brazil	2x	18
I2	Calabozo, Venezuela	2x	12
I26	Santa Rosa, Costa Rica	2x	28
I28	Santa Rosa, Costa Rica	2x	8
I29	La Pacifica, Costa Rica	2x	26
I31	Juremal, Brazil	2x	40
I35	El Salvador	2x	35
I8	Rio Piedras, P.R.	4x	14
I10	Parbueyon, P.R.	4x	40
I11	Joyuna, P.R.	4x	26
I12	Dorado, P.R.	4x	13
I13	Manati, P.R.	4x	11
I14	Hwy 2 at 863, P.R.	4x	28
I15	Hwy 2 at 671, P.R.	4x	6
I16	Tortuguera, P.R.	4x	20
I17	Jarabacoa, D.R.	4x	15
I18	Santo Domingo, D.R. 1	4x	29
I19	Cabrete, D.R.	4x	10
I20	San FCO de Macoris, D.R.	4x	28
I22	Lucas Diaz, D.R.	4x	25
I23	Bani at Playa, D.R.	4x	12
I24	St Cristobal, D.R.	4x	21
I25	Santo Domingo, D.R. 2	4x	27
E2	Manaus, Brazil 1	4x	23
E3	Belém, Brazil	4x	25
E6	Manaus, Brazil 1	4x	21
E7	Recife, Brazil	4x	21
E8	Aracaju, Brazil	4x	28
E9	Maceio, Brazil	4x	24
E10	Vitória de S. Antão, Brazil	4x	18

I = variety *intermedia*,

E = variety *elegans*,

P.R. = Puerto Rico,

D.R. = Dominican Republic, x = 5.

Dominican Republic. Note that population I31 has recently been found to be intersterile with other diploid populations and may represent an undescribed species of *Turnera* (Arbo & Fernandez, 1987; M. M. Arbo, personal communication). Nevertheless, it is included in the analyses for comparisons of amounts of genetic variation between diploids and tetraploids (below). Diploid population I1 is considered to be *T. ulmifolia* var. *intermedia*, although, it is polymorphic for the presence of a purple petal spot characteristic of *T. ulmifolia* var. *elegans* (Shore & Barrett, 1987; Barrett, 1978).

Horizontal starch gel electrophoresis was used to obtain allozyme data for 10 enzyme systems. Aconitase (*Aco*), esterase (*Est*), glutamate dehydrogenase (*Gdh*), glutamate oxaloacetate transaminase (*Got*), glucose-phosphate isomerase (*Gpi*), leucine aminopeptidase (*Lap*), 6-phosphogluconate dehydrogenase (*Pgd*) and phosphoglucomutase (*Pgm*), were assayed following Shore & Barrett (1987). Malate dehydrogenase (*Mdh*) and alcohol dehydrogenase (*Adh*) were assayed following Cardy *et al.* (1980). Flower buds were used for the electrophoretic assays as they provided high levels of enzyme activity. For each enzyme, the locus coding the most anodally migrating protein was designated as locus 1, the next locus 2, etc., except where information was available on the intra-cellular location of the isozyme coded by the locus (Shore & Barrett, 1987). Isozymes localized in the plastids are denoted by the letter 'p' and cytosolic isozymes by 'c'. Allozymes coded by different alleles were designated by lower case letters, with the allele coding for the most anodally migrating allozyme designated 'a', the next 'b', etc. For ease of presentation, some alleles were re-coded for studies of inheritance (see below).

The inheritance of three isozyme loci, *Adh-2* for *T. ulmifolia* var. *intermedia* and *Aco-2* and *Gpi-c* for var. *elegans* was determined for tetraploids using controlled crosses of parents of known electrophoretic phenotype and assaying the resulting progeny. For *Adh-2* (a dimeric enzyme), a plant putatively of tetrasomic genotype *fffs* was crossed to four individuals homozygous (genotype *ffff*) at this locus. A reciprocal cross was done for one of the individuals thus providing a total of five families. A total of 96 progeny was scored for *Adh-2* activity by assaying mature pollen in which the locus is expressed.

For *T. ulmifolia* var. *elegans*, a heterozygous plant that was putatively tetra-allelic at the *Aco-2* locus was obtained. The genotype of the plant was *abcd*, where each letter refers to one of the four alleles producing electrophoretically distinguishable monomeric allozymes at this locus. The plant was crossed reciprocally to an individual of homozygous genotype, *bbbb*, and

85 progeny were scored. The inheritance of *Gpi-c* (the cytosolic isozyme locus of the dimeric enzyme, glucosephosphate isomerase) was also assessed in *T. ulmifolia* var. *elegans*. An individual with a putative tri-allelic genotype was crossed with an individual that was di-allelic as follows: $fmms \times mmms$ where f , m , and s , are three alleles at this locus. A total of 83 progeny was scored.

For all three loci, goodness of fit and heterogeneity tests were done to both tetrasomic (expected for autotetraploids) and two-locus disomic (expected for allotetraploids) models of inheritance, using the G statistic (Sokal & Rohlf, 1981). Tetrasomic ratios were predicted assuming no double reduction occurred at these loci (Allard, 1960).

A total of 652 individuals from 30 populations, representing both taxonomic varieties and ploidal levels, were analysed electrophoretically to compare levels of genetic variation across ploidal levels and taxa. For the 10 enzyme systems assayed, 14 loci could be resolved sufficiently well for both diploids and tetraploids. These were: *Aco-1*, *Aco-2*, *Adh-1*, *Est*, *Gdh*, *Got-1*, *Got-2*, *Gpi-c*, *Gpi-p*, *Lap-1*, *Lap-2*, *Mdh-1*, *Pgd-c*, and *Pgm-c*. The inheritance of nine of the 14 loci has been determined for diploid *T. ulmifolia* (Shore & Barrett, 1987). For tetraploids, the banding patterns were interpreted as if they were the result of tetrasomic inheritance, given the evidence below. It was necessary to infer the dosage of alleles from the relative intensity of the bands to determine the genotype of each tetraploid plant at each locus (e.g. Figs 1 and 2).

The percentage of polymorphic loci, mean number of alleles per locus, mean number of alleles per polymorphic locus, Nei's (1973) index of gene diversity and the observed proportion of heterozygosity were estimated for each population. Mean levels of genetic variation for each variety at each ploidal level were compared statistically, using the Wilcoxon rank-sum test, setting the experimentwise error rate at 0.05, for the three comparisons.

The partitioning of total gene diversity (H_T) among populations (D_{ST}) of each variety at each ploidal level, as well as within populations (H_S), was assessed using

the diversity analysis of Nei (1973). Gene diversity among populations relative to total diversity was calculated as $G_{ST} = D_{ST}/H_T$. In addition, genetic distances were calculated among all pairs of populations followed by a UPGMA cluster analysis (Nei, 1987).

Results

Inheritance of isozymes

To test the hypothesis that tetraploid *T. ulmifolia* var. *elegans* and *intermedia* are autopolyploids, the inheritance of three isozyme loci was investigated. For *Adh-2* in *T. ulmifolia* var. *intermedia*, segregation ratios from the cross $ffss \times ffff$ are presented in Table 2. The parental and three progeny genotypes are shown in Fig. 1. Note that heterozygous genotypes, $fffs$ and $ffss$, are readily distinguished by differences in banding intensity. The results fit the expected tetrasomic ratios while deviating significantly from a two-locus disomic model of inheritance. Progeny ratios among replicate crosses were homogeneous (Table 2).

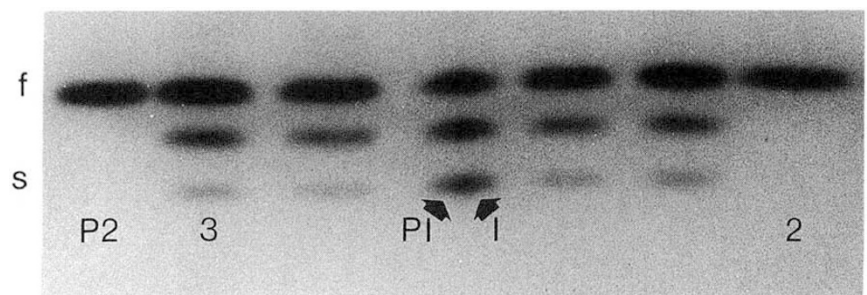
The inheritance of two isozyme loci was determined for tetraploid *T. ulmifolia* var. *elegans*. For *Gpi-c*, progeny of the cross $fmms \times mmms$ were scored and a photograph showing parental and progeny genotypes

Table 2 Tetrasomic inheritance of *Adh-2* in tetraploid *Turnera ulmifolia* var. *intermedia*. Putative genotypes crossed were $ffss \times ffff$ and reciprocal. Goodness of fit G tests to tetrasomic and two-locus disomic models of inheritance are provided as is a G statistic for heterogeneity among crosses

	Genotype			G	d.f.	P
	$ffff$	$fffs$	$ffss$			
Progeny observed	20	60	16			
Expected tetrasomic	16	64	16	1.18	2	>0.50
Expected disomic	24	48	24	6.51	2	<0.05
$G_{\text{heterogeneity}} = 8.39, \text{d.f.} = 8, P > 0.25$						

d.f. = degrees of freedom.

Fig. 1 Photograph of an isozyme gel for the *Adh-2* locus. Parental (P1 and P2) and segregating progeny genotypes (lanes 1, 2 and 3) are indicated. Three different genotypes occur. The genotypes are: P1 and 1 = $ffss$; P2 and 2 = $ffff$; 3 = $fffs$.



may be found in Barrett & Shore (1989). Segregation ratios fit a tetrasomic model of inheritance and deviate significantly from a two-locus disomic model of inheritance (Table 3). The data also illustrate the occurrence of heterozygous plants containing three alleles at a single locus with some alleles carried in different doses.

For *Aco-2*, the cross *abcd* × *bbbb* was made and progeny were scored and compared to expected tetrasomic ratios (Table 4). The parental plants and the six different progeny genotypes are shown in Fig. 2. No two-locus disomic model of inheritance can account for the production of six progeny genotypes, thus discounting this as a possible mechanism of inheritance. The progeny genotypes and segregation ratios are, however, consistent with a tetrasomic model of inheritance. There is no evidence of double reduction at this

locus as additional progeny genotypes, that would uniquely mark the occurrence of double reduction, were not observed. The locus further illustrates the manner in which tetrasomic genotypes can be determined from electrophoretic phenotypes (Fig. 2).

Isozyme variation in natural populations

Levels of genetic variation within each population are presented in Table 5 and means for each variety at each ploidal level are provided in Table 6. Interestingly, there are differences in the levels of genetic variation for tetraploids of the two varieties (Tables 5 and 6). Tetraploid *T. ulmifolia* var. *elegans* exhibits significantly greater levels of genetic variation, as assessed by a Wilcoxon rank-sum test, for percentage polymorphic

Table 3 Tetrasomic inheritance of *Gpi-c* in var. *elegans*. The cross *fnms* × *mmms* was carried out. Goodness of fit *G* tests for tetrasomic and two-locus disomic models of inheritance are provided

	Genotype						<i>G</i>	d.f.	<i>P</i>
	<i>fnmm</i>	<i>fnms</i>	<i>fnss</i>	<i>mmmm</i>	<i>mmms</i>	<i>mmss</i>			
Progeny observed	11	18	6	5	20	23			
Expected tetrasomic	13.8	20.8	6.9	6.9	20.8	13.8	6.8	5	>0.2
Expected disomic	10.4	20.8	10.4	10.4	20.8	10.4	17.4	5	<0.01

d.f. = degrees of freedom.

Table 4 Tetrasomic inheritance of *Aco-2* in tetraploid *Turnera ulmifolia* var. *elegans*. Putative genotypes crossed were *abcd* × *bbbb* and reciprocal. No two-locus disomic model can account for the occurrence of six progeny genotypes from this cross. Goodness of fit to a tetrasomic model of inheritance and heterogeneity *G* statistics are provided

	Genotype						<i>G</i>	d.f.	<i>P</i>
	<i>bbcd</i>	<i>bbbd</i>	<i>bbbc</i>	<i>abbd</i>	<i>abbc</i>	<i>abbb</i>			
Progeny observed	19	14	11	13	14	14			
Expected tetrasomic	14.17	14.17	14.17	14.17	14.17	14.17	2.36	5	>0.75
<i>G</i> _{heterogeneity} = 5.09, d.f. = 5, <i>P</i> > 0.25									

d.f. = degrees of freedom

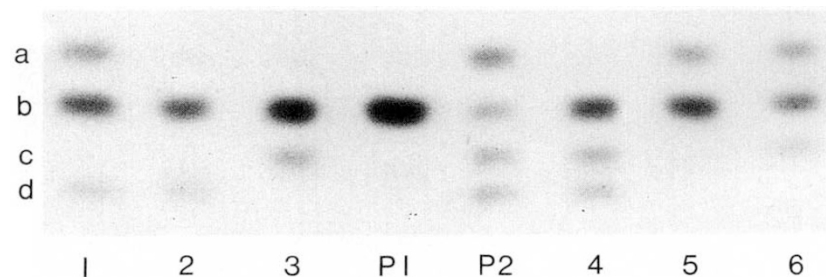


Fig. 2 Photograph of an isozyme gel for the *Aco-2* locus. Parental (lanes P1 and P2) and the six progeny genotypes observed (lanes 1–6) are shown. The genotypes are: P1 = *bbbb*; P2 = *abcd*; 1 = *abbd*; 2 = *bbbd*; 3 = *bbbc*; 4 = *bbcd*; 5 = *abbb*; 6 = *abbc*.

Table 5 Estimates of isozyme variation within populations of *T. ulmifolia*

Population	Ploidal level	Percentage polymorphic loci	Mean alleles/locus	Mean alleles/polymorphic locus	Gene diversity	Observed heterozygosity
I1	2x	50	1.5	2.0	0.16	0.17
I2	2x	43	1.6	2.3	0.09	0.08
I26	2x	43	1.5	2.2	0.11	0.09
I28	2x	50	1.6	2.1	0.15	0.10
I29	2x	36	1.4	2.2	0.08	0.07
I31	2x	43	1.6	2.5	0.11	0.08
I35	2x	57	1.6	2.1	0.19	0.17
I8	4x	21	1.2	2.0	0.03	0.06
I10	4x	14	1.1	2.0	0.01	0.02
I11	4x	21	1.2	2.0	0.05	0.10
I12	4x	14	1.1	2.0	0.01	0.02
I13	4x	14	1.1	2.0	0.05	0.12
I14	4x	14	1.1	2.0	0.04	0.07
I15	4x	14	1.1	2.0	0.03	0.07
I16	4x	14	1.1	2.0	0.03	0.05
I17	4x	14	1.1	2.0	0.03	0.05
I18	4x	29	1.3	2.0	0.06	0.09
I19	4x	7	1.1	2.0	0.04	0.05
I20	4x	21	1.2	2.0	0.02	0.04
I22	4x	36	1.4	2.0	0.04	0.07
I23	4x	14	1.1	2.0	0.03	0.07
I24	4x	29	1.3	2.0	0.06	0.10
I25	4x	43	1.4	2.0	0.06	0.10
E2	4x	71	2.2	2.7	0.24	0.32
E3	4x	43	1.6	2.3	0.21	0.34
E6	4x	57	1.7	2.3	0.24	0.39
E7	4x	64	2.1	2.8	0.31	0.45
E8	4x	79	2.2	2.5	0.31	0.49
E9	4x	71	2.1	2.6	0.30	0.50
E10	4x	71	2.2	2.7	0.27	0.42

Table 6 Mean levels of isozyme variation for each taxon/ploidal level. Means sharing the same letter (a, b or c) are not significantly different, based upon the Wilcoxon rank-sum test

Taxon	Ploidal level	Percentage polymorphic loci	Mean alleles/locus	Mean alleles/polymorphic locus	Gene diversity	Observed heterozygosity
<i>Intermedia</i>	2x	45.9 ^a	1.55 ^a	2.20 ^a	0.12 ^a	0.11 ^a
<i>Intermedia</i>	4x	20.1 ^b	1.20 ^b	2.00 ^b	0.04 ^b	0.07 ^a
<i>Elegans</i>	4x	65.3 ^c	2.03 ^c	2.56 ^c	0.27 ^c	0.42 ^b

loci, mean number of alleles per locus and gene diversity, as well as having the greatest levels of observed heterozygosity. In marked contrast, tetraploid populations of *T. ulmifolia* var. *intermedia* exhibited the

lowest estimates (Table 6). The means for diploids were intermediate.

Gene diversity was partitioned within and among populations of each variety at each ploidal level. Tetra-

Table 7 Gene diversity statistics for *T. ulmifolia* vars. *elegans* and *intermedia* at each ploidal level

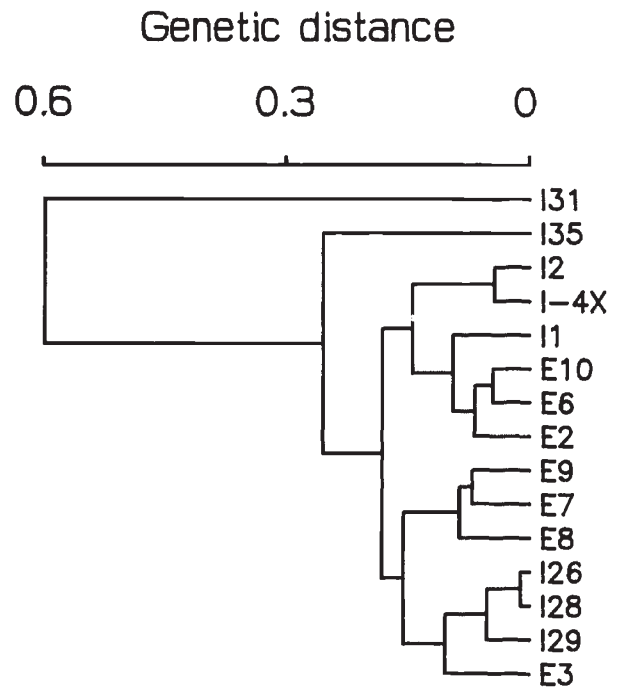
Taxon	Ploidal level	H_S	D_{ST}	H_T	G_{ST}
<i>Intermedia</i>	2x	0.12	0.19	0.31	0.60
<i>Intermedia</i>	4x	0.04	0.01	0.05	0.21
<i>Elegans</i>	4x	0.27	0.07	0.34	0.21

ploidy *T. ulmifolia* var. *intermedia* exhibited low levels of total diversity ($H_T=0.05$), a greater proportion of which occurred within populations (Table 7). *T. ulmifolia* var. *elegans* exhibited the largest total gene diversity ($H_T=0.34$), again with the greater proportion occurring within populations ($H_S=0.27$). Diploid *T. ulmifolia* var. *intermedia* had a total gene diversity similar to that of var. *elegans*, however, 60 per cent of the diversity ($G_{ST}=0.6$) was partitioned between populations as compared to var. *elegans*, for which 21 per cent of the gene diversity ($G_{ST}=0.21$) occurred among populations.

Gene diversity for diploid *T. ulmifolia* var. *intermedia* populations was also calculated, excluding population I31, the potentially new species of *Turnera*. When this was done, the following diversity values were obtained: $H_S=0.13$, $D_{ST}=0.12$, $H_T=0.24$, $G_{ST}=0.48$. This analysis had the effect of reducing the total diversity as well as the between population component of diversity. Diploid *T. ulmifolia* var. *intermedia*, however, still had the largest proportion of gene diversity among populations, in comparison with the tetraploids.

To provide information on the possible origins of the polyploids, genetic distances among all populations were calculated and a UPGMA cluster analysis was used to construct a dendrogram (Fig. 3). In addition, the numbers of unique alleles present in each variety at each ploidal level were enumerated. Diploid *T. ulmifolia* var. *intermedia* (excluding population I31) had three unique alleles; tetraploid var. *intermedia*, one; and tetraploid var. *elegans*, six. The numbers of alleles present in the tetraploids that were not present in the diploids were also counted. For tetraploid *T. ulmifolia* var. *intermedia*, two alleles occurred that did not occur in the diploid populations. For tetraploid *T. ulmifolia* var. *elegans*, nine alleles occurred which were not represented in the diploids.

Diploid population I31 from Brazil proved to be genetically distant to the remaining populations (Fig. 3) supporting the crossing data of Arbo & Fernandez (1987), which suggests that it should be recognized as a new species of *Turnera*. All 16 tetraploid populations

**Fig. 3** Dendrogram produced from UPGMA clustering of genetic distances among populations of *Turnera ulmifolia*. I-4X represents a single cluster composed of all tetraploid var. *intermedia* populations.

of *T. ulmifolia* var. *intermedia* clustered together and are represented as I-4X (Fig. 3). Interestingly, these tetraploids clustered most closely with a diploid population of *T. ulmifolia* var. *intermedia* from Venezuela. The three Costa Rican diploids (populations I26, I28 and I29) form a cluster which also contains a Brazilian population (E7) of tetraploid *T. ulmifolia* var. *elegans*. Diploid population I1 clusters with some tetraploid *T. ulmifolia* var. *elegans* populations (populations E2, E6 and E10).

Discussion

The demonstration of tetrasomic inheritance at isozyme loci provides an independent source of evidence for autopolyploidy in tetraploid *T. ulmifolia* vars. *elegans* and *intermedia*. Coupled with cytological observations of chromosome configurations at meiosis (Raman & Kesavan, 1964; Arbo & Fernandez, 1983; Fernandez, 1987; Shore, 1991), and studies of the inheritance of distyly in tetraploids (Shore & Barrett, 1985b), it provides strong support for autotetraploidy within this species complex. Thus, these taxonomic varieties of *T. ulmifolia* may be added to the relatively short list of natural autotetraploid plants (reviewed in Soltis & Rieseberg, 1986; Krebs & Hancock, 1989).

Empirical studies of the genetic consequences of autopolyploidy are few in number (Soltis & Rieseberg, 1986), however, the theoretical consequences have been examined. The population genetics of polysomy was formulated by Haldane (1930) and extended by other workers (e.g. Bennett, 1968; Wright, 1969; Mayo, 1971; McConnell & Fyfe, 1975; Mayo, 1983). Autopolyploid populations should maintain greater levels of heterozygosity and exhibit greater resistance to allele frequency shifts due to drift than an otherwise comparable diploid population.

The manner in which an autopolyploid population becomes established may also have an important influence on the extent of genetic variation within the population. In particular, the amount of genetic variation that traverses the bottleneck between diploid and autopolyploid needs to be considered both theoretically and investigated experimentally. Furthermore, the history of the autopolyploid population and time since its origin need to be taken into account. For example, gene exchange with diploid populations via unreduced gametes (DeWett, 1979), or hybridization with autopolyploid populations of independent origin, might result in an influx of genetic variation.

Diploid and tetraploid *T. ulmifolia* var. *elegans* are sympatric in parts of their range (Shore & Barrett, 1986; Barrett & Shore, 1987), providing opportunities for gene exchange. No such opportunity, however, occurs for island populations of tetraploid *T. ulmifolia* var. *intermedia*. This may account for the striking difference in isozyme variation in the two autotetraploids. An additional possibility is that the occurrence of tetraploid *T. ulmifolia* var. *intermedia* on islands, and the resulting founder effect following long-distance dispersal could account for the low levels of isozyme variation. Island populations of *Eichhornia paniculata* (Glover & Barrett, 1987) and *Lycopersicon cheesmanii* (Rick & Fobes, 1975) have been found to be genetically depauperate in comparison with mainland populations.

Not only did tetraploid *T. ulmifolia* var. *elegans* have the greatest levels of genetic variation but, in addition, possessed nine alleles not found in the sample of diploid populations (six alleles were unique to tetraploid *T. ulmifolia* var. *elegans*). This could indicate that *T. ulmifolia* var. *elegans* has undergone hybridization with locally differentiated diploid populations that were not included in this study. Alternatively, a reduction in the effects of drift due to autopolyploidy and the accumulation of novel alleles via mutation, might account for these observations. Additional population samples, particularly of diploid *T. ulmifolia* var. *elegans*, are required to evaluate these alternatives.

Since tetraploid populations of *T. ulmifolia* are

distylous and self-incompatible (Shore & Barrett, 1985b; Barrett & Shore, 1987), it is highly improbable that a single tetraploid plant could establish an obligately outcrossing, distylous, autotetraploid population. The number and source of individuals giving rise to autotetraploid populations and varieties is, however, unknown. The dendrogram (Fig. 3) indicates that tetraploid *T. ulmifolia* var. *intermedia* populations are most similar to a diploid var. *intermedia* population from Venezuela which, however, was represented by a small sample of plants. Different populations of tetraploid *T. ulmifolia* var. *elegans* clustered with different diploid var. *intermedia* populations, perhaps suggesting multiple origins of this variety. Information on the number of independent origins of autotetraploidy in *T. ulmifolia* will likely require the use of molecular biological methods, following the approach used by Soltis *et al.* (1989) for autopolyploid *Heuchera micrantha*.

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