

**GENETIC VARIABILITY IN *MUSCARI COMOSUM* (LILIACEAE).
I. A COMPARATIVE ANALYSIS OF CHROMOSOME
POLYMORPHISMS IN SPANISH AND AEGEAN POPULATIONS**

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1. INTRODUCTION

GARBARI (1966, 1969) has reported a stable structural polymorphism in populations of Italian *Muscari comosum*. This was for a pericentric inversion in the 2nd largest pair of chromosomes. The same polymorphism was subsequently seen by Bentzer (1972) and Bentzer and Ellmer (1975) in material from several Aegean Islands (Greece). These authors also reported a heteromorphism for the size of the chromosome concerned. A similar situation has since been noted in populations from Egypt (Bentzer, unpublished data, cited in Bentzer, 1972). This paper reports a cytological analysis of seven Spanish populations, in which the arrangement frequencies are compared with those present in the Aegean Islands.

2. MATERIALS AND METHODS

A total of 311 bulbs from seven Spanish populations (table 1) were collected during 1978, 1979, and 1980. The bulbs chosen were usually

TABLE 1

Number of plants with the various cytotypes of chromosome 2

Key	Population	Sample size	Cytotypes						
			+ / +	+ / i	i / i	+ / +1	+ / i1	i / +1	i / i1
GAR	Gargantilla (Madrid)	100	44	30	4	7	8	6	1
COL	Colmenar (Málaga)	50	26	16	1	—	4	—	3
SIL	Silleta (Granada)	50	20	18	5	2	3	1	1
PAD	Padul (Granada)	35	31	4	—	—	—	—	—
CAZ	S ^a Cázulas (Granada)	18	10	4	2	1	—	—	1
MOL	El Mojar (Madrid)	18	10	6	—	—	2	—	—
COZ	Cozviñar (Granada)	40	35	5	—	—	—	—	—
	Total	311	176	83	12	10	17	7	6

more than one meter apart. Young inflorescences were fixed in ethanol: acetic acid (3 : 1). The total size and arm proportion of the first and second chromosome pairs were measured in five metaphase cells from the root tips of each bulb. The meiotic behaviour of PMCs was analysed by staining young anthers in acetic-carmin.

3. RESULTS

(i) *Cytogenetic analysis*

All the bulbs analysed had the diploid value typical for this species with $2n = 18$ (fig. 1). Four types of chromosome 2 were present and these agree with those described by Bentzer and Ellmer (1975) in Aegean populations: Normal (+) (fig. 1), inversion type (i) (fig. 2), inversion with extra long arm (il) (fig. 3), and normal with extra long arm (+l) (fig. 4). These four types have the same *r*-values and proportions reported by Bentzer (1972) and Bentzer and Ellmer (1975). Because the centromere position differs without affecting the chromosome size, the inversion chromosome type probably originated by a pericentric inversion (Bentzer, 1972). Although a centric transposition cannot be excluded, for the purpose of this paper it may be regarded as an inversion rearrangement. This chromosome pair shows a normal meiotic behaviour in both homozygotes (+/+ or i/i). In some (20 per cent) of the diakinesis cells from individuals carrying a heterozygous duplication (*i.e.*, +l or il) autosynapsis of the distal part of the longer chromosome is found (fig. 5).

(ii) *Chromosome variability in Spanish populations*

The number of plants with the various combinations of chromosome 2, and the sample size in each population, are shown in table 1. In all seven populations the + chromosome is the most frequent, but the i type is present in substantial proportions in all except for PAD and COZ populations. The +l and il chromosomes are found in low frequencies in GAR, SIL, and CAZ, while only the il type is present in the COL and MOL populations. All the Spanish populations lack homozygotes for the extra long arm (*i.e.*, combinations +l/+l, il/il, or +l/il).

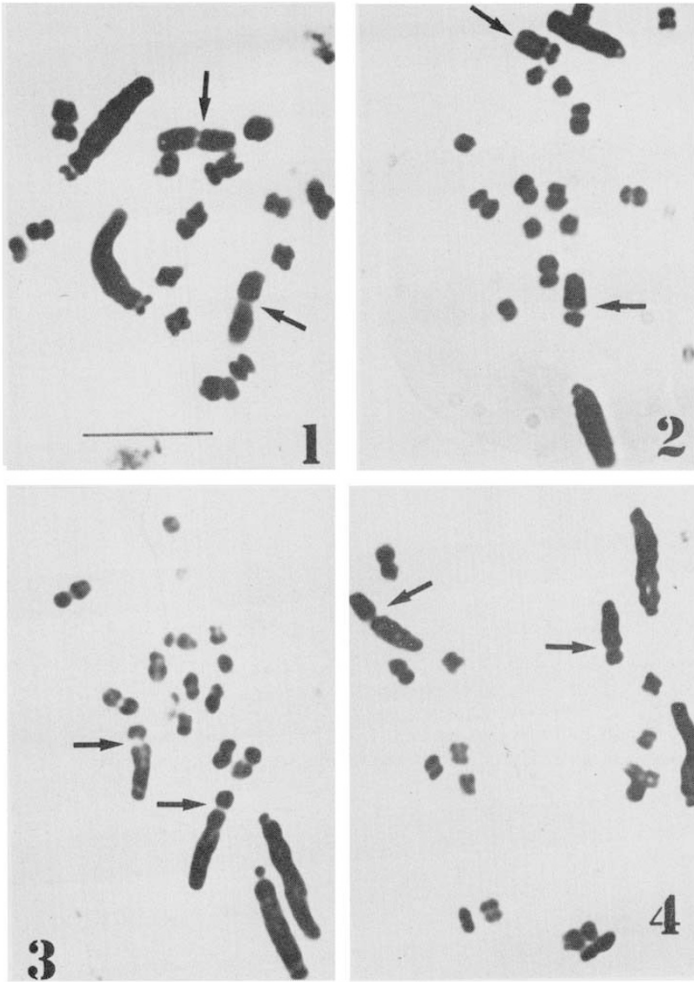
(iii) *A comparison of the Spanish and Aegean populations*

Our data from the seven Spanish populations of *M. comosum* are combined with those of Bentzer and Ellmer (1975) on six Aegean populations in table 2. An estimate of the degree of chromosome differentiation for all populations is possible through the use of the distance statistic of Prevosti (1974) and Prevosti *et al.* (1975). This allows one to construct a distance matrix and from this the dendrogram shown in the fig. 6 was obtained by a single-linkage cluster analysis (Davies, 1971). We have also analyzed the total inversion frequencies, the observed inversion heterozygosity, and the means for each geographical area (table 3).

4. DISCUSSION

The populations of *Muscari comosum* from the Iberian Peninsula (the western end of its Mediterranean distribution) carry the same chromosome variants which Bentzer and Ellmer (1975) reported from the Aegean Islands (central zone). As Bentzer (1972) indicated, one of these is probably a pericentric inversion; on the basis of its meiotic behaviour the other is probably a segmental duplication. If the rearrangements involved in this

Plate I



FIGS. 1-4.—Metaphase plates from $+/+$ (Fig. 1), i/i (Fig. 2), i/il (Fig. 3), and $+/+1$ (Fig. 4) individuals. Arrows indicate the second chromosome pair. Scale equals $5\ \mu$.

Plate II

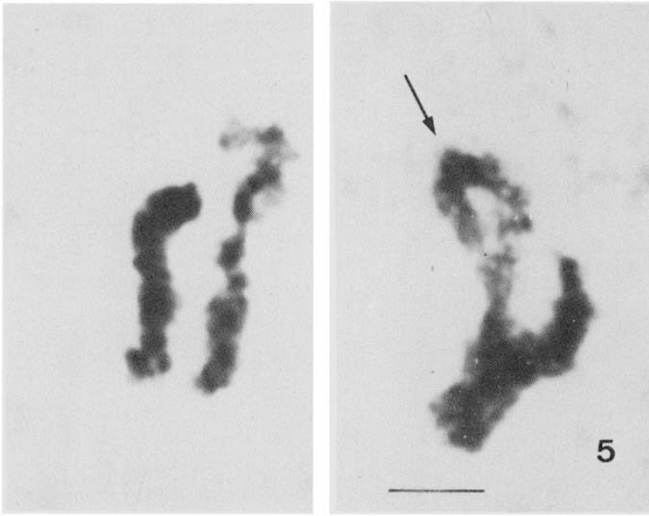


FIG. 5.—Structural heterozygote bivalent from *il/i* individual. Normal synapsis (left) and autosynapsis (right). Arrow indicates autosynapsis. Scale equals $2\ \mu$.

TABLE 2
Arrangement frequencies in Spanish and Aegean populations

Population	Sample size	Arrangements			
		+	i	non-l	l
(a) Spain					
GAR	200	0.730	0.270	0.890	0.110
COL	100	0.720	0.280	0.930	0.070
SIL	100	0.660	0.340	0.930	0.070
PAD	70	0.943	0.057	1.000	—
CAZ	36	0.722	0.278	0.944	0.056
MOL	36	0.778	0.222	0.944	0.056
COZ	80	0.938	0.062	1.000	—
(b) Greece					
600 (Kithnos)	36	0.417	0.583	0.972	0.028
601 (Kithnos)	144	0.708	0.292	0.944	0.056
602 (Kithnos)	106	0.613	0.387	0.858	0.142
603 (Rodhos)	84	0.464	0.536	0.940	0.060
604 (Rodhos)	52	0.481	0.519	0.865	0.135
605 (Kithnos)	74	0.703	0.297	0.919	0.081

study are unique in origin, their distribution over such a wide range can only be attributed to the migration of either individuals or gametes.

Bentzer and Ellmer (1975) explained the different frequencies of chromosome types found in the Aegean Islands in terms of genetic drift. From our data we would argue that adaptive and historical factors, related to the colonization process, may be also important to explain the total geographical distribution of both chromosome rearrangements.

Like Bentzer and Ellmer (1975) we have not found homozygous individuals for the extra long arm (*i.e.*, *il/il*, *+1/+1*, or *+1/il*). We appreciate that the low frequencies of *+1* and *il* chromosomes produce a very low probability of finding them together in one plant. Nevertheless, when a Student *t*-test with the angular transformation of the data (Sokal and Rohlf, 1969) is applied to the Spanish population from GAR, which has the highest sample size, a statistically significant deficiency of duplication homozygotes is found ($t_s = 2.20$; $P: 0.05 - 0.01$). Likewise, if the expected frequencies of chromosome types, as computed from the rearrangement frequencies (table 2), are compared with the observed frequencies in the Spanish population from COL, a statistically significant deficiency of *+1* chromosomes is found ($t_s = 3.20$; $P: 0.01 - 0.001$). There are also significantly fewer *il* chromosomes in the population 601 from Kithnos than is expected ($t_s = 2.18$; $P: 0.05 - 0.01$). Therefore, it seems that the chromosomes carrying the extra material may be selected against.

On the other hand, from the analysis of the dendrogram (fig. 6), we can conclude that:

(1) If population 600 is discounted, the populations from Kithnos tend to cluster with the Spanish ones and appear more distantly related to the populations from Rodhos. The higher similarity between Spain and Kithnos is due to their lower inversion frequencies. The Rodhos collection, on the other hand, shows a higher frequency of inversion chromosomes. When inversion frequencies in Aegean and Spanish populations are compared

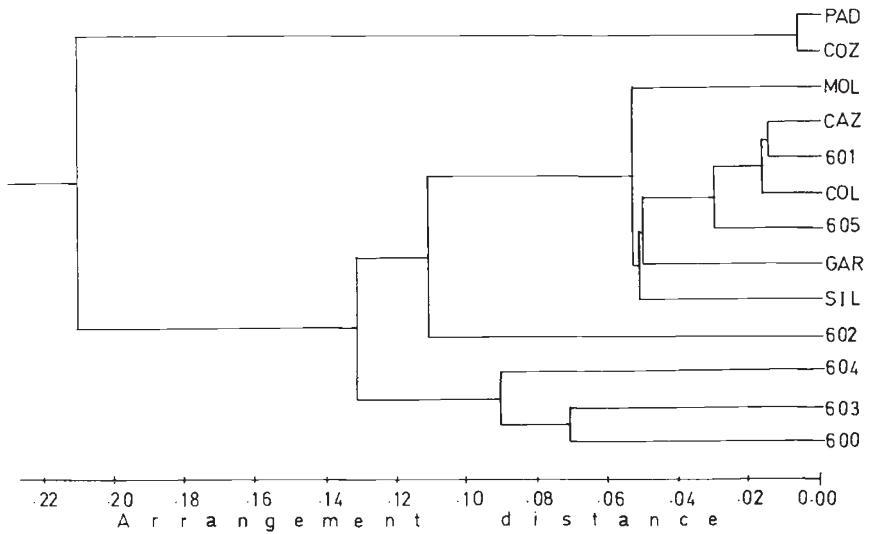


FIG. 6.—Dendrogram showing the relationships between Spanish and Aegean populations of *M. comosum*, as inferred from their arrangement distance matrix.

(table 3), striking differences are found. The mean inversion frequencies are higher in Rodhos, intermediate in Kithnos, and lower in the Iberian Peninsula. More samples from other geographical areas need to be analyzed to determine whether there is any clinal relationship which relates to colonization and expansion of *M. comosum*.

(2) The two Spanish populations from PAD and COZ appear as the most differentiated ones. These populations offer an interesting opportunity to test the respective roles played by natural selection and genetic drift in maintaining the inversion polymorphism in question. Both Spanish populations have the lowest inversion frequencies (table 3). Their heterozygosity values (table 3) are also significantly lower than those present in the remaining Spanish and Aegean populations. Furthermore, the two Spanish

TABLE 3

Total inversion frequencies, observed inversion heterozygosities and means for each geographical area in Mediterranean populations of *M. comosum*

	Population	Inversion frequency	Mean	Observed heterozygosity	Mean
Rodhos	603	0.54	0.53 ± 0.01	0.37	0.48 ± 0.11
	604	0.52		0.58	
	600	0.58		0.72	
Kithnos	601	0.29	0.39 ± 0.07	0.39	0.56 ± 0.07
	602	0.38		0.62	
	605	0.30		0.49	
	GAR	0.27		0.44	
	COL	0.28		0.40	
Spain	SIL	0.34	0.22 ± 0.04	0.44	0.31 ± 0.06
	PAD	0.06		0.13	
	CAZ	0.28		0.22	
	MOL	0.22		0.44	
	COZ	0.06		0.13	

populations show characteristics which are typical of an ecological border, namely: recent colonization (they were cultivated fields ten years ago), low population size, and differential edaphic conditions.

The PAD and COZ populations have a normal level of allozyme polymorphism, as compared with the remaining Spanish populations (Pascual *et al.*, in preparation). In both PAD and COZ there is therefore no restriction of genic variability. This constitutes further support for the well established conclusion that allozyme polymorphisms show more uniform geographical distribution than chromosome polymorphisms (Ayala *et al.*, 1971; González-Duarte *et al.*, 1973; Powell, 1973; Saura and Lakovaara, 1973; Lewontin, 1974).

One final conclusion emerges from our data. If the low inversion heterozygosities from PAD and COZ populations are maintained by genetic drift (*e.g.*, founder effect), then the normal level of allozyme variability is difficult to explain. On the other hand, if the low inversion heterozygosities from these two populations are maintained by natural selection, the existence of a normal level of allozyme polymorphism in three particular enzymes can be readily explained.

In conclusion, although genetic drift cannot be rejected, we think that the interaction between adaptive and historical factors, related to the colonization process, have played the major role in determining the geographical distribution of the two coupled stable chromosome polymorphisms present in *Muscari comosum*.

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