

GENE FREQUENCIES IN WILD POPULATIONS OF *TRIFOLIUM REPENS*

I. DISTRIBUTION BY LATITUDE

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I. INTRODUCTION

THE presence of hydrocyanic acid (HCN) in *Trifolium repens* L. has been realised for some time (Mirande, 1912). The plants possess cyanogenetic glucosides consisting of 80 per cent. lotaustralin and 20 per cent. linamarin (Melville and Doak, 1940) and the term glucoside is used in this paper to denote these cyanogenetic glucosides. The enzyme linamarase hydrolyses the glucoside into its components with the production of HCN.

Research has been carried out on hydrocyanic acid in *T. repens*, from three points of view: its amount in the plants (Doak, 1933; Rigg *et al.*, 1933; and Askew, 1933); its significance in seed certification (Foy and Hyde, 1937); and its possible role in contributing to bloat in ruminants (Evans and Evans, 1948). Further investigations have been concerned with the isolation of the glucoside (Melville and Doak, 1940), and of the enzyme (Coop, 1940) and their inheritance (Williams, 1939; Corkill, 1942; Atwood and Sullivan, 1943). Genetical studies (Corkill, 1942; Atwood and Sullivan, 1943) have shown that two independent genes determine the production of the glucoside *Ac-ac*, and enzyme *Li-li*. *T. repens* plants bearing both dominant genes crossed with the double recessive, gave a normal dihybrid segregation in F_2 .

Previous unpublished studies made by the late R. D. Williams had shown that the percentage of cyanogenetic plants varied from 0-100 in populations of different geographical origin. The purpose of the present investigation was to determine the gene frequency in wild populations and to relate it to environmental factors. The four phenotypes:

- (1) Glucoside and enzyme (*AcLi*)
- (2) Glucoside only (*Acli*)
- (3) Enzyme only (*acLi*)
- (4) Neither glucoside nor enzyme (*accli*)

were distinguished by the picric acid test, using isolated lotaustralin and linamarase solutions as described by Corkill (1940). This technique, however, was modified by retesting the plants provisionally classified as the *Acli* phenotype with a solution of isolated lotaustralin

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to detect small amounts of enzyme, as will be described in a later paper. The Hardy-Weinberg formula ($p^2 + 2pq + q^2 = 1$) was applied to determine the genotypic frequencies of the populations (Hardy, 1908; Weinberg, 1908).

A collection of wild *T. repens* seeds was made from most European and some Near Eastern countries. This collection was made possible by the kind co-operation of the British Council, Agricultural Experimental Stations, Universities and Agricultural Ministries in the countries concerned. They were requested to adhere to the following conditions of seed collection: 20 wild *T. repens* seed heads to be collected from five different ecological areas around the named locality or "collection centre"; the collection to be of wild *T. repens* only, and the place of collection to be well isolated from any cultivated land or reseeded pasture.

2. WILD *T. REPENS* POPULATIONS

Wild *T. repens* is widely distributed in Europe and Asia, where it can be found in pastures, meadows, on road sides, river banks, etc. Kousnetzoff (1926) has suggested that *T. repens* and most of the other *Trifolium* species have their centre of origin in the Mediterranean region. Wild *T. repens* has spread throughout Europe, as far as the Lake Baikal region on the mainland of Asia and to Japan and Ceylon, growing at altitudes up to 2200-2300 m. above sea level in the Alps and probably up to the same altitude in other mountain regions. *T. repens* is not known to have been indigenous outside the Old World.

The genetical structure of European and Near Eastern populations

The *phenotypic structure* of populations can be expressed by the percentage frequency of the four phenotypes (*AcLi*, *Acli*, *acLi* and *acli*) and these are set out in Appendix 1. Considerable variation exists between populations in regard to their phenotypic proportions. Heterogeneity tests, using 2×2 contingency tables, were made to determine whether any differences existed between the phenotype frequency in samples from different geographical regions, and for this purpose the four phenotypes were grouped into two as follows: (a) *AcLi* phenotype; (b) *Acli*, *acLi* and *acli* phenotypes.

An analysis of the population samples from geographical regions shows that the Mediterranean populations possess the highest frequency of *AcLi* phenotype. In Greece (Athens), for example, the frequency is 100 per cent., and in Israel (Huleh District), Italy (Lucca), Spain (La Coruña) the *Acli* and *acli* phenotypes occur only occasionally. Samples from northern Italy (San Daniele del Friuli, Milan), however, differ markedly from the above populations, the majority of the plants being of the *acli* type. In some of the populations from Ireland (Longford), Great Britain (Aberystwyth), France (Finistère), two to

four phenotypes are found, but in all cases the *AcLi* type predominates. The decrease in the frequency of the *AcLi* phenotype from the Mediterranean region to France is significant (heterogeneity $\chi^2 = 11.62$, $P < 0.001$), as is also the decrease in frequency from France to Great Britain ($\chi^2 = 14.72$, $P < 0.001$). All four phenotypes are represented in most of the German and Swiss populations (Hohenheim, Lausanne, Zürich), but the *AcLi* phenotype decreases in favour of the *Acli*, *acLi* and *acli* phenotypes; the difference between the British and German populations is so obvious as to require no statistical proof. A Dutch

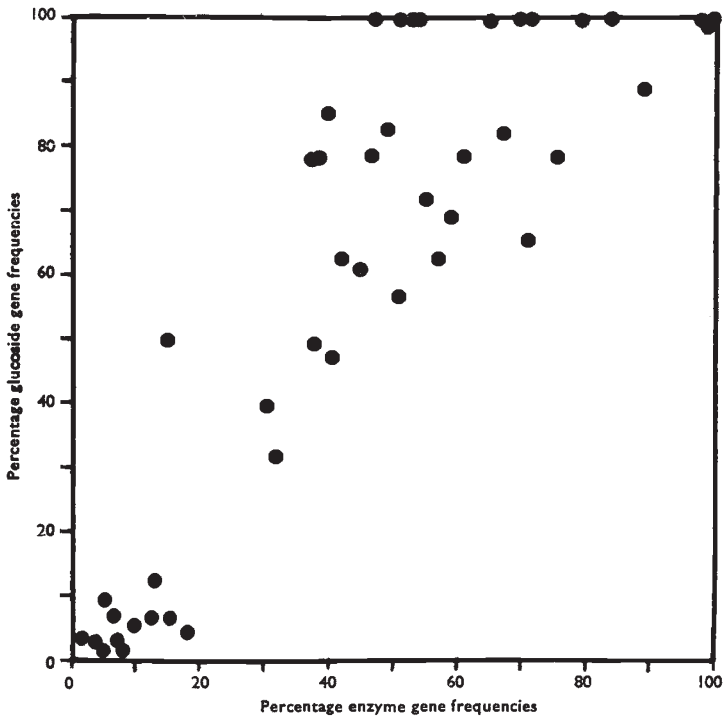


FIG. 1.—Frequencies of the dominant glucoside lotaustralin (*Ac*) and enzyme linamarase (*Li*) genes in European and Near Eastern wild populations of *Trifolium repens*.

sample (Breda) shows approximately equal proportions of the four phenotypes. The trend continues in Denmark (North Jutland), Germany (Hanover, Straubing), Austria (Rupprechtshofen), Netherlands (Groningen), where the *acli* phenotype becomes predominant. Marked trends are also noticeable in other central, eastern, and northern European samples. From the analyses it appears that the frequency of the *acli* phenotype reaches a maximum in Norwegian (Tjøtta), Hungarian (Budapest), and Polish (Kościan) samples. Unpublished data of R. D. Williams on Swedish and Russian populations indicate the existence of populations consisting entirely of acyanogenetic plants.

Genotypic structure.—In the investigation of the genotypic structure of populations, the proportion of dominant homozygous (p^2), heterozygous ($2pq$) and recessive homozygous (q^2) genotypes was determined by the Hardy-Weinberg formula. The genotypic frequencies of glucoside (*Ac-ac*) and enzyme genes (*Li-li*) are presented in Appendix 2.

When the glucoside and enzyme gene frequencies are plotted against one another (fig. 1) they show a gradation from very low values to extremely high values. As will be seen later, moreover, this gradation forms a close parallel with the geographical origin ranging from the low values for north-eastern Europe to the high values for the Mediterranean region.

3. RELATION OF GENE FREQUENCY TO THE ENVIRONMENTAL FACTORS

Marked differences have been found to occur in the genetical structure of wild *T. repens* populations from different regions in Europe and the Near East. In view of the progressive increase in the frequency of recessive genes in moving from south to north-east Europe, an attempt has been made to correlate this with environmental factors.

No correlation can be discerned between the gene frequency of populations and the rainfall of the regions in which they exist, the annual isotherms, or the July isotherm. However, it was noted that the January isotherms were very closely correlated with the gene frequency distribution. Fig. 2 shows the distribution and frequency of the glucoside gene in wild *T. repens* populations superimposed on the January isotherms in Europe and the Near East. The 46.3° F. isotherm passes through Spain, Italy, Greece and Turkey. In all samples which originated from regions near to, or on the warm side of this line, the dominant gene frequency is, in general, high. The proportion of the plants bearing the glucoside gene in the populations averages 94.8 per cent., though it reaches 100 per cent. in many cases. In the region between the 46.3° to 40.0° F. isotherms, while some samples of *T. repens* still show very high frequencies of the dominant gene, the greater number of the populations show an increase in the frequency of the recessive gene, ranging from 0 to 37.4 per cent. This area includes Great Britain, Ireland, the west and south coast of France and the northern part of Turkey. Only a few seed samples were collected in the next temperature belt from the 40.0 to 35.6° F. isotherms. These *T. repens* populations still have a high degree of dominant gene frequency. From the 35.6 to 32.0° F. lines, there is a marked change in the gene frequency pattern. A large number of samples from the Netherlands, Belgium, Germany, Switzerland, Liechtenstein and Denmark were tested and gave ample evidence of this trend towards a high rate of change in the gene frequency structure between these two isotherms. Investigations of

the gene frequency of Belgian and Dutch wild material show a continuous reduction in dominant gene frequency from Brussels (46.7 per cent.), through Breda (31.6 per cent.), to Groningen (9.3 per cent.). A similar pattern is apparent between these two isotherms in Switzerland and Liechtenstein (Lausanne, 62.4 per cent. ; Zürich,



FIG. 2.—Distribution and frequency of the glucoside lotaustralin gene in European and Near Eastern wild populations of *Trifolium repens* L.

Black section : Dominant gene frequency.

White section : Recessive gene frequency.

— January isotherm (refer to the Appendix 1 for the location numbers on the map).

49.4 per cent. ; Schaanwald, 39.4 per cent.). Striking variations were noted in the gene frequency of German populations. The three west German samples possess a high proportion of dominant genes (Hohenheim, 61.8 per cent. ; Frankfurt a/M, 57.7 per cent. ; Giessen, 50.0 per cent.), while the other two samples from locations very near the 32.0° F. isotherm have low dominant gene frequencies (Hanover, 6.4 per cent. ; Schessel, 4.3 per cent.). After crossing the 32.0° F.

isotherm the proportion of dominant genes in wild *T. repens* continues to decrease. This process, however, is much less pronounced (Germany, Straubing, 4.8 per cent. ; Austria, Rupprechtshofen, 6.5 per cent. ; Hungary, Budapest, 1.2 per cent.), than in the previous temperature belt. The 32.0° F. January isotherm occurs in the Po Valley in-

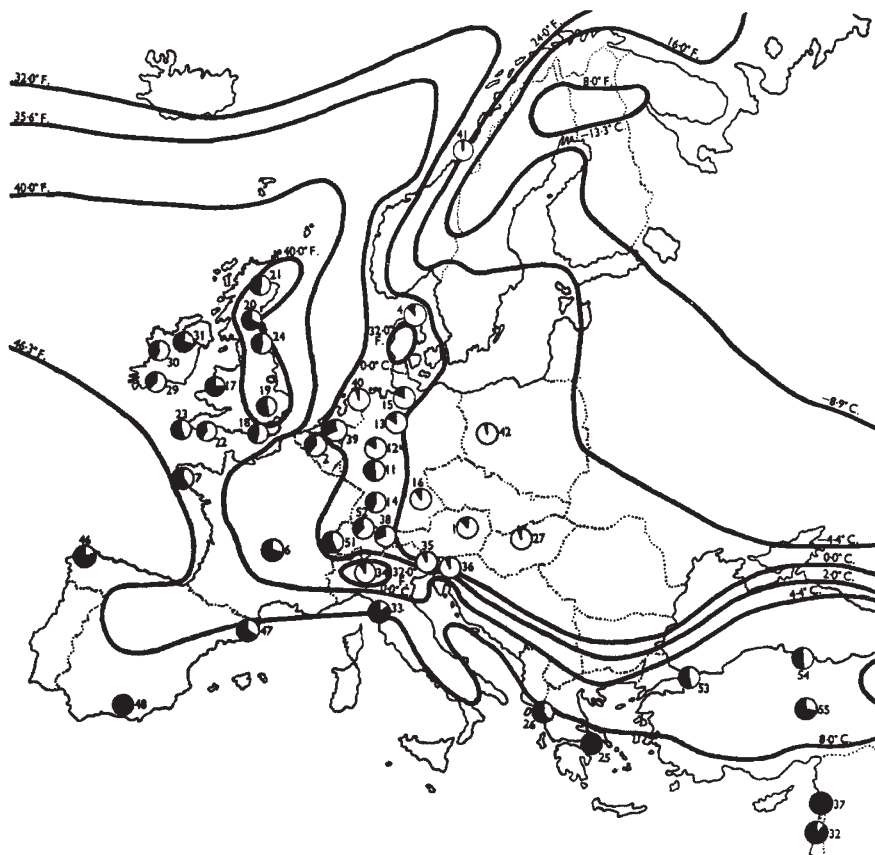


FIG. 3.—Distribution and frequency of the enzyme linamarase gene in European and Near Eastern wild populations of *Trifolium repens* L.

Black section : Dominant gene frequency.

White section : Recessive gene frequency.

— January isotherm (refer to the Appendix 1 for the location numbers on the map).

dependently of the main isotherms across Europe. This low winter temperature seems to explain why the wild populations of the Po Valley and a cultivated strain of Ladino white clover have low percentages of dominant genes and in this respect resemble samples obtained from below the 32.0° F. line in central Europe. The lowest dominant glucoside gene frequency occurs below the 16.0° F. isotherm. No samples were obtained from Russia and Sweden for this present

investigation and the data presented have been taken from the unpublished data of R. D. Williams. According to the results of this previous investigation, in two samples out of four, cyanogenetic plants occur, but the gene frequency is reduced to very low values (U.S.S.R., Leningrad, 1.2 per cent., Orel, 0.4 per cent.). No plants with double dominant genes were found in populations from northern and eastern Europe, including Sweden (Luleå) and U.S.S.R. (Jaroslavl). A similar genetical pattern can be established in the case of the geographical distribution of the enzyme gene frequency which is illustrated in fig. 3.

The variation in the occurrence of the dominant enzyme gene is much more pronounced than that for the glucoside gene above the 40.0° F. isotherm. In the course of the investigation, populations were found with 37.9 to 100 per cent. dominant gene frequencies. The frequency of occurrence of the recessive gene was greater in Great Britain, Ireland and Turkey than in samples from the rest of the region above the 40.0° F. isotherm. In a manner similar to that of the glucoside gene, the enzyme gene composition of populations continues to alter along, and on the colder side of, the 35.6° F. isotherm. From the data relating to the dominant enzyme gene proportions, a rapid reduction towards the 32.0° F. isotherm in Belgian, Dutch, Swiss, Liechtenstein, German and Danish samples is apparent, and this conforms closely with similar reductions found in the proportions of the glucoside gene. Due to the fact that only a limited number of samples were available from the eastern half of Europe, no tests could be carried out on populations outside the 24.0° F. isotherms. Samples obtained from between the 32.0° F. and 24.0° F. isotherms have been shown to maintain the previous trend in the reduction of the dominant enzyme gene frequency and in being closely correlated with decreasing mean winter temperature.

Fig. 4 compares the mean glucoside and enzyme gene frequency of populations originating from regions between different January isotherms. It establishes a reduction in dominant allele frequencies in the successive zones, from high to low temperatures. In zones I, II and III the glucoside gene is more common than the enzyme gene. This is reversed in zone IV, where the mean frequency of the enzyme gene is more than twice as high as that of the glucoside gene. No reliable data were available concerning the enzyme gene frequency in zone V, although a greater frequency of this gene than the corresponding glucoside gene frequency is indicated by the regression coefficient.

In order to test the relationship of January temperature and the gene frequency the regression coefficient was calculated. The *t* test showed that both of the obtained values were highly significant, being $t_{(44)} = 9.00$ in the case of the glucoside gene frequency and $t_{(44)} = 8.98$ in the case of the enzyme gene (P in both cases less than 0.001). Both coefficients are highly significant, which demonstrates

that a close relationship exists between decreasing dominant gene frequencies and decreasing January mean temperatures. From the analyses, it was also apparent that the dominant glucoside gene frequency decreases by 4.23 and the enzyme gene frequency by 3.16 per cent. for each reduction of 1° F. (0.55° C.) January mean temperature. It is also of interest to consider whether the respective rates of decrease in glucoside and enzyme gene frequencies differ. From the analysis of variance of the decreases of the two gene frequencies it appears that the difference between the two regression

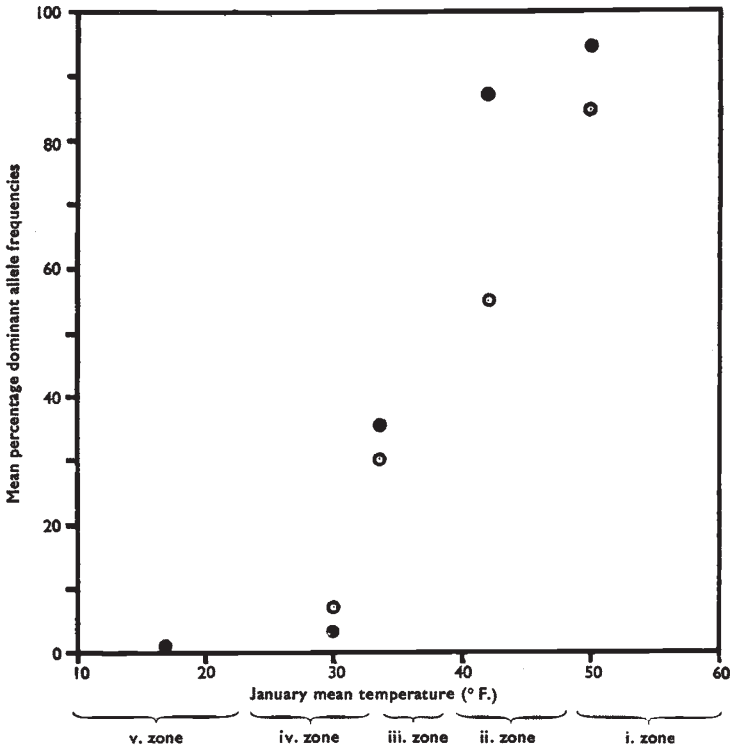


FIG. 4.—Mean frequencies of the glucoside lotaustralin (*Ac*) and enzyme linamarase (*Li*) alleles in different January temperature zones. (● = glucoside allele ; ○ = enzyme allele.)

coefficients is significant ($t_{(88)} = 2.206$ with a probability of between 0.05-0.02), which suggests that the glucoside gene frequency decreases by a significantly greater amount than does the enzyme gene frequency.

4. GEOGRAPHICAL VARIATION AND EVOLUTION

Several authors have discussed the problem of geographical divergence, especially in human and animal populations. Boyd (1939) and Lundman (1948) studied the geographical distribution and frequency of the blood groups in man. Crampton (1916, 1932)

showed the variation of the dextral and sinistral spirals of *Partula suturalis vexillum*. The frequency of white females in the orange race of *Colias chrysotheme* (= *eurytheme*) showed a change from 74 per cent. in the North San Joaquin Valley to 13 per cent. in the Imperial Valley (Hovanitz, 1944). Dobzhansky (1939, 1944) found differences in the incidence of gene arrangements in the third chromosome among populations of *Drosophila pseudoobscura* and *Drosophila persimilis* derived from what was presumably a continuously inhabited region.

The investigations strongly indicate that geographical factors (climate, solar intensity, day-length, etc.) generally play an important role in geographical speciation. It has also been shown that organisms are able to adapt themselves to environmental gradients (*e.g.* latitude, altitude, temperature, etc.) and that natural selection moulds the character of corresponding adaptive clines. This relationship between organisms and their environments is expressed in the rules of Bergmann, Allen and Gloger (Goldschmidt, 1940). These state that there is a definite correlation between body size or pigmentation increases and geographical gradients. Bergmann's rule states that the body size of warm-blooded animals usually increases with decrease in temperature.

Beside the morphological characters, it became evident that the physiological character of species is also subjected to adaptive selection. Timoféeff-Ressovsky (1935) showed in his investigation that *Drosophila funebris* populations derived from western Europe are especially susceptible to both extremes of temperature; the Russian and Siberian ones are particularly resistant, while *D. funebris* from the Mediterranean region is susceptible to cold, but resistant to heat. The investigation of the gene frequency in *T. repens* populations also shows the occurrence of gene frequency clines. The samples of Mediterranean origin possess the highest frequency of dominant glucoside and enzyme genes. A continuous decrease of the dominant gene frequency takes place in western and central Europe, the proportions becoming extremely low and even absent in north eastern Europe. The distribution of the frequency of both chemical characters, the presence or absence of which is governed by these two genes, shows an outstanding correlation with the January isotherms. From this it is apparent that these clines are caused by a geographical factor acting upon European wild *T. repens* populations, resulting in a gene frequency pattern ranging from a high dominant gene frequency to a very low one, depending on the factor pressure prevailing.

The picric acid test provides a means of assessing the amount of glucoside present (Melville *et al.*, 1940), as well as its frequency in the population sample. In Mediterranean samples the fresh leaves were found to contain a high proportion of HCN (more than 0.02 per cent.) whereas a large proportion of the northern European glucoside-containing plants produced very little HCN (0.006-0.02 per cent.).

Such quantitative differences are regarded as being due to the effects of modifying genes (Williams, 1939). Differences also occur in the quantity of enzyme produced. The large proportion of plants in the northern regions with a low output of gene products is probably due to natural selection.

Although no other characters were closely studied in this investigation, several observations indicate variability in other factors in the species. The leaf of *T. repens* usually bears a white marking. It is known that the presence of this character is governed by a single dominant gene (Atwood and Sullivan, 1943), the recessive allele producing no leaf mark. On the basis of a preliminary selection it was possible to distinguish six differently shaped leaf marks. Nearly every plant of Mediterranean *T. repens* has a leaf mark. All six shapes appeared and they were clearly marked on the leaf. The dominant gene was still abundantly evident in central Europe, but the recessive type was found to be more frequent than in the previous region and the leaf mark shape was less pronounced. The Norwegian population contained an increased proportion of the recessive type, with a very faint mark and little variation in shape. Further genetical research is necessary, particularly with regard to the genetical relationship between leaf mark types, before this character can be used in determining geographical gene frequency.

Comparisons of the length of the corolla tubes also disclose marked differences. The Mediterranean *T. repens* appeared to possess uniformly long corolla tubes. In contrast, the Norwegian type has a short corolla tube. *T. repens* is a highly self-sterile species and is generally pollinated by bees. This relationship between the plant and the bee makes an investigation by Alpatov (1929) particularly interesting. He studied the tongue length range of the honey bee in the plain of European Russia. Alpatov found the shortest tongue length in the northern bee populations and that the tongue length gradually increased in populations, as their origin moved southwards. If in *T. repens* populations the corolla tubes were found similarly to vary in size, *i.e.* gradually increasing in length from the shortest in the north to the longest type in the south, such evidence would be of considerable interest and importance, both biologically and practically.

Further differentiation can be mentioned in respect of habit of growth and flowering time. Generally speaking, the southern European *T. repens* appears as an erect type, with large leaves, thick stems and early flowering; the northern European (Norwegian) type, on the other hand, is prostrate with smaller leaves, thinner stems and is late flowering.

Apart from the larger and more general character clines that have been discussed, wild *T. repens* can be divided into several ecotypes, each possessing features adapted to local microgeographical conditions. Previous investigators of geographical variation in the heritable

characters of species have attributed this variation to genetical drift, adaptive selection, migration, isolation, etc.

Before considering the evolutionary mechanism of gene frequency variation in wild *T. repens* populations, the correlation between the decreasing winter temperature and the parallel increasing proportion of recessive genes should be stressed. Modern evolutionary views suggest that only a very low rate of mutation occurrence can be expected, while the importance of natural selection in the case of adaptive geographical variation has been pointed out. In the light of these modern theories, it can be expected that however the recessive mutant type arises in *T. repens* populations, the rate of frequency should be rather low. Because of the adaptive advantage of the carrier of the mutant gene under natural selection, the winter temperature pressure was an effective factor in the case of the widely distributed species *T. repens*. The process of change in the genetic structure of the species presumably continued until an equilibrium was reached between the selection pressure and the proportion of genotypes in *T. repens* in given regions. It is interesting to note that the selection pressure resulted in similar gene frequency clines for both glucoside and enzyme genes. However, the extent of the reaction to the selection pressure was not in all cases the same. In the warmer January temperature zones (zones I, II and III), a larger proportion of mutant recessive enzyme than glucoside genes was brought forward by the selection pressure; by contrast, in zone IV, the recessive glucoside gene showed a significantly greater frequency, compared with the recessive enzyme genes, giving a balance between the gene frequency composition and the environmental gradient.

Further studies of different character clines in the species may reveal other progressive variation clines. The plastic nature of species makes the existence of other such character clines quite probable. These clines may show correlation with some geographical factor other than January isotherms, such as latitude.

The unpublished work of R. D. Williams was concerned with the determination of cyanogenetic plant proportions which correspond to *AcLi* phenotype. His method does not disclose the *Acli* and *acLi* phenotypes and it is therefore not suitable for the purpose of gene frequency calculations. Because no seed samples from Czechoslovakia, Russia and Sweden became available in time for the present investigation, the data of R. D. Williams were used for calculating purposes. Owing to the extremely low gene frequency, the limitation of the test can be neglected.

It may be concluded from the investigation, that in the wild *T. repens* populations there exists a gene frequency cline which is conditioned by the winter temperature prevailing, and that geographical speciation has given rise to several subspecies of wild *T. repens* in Europe and the Near East.

5. SUMMARY

1. The phenotypic and genotypic structures of wild *Trifolium repens* populations were investigated. By means of a modified picric acid test, *T. repens* plants were classified into four phenotypes (1) Glucoside and enzyme (*AcLi*); (2) Glucoside only (*Acli*); (3) Enzyme only (*acLi*) and (4) neither glucoside nor enzyme (*acli*) according to the presence of glucoside lotaustralin (*Ac-ac*) and enzyme linamarase (*Li-li*) genes in dominant or recessive conditions. The genotypic structure was determined by the Hardy-Weinberg formula

$$(p^2 + 2pq + q^2 = 1).$$

2. There was a continuous gradual decrease in the frequencies of glucoside lotaustralin and enzyme linamarase genes over the whole range from 100 to 0 per cent. as the source of the samples of wild *T. repens* populations moved from the Mediterranean region to north-eastern Europe.

3. The distribution of the dominant allele frequencies in the populations was closely correlated with the January isotherms.

4. A decrease of 1° F. in January mean temperature resulted in a reduction of 4.23 per cent. in the frequency of the glucoside gene, and a reduction of 3.16 per cent. in the frequency of the enzyme gene.

5. Thus January temperatures have played an important role through natural selection in the evolution of subspecies in *T. repens*.

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[Continued on p. 78.]

APPENDIX I

Frequency of the four phenotypes in European and Near Eastern wild populations of Trifolium repens (per cent.)

No.	Localities	<i>AcLi</i>	<i>Acli</i>	<i>acLi</i>	<i>acli</i>	No. of plants
1	<i>Austria</i> Rupprechtshofen . . .	7·8	4·8	15·5	71·9	103
2	<i>Belgium</i> Brussels	43·2	28·4	21·0	7·4	81
3	<i>Czechoslovakia</i> Bohemia *	3·1	96·9	96
4	<i>Denmark</i> North Jutland	9·1	13·7	14·7	62·5	197
5	<i>France</i> Cherbourg	85·7	14·3	0·0	0·0	35
6	Clermont	82·8	6·1	9·1	2·0	99
7	Finistère	80·2	15·1	4·7	0·0	106
8	La Rochelle	81·7	18·3	0·0	0·0	60
9	Paris	84·3	15·7	0·0	0·0	19
10	St Brienc	92·8	7·2	0·0	0·0	236
11	<i>Germany</i> Frankfurt a/M	64·8	17·1	11·6	6·5	199
12	Giessen	17·5	57·5	10·0	15·0	40
13	Hanover	5·6	6·8	22·8	64·8	162
14	Hohenheim	60·1	25·3	8·9	5·7	158
15	Schessel	8·3	0·0	25·0	66·7	24
16	Straubing	8·3	1·1	10·4	80·2	96
17	<i>Great Britain</i> Aberystwyth	90·1	5·1	4·0	0·8	640
...	Bangor	58·8	36·5	3·5	1·2	85
18	Brighton	72·1	27·9	0·0	0·0	136
19	Cambridge	79·1	20·9	0·0	0·0	187
20	Edinburgh	85·9	10·9	3·2	0·0	92
21	Nairn	72·5	24·5	1·5	1·5	200
22	Torquay	64·6	33·3	0·0	2·1	48
23	St Ives	73·8	18·6	6·4	1·2	172
24	Sunderland	68·2	27·2	2·9	1·7	173
25	<i>Greece</i> Athens	100·0	0·0	0·0	0·0	193
26	Yanninan	76·5	13·9	7·0	2·6	115
27	<i>Hungary</i> Budapest	1·5	0·8	9·2	88·5	131
28	Magyaróvár	1·2	2·4	96·4	...	83
29	<i>Ireland</i> Cork	58·4	36·8	3·0	1·8	166
30	Galway	59·1	26·9	7·5	6·5	93
31	Longford	90·9	9·1	0·0	0·0	44
32	<i>Israel</i> Huleh District	98·8	0·0	0·0	1·2	85

APPENDIX I—continued

No.	Localities	<i>AcLi</i>	<i>Acli</i>	<i>acLi</i>	<i>acli</i>	No. of plants
<i>Italy</i>						
33	Lucca	97.4	2.6	0.0	0.0	76
34	Milan	1.8	1.7	15.8	80.7	171
35	San Daniele del Friuli	1.2	5.0	12.4	81.4	161
<i>Jugoslavia</i>						
36	Ljubljana	3.0	10.4	9.6	77.0	135
<i>Lebanon</i>						
37	100.0	0.0	0.0	0.0	118
<i>Liechtenstein</i>						
38	Schaanwald	28.7	34.5	23.0	13.8	87
<i>Netherlands</i>						
39	Breda	27.5	25.7	26.6	20.2	109
40	Groningen	0.0	17.7	9.7	72.6	62
<i>Norway</i>						
41	Tjøtta	0.0	6.0	4.0	90.0	50
<i>Poland</i>						
42	Kościan	3.0	1.0	5.0	91.0	100
<i>Russia</i>						
43	Jaroslavl *	0.0	100.0			122
44	Leningrad *	2.7	97.3			111
45	Orel *	0.8	99.2			130
<i>Spain</i>						
46	La Coruña	95.5	4.5	0.0	0.0	89
47	Gerona	87.7	12.3	0.0	0.0	73
48	Granada	100.0	0.0	0.0	0.0	112
<i>Sweden</i>						
49	Luleå *	0.0	100.0			69
50	Stockholm *	1.7	98.3			59
<i>Switzerland</i>						
51	Lausanne	70.6	15.2	10.9	3.3	184
52	Zürich	48.8	25.6	12.2	13.4	82
<i>Turkey</i>						
53	Adapazari	78.4	21.6	0.0	0.0	65
54	Samsun	76.1	23.9	0.0	0.0	113
55	Kayseri	91.8	8.2	0.0	0.0	110

* From the unpublished data of the late R. D. Williams.

APPENDIX II

Frequency of the glucoside and enzyme genotypes in European and Near Eastern wild populations of Trifolium repens

Localities	Glucoside genotypes			Enzyme genotypes		
	p^2	$2pq$	q^2	p^2	$2pq$	q^2
	<i>AcAc</i>	<i>Acac</i>	<i>acac</i>	<i>LiLi</i>	<i>Lili</i>	<i>lili</i>
<i>Austria</i>						
Rupprechtshofen	0·004	0·122	0·874	0·015	0·218	0·767
<i>Belgium</i>						
Brussels	0·218	0·498	0·284	0·162	0·480	0·358
<i>Denmark</i>						
North Jutland	0·015	0·213	0·772	0·016	0·222	0·762
<i>France</i>						
Cherbourg	1·000	0·000	0·000
Clermont	0·445	0·444	0·111	0·512	0·407	0·081
Finistère	0·613	0·340	0·047	0·374	0·475	0·151
La Rochelle	1·000	0·000	0·000
Paris	1·000	0·000	0·000
St Brienc	1·000	0·000	0·000
<i>Germany</i>						
Frankfurt a/M	0·331	0·488	0·181	0·264	0·500	0·236
Giessen	0·250	0·500	0·250	0·022	0·253	0·725
Hanover	0·004	0·120	0·876	0·024	0·260	0·716
Hohenheim	0·382	0·472	0·146	0·197	0·493	0·310
Schessel	0·001	0·082	0·917	0·033	0·300	0·667
Straubing	0·002	0·092	0·906	0·010	0·178	0·812
<i>Great Britain</i>						
Aberystwyth	0·622	0·330	0·048	0·593	0·348	0·059
Bangor	0·613	0·340	0·047	0·149	0·474	0·377
Brighton	1·000	0·000	0·000	0·222	0·499	0·279
Cambridge	1·000	0·000	0·000	0·295	0·496	0·209
Edinburgh	0·671	0·296	0·033	0·449	0·442	0·109
Nairn	0·685	0·285	0·030	0·240	0·500	0·260
Torquay	0·732	0·247	0·021	0·164	0·482	0·354
St Ives	0·525	0·399	0·076	0·308	0·494	0·198
Sunderland	0·616	0·338	0·046	0·214	0·497	0·289
<i>Greece</i>						
Athens	1·000	0·000	0·000	1·000	0·000	0·000
Yanninan	0·477	0·427	0·096	0·352	0·483	0·165
<i>Hungary</i>						
Budapest	0·0001	0·0228	0·9771	0·003	0·104	0·893
<i>Ireland</i>						
Cork	0·609	0·343	0·048	0·144	0·471	0·385
Galway	0·392	0·468	0·140	0·179	0·488	0·333
Longford	1·000	0·000	0·000	0·488	0·421	0·091
<i>Israel</i>						
Huleh District	0·794	0·194	0·012	0·794	0·194	0·012

APPENDIX II—continued

Localities	Glucoside genotypes			Enzyme genotypes		
	p^2	$2pq$	q^2	p^2	$2pq$	q^2
	<i>AcAc</i>	<i>Acac</i>	<i>acac</i>	<i>LiLi</i>	<i>Lili</i>	<i>lili</i>
<i>Italy</i>						
Lucca	1·000	0·000	0·000	0·702	0·272	0·026
Milan	0·0004	0·0347	0·9649	0·008	0·167	0·825
San Daniele del Friuli	0·001	0·061	0·938	0·005	0·132	0·863
<i>Jugoslavia</i>						
Ljubljana	0·005	0·128	0·867	0·004	0·122	0·874
<i>Lebanon</i>						
.	1·000	0·000	0·000	1·000	0·000	0·000
<i>Liechtenstein</i>						
Schaanwald	0·155	0·477	0·368	0·093	0·424	0·483
<i>Netherlands</i>						
Breda	0·100	0·432	0·468	0·104	0·437	0·459
Groningen	0·009	0·169	0·822	0·003	0·094	0·903
<i>Norway</i>						
Tjøtta	0·0009	0·0591	0·9400	0·0004	0·0396	0·9600
<i>Poland</i>						
Kościan	0·0004	0·0396	0·9600	0·002	0·078	0·920
<i>Spain</i>						
La Coruña	1·000	0·000	0·000	0·621	0·334	0·045
Gerona	1·000	0·000	0·000	0·421	0·456	0·123
Granada	1·000	0·000	0·000	1·000	0·000	0·000
<i>Switzerland</i>						
Lausanne	0·390	0·469	0·141	0·325	0·490	0·185
Zürich	0·244	0·500	0·256	0·181	0·429	0·390
<i>Turkey</i>						
Adapazari	1·000	0·000	0·000	0·287	0·498	0·215
Samsun	1·000	0·000	0·000	0·261	0·500	0·239
Kayseri	1·000	0·000	0·000	0·510	0·408	0·082

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