

Tissue-specific activation of mitogen-activated protein kinases for expression of transthyretin by phenylalanine and its metabolite, phenylpyruvic acid

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DOI 10.3858/emm.2010.42.2.012

Accepted 26 November 2009
Available Online 30 November 2009

Abbreviations: C/EBP, CCAAT/enhancer binding protein; HNF, hepatocyte nuclear factors; PAH, phenylalanine hydroxylase; SAPK, stress-activated protein kinase; TTR, transthyretin; Ttr, transthyretin

Abstract

Phenylketonuria is an autosomal recessive disorder caused by a deficiency of phenylalanine hydroxylase. Transthyretin has been implicated as an indicator of nutritional status in phenylketonuria patients. In this study, we report that phenylalanine and its metabolite, phenylpyruvic acid, affect MAPK, changing transthyretin expression in a cell- and tissue-specific manner. Treatment of HepG2 cells with phenylalanine or phenylpyruvic acid decreased transcription of the *TTR* gene and decreased the transcriptional activity of the *TTR* promoter site, which was partly mediated through HNF4 α . Decreased levels of p38 MAPK were detected in the liver of phenylketonuria-affected mice compared with wild-type mice. In contrast, treatment with phenylalanine increased transthyretin expression and induced ERK1/2 activation in PC-12 cells; ERK1/2 activation was also elevated in the brainstem of phenylketonuria-affected mice. These findings may explain between-tissue differences in gene expression, including *Ttr* gene expression, in the phenylketonuria mouse model.

Keywords: hepatocyte nuclear factor 4; mitogen-activated protein kinases; mouse; phenylketonurias; prealbumin

Introduction

Phenylketonuria (MIM 261600) is an autosomal recessive metabolic disorder in which insufficient phenylalanine hydroxylase (PAH; EC 1.14.16.1) activity results in the accumulation of phenylalanine and its metabolites in the blood and other organs. High levels of phenylalanine and its metabolites cause cognitive dysfunction, low brain weight, and changes in myelin structure (Scriver *et al.*, 2001; Martynyuk *et al.*, 2005). As phenylketonuria patients should consume a phenylalanine-restricted diet from birth to prevent brain dysfunction, it is important to monitor their nutritional status (Scriver *et al.*, 2001). The serum level of transthyretin (MIM 176300), formerly called prealbumin, has been implicated as an indicator of the nutritional status of phenylketonuria patients (Shenton *et al.*, 1983; Acosta *et al.*, 1999; Arnold *et al.*, 2002).

Transthyretin is a 55-kDa protein consisting of four identical subunits of 127 amino acids each (Episkopou *et al.*, 1993). Transthyretin is the primary carrier of thyroid hormones and retinol and is mainly synthesized in the liver and choroid plexus (Episkopou *et al.*, 1993). Transthyretin has been associated with amyloidosis, including familial amyloidotic polyneuropathy and Alzheimer's dementia (Costa *et al.*, 1978; Choi *et al.*, 2007; Buxbaum *et al.*, 2008). In addition, transthyretin is involved with many acute-phase proteins when the liver initiates an acute-phase response to acute injury or infection (Wang and Burkner, 2007). As serum transthyretin level has been implicated as an indicator of nutritional status in phenylketonuria patients, a better understanding of the regulation of *TTR* gene expression by phenylalanine and its metabolites is essential (Shenton *et al.*, 1983; Arnold *et al.*, 2002).

The *Ttr* gene is transcriptionally regulated by two DNA regions: a proximal -150 to -90 bp promoter region and a distal 100-nucleotide enhancer located -2 kb upstream of the mRNA cap site (Samadani and Costa, 1996). The *Ttr* promoter and enhancer regions have been identified in experiments with transgenic mice, and have DNA binding sites for four hepatocyte nuclear factors (HNFs), CCAAT/enhancer binding protein (C/EBP), and AP-1/cJun (Costa and Grayson, 1991). More

specifically, the *Ttr* proximal promoter region has binding sites for HNF1, HNF3, HNF4, HNF6, and AP-1. The *Ttr* enhancer region has binding sites for HNF3, HNF4, and C/EBP.

The mouse model of phenylketonuria (BTBR-*Pah*^{enu2}) has a homozygous F263S mutation of the *Pah* gene, which is created by an alkylating agent, *N*-ethyl-*N*-nitrosourea (McDonald *et al.*, 1990; McDonald and Charlton, 1997). The phenotype of the phenylketonuria mouse model closely resembles that of phenylketonuria patients: hypopigmentation, elevated serum and brain phenylalanine levels, and neurological abnormalities (McDonald *et al.*, 1990; Surendran *et al.*, 2004). Previously, we found that *Ttr* gene expression was downregulated in the liver and serum of phenylketonuria-affected mice compared with wild-type mice, although all mice were given *ad libitum* access to a diet containing phenylalanine (Park *et al.*, 2009). In this report, we describe the mechanism by which phenylalanine and its metabolite, phenylpyruvic acid, regulate transcription of the *Ttr* gene in phenylketonuria.

Results

Phenylalanine and phenylpyruvic acid treatment of HepG2 cells reduces *TTR* mRNA and inhibits the activity of the *TTR* gene promoter

We performed real-time PCR analysis to determine whether the *TTR* mRNA level is altered by phenylalanine or phenylpyruvic acid treatment. Exposure of HepG2 cells to 0.9 mM phenylalanine reduced *TTR* transcription by $24.8 \pm 6.5\%$ after 3 h ($P < 0.01$), by $13.5 \pm 6.5\%$ after 6 h ($P < 0.05$), and by $38.3 \pm 6.6\%$ after 18 h compared with the control ($P < 0.01$; Figure 1A). Treatment of HepG2 cells with 1.2 mM phenylpyruvic acid resulted in a similar suppression of *TTR* transcription, except for a shift in the time course: transcription decreased to $66.6 \pm 11.9\%$ of that of the control after 15 min, and maximal repression of transcription was observed after 1 h ($P < 0.001$; Figure 1B).

To assess the effects of phenylalanine and phenylpyruvic acid on *Ttr* promoter-mediated transactivation potential, a luciferase assay using a luciferase reporter containing a mouse *Ttr* promoter site was performed. As phenylalanine and phenylpyruvic acid dissolve in PBS and DMSO, respectively, PBS- and DMSO-treated cells were used as controls. Exposure of HepG2 cells to 0.9 mM phenylalanine decreased luciferase activity by $38.2 \pm 24.9\%$ after 6 h ($P < 0.05$) and by $46.2 \pm 18.9\%$ after 18 h compared with the control ($P < 0.01$; Figure 1C). Luciferase activity decreased

after 3 h and 6 h of 1.2 mM phenylpyruvic acid treatment (Figure 1D). Maximal repression occurred after 3 h, when luciferase activity was $53.1 \pm 22.5\%$ of that of the control ($P < 0.01$; Figure 1D). Increments in the concentration of phenylalanine induced marked decreases in luciferase activity in a dose-dependent manner (Figure 1E). In contrast, a small amount of phenylpyruvic acid (0.5 mM) resulted in a decrease in luciferase activity similar to that obtained with a greater amount of phenylpyruvic acid ($P < 0.05$; Figure 1F). These findings suggest that modification of *TTR* gene transcription by phenylalanine and phenylpyruvic acid is mediated through the *TTR* promoter region.

Phenylalanine and phenylpyruvic acid decrease the level of HNF4 α protein in HepG2 cells, and the level of p38 MAPK is low in the liver of phenylketonuria-affected mice

To identify transcription factors involved in phenylalanine- and phenylpyruvic acid-mediated *TTR* gene alteration, we analyzed protein levels of transcription factors that bind to the *TTR* promoter region. HNF4 α nuclear protein level in HepG2 cells transiently decreased to $47.3 \pm 13.0\%$ of that of the control after 1 h of 0.9 mM phenylalanine treatment ($P < 0.05$; Figure 2A). The HNF4 α nuclear protein level in HepG2 cells started to decrease after 5 min of 1.2 mM phenylpyruvic acid treatment and was $61.6 \pm 2.8\%$ of that of the control after 30 min ($P < 0.01$; Figure 2B). Treatment of HepG2 cells with 0.9 mM phenylalanine or 1.2 mM phenylpyruvic acid did not alter nuclear protein levels of HNF1, HNF3 α/β , C/EBP α , or C/EBP β (data not shown).

As MAPK has been reported to play an important role in the regulation of HNF4 α activity (Hatzis *et al.*, 2006; Li *et al.*, 2006; Xu *et al.*, 2007), we analyzed MAPK alterations in the liver of the phenylketonuria mouse model. Total levels of p38 MAPK protein in liver tissue were significantly lower in homozygous *Pah*^{enu2} mice than in wild-type mice: those of 5-week-old male homozygous mice were $52.2 \pm 12.5\%$ lower ($P < 0.01$; Figure 3A); those of 30-week-old male homozygous mice were $32.1 \pm 3.2\%$ lower ($P < 0.01$; Figure 3B); and those of 43-week-old female homozygous mice were $36.3 \pm 14.9\%$ lower ($P < 0.05$; Figure 3C). JNK/SAPK and ERK1/2 phosphorylation was not detected in liver tissues of the phenylketonuria mouse model (data not shown). We also monitored p38 MAPK and ERK1/2 activation in HepG2 cells treated with 0.9 mM phenylalanine or 1.2 mM phenylpyruvic acid. However, the phosphorylated form of p38 MAPK was

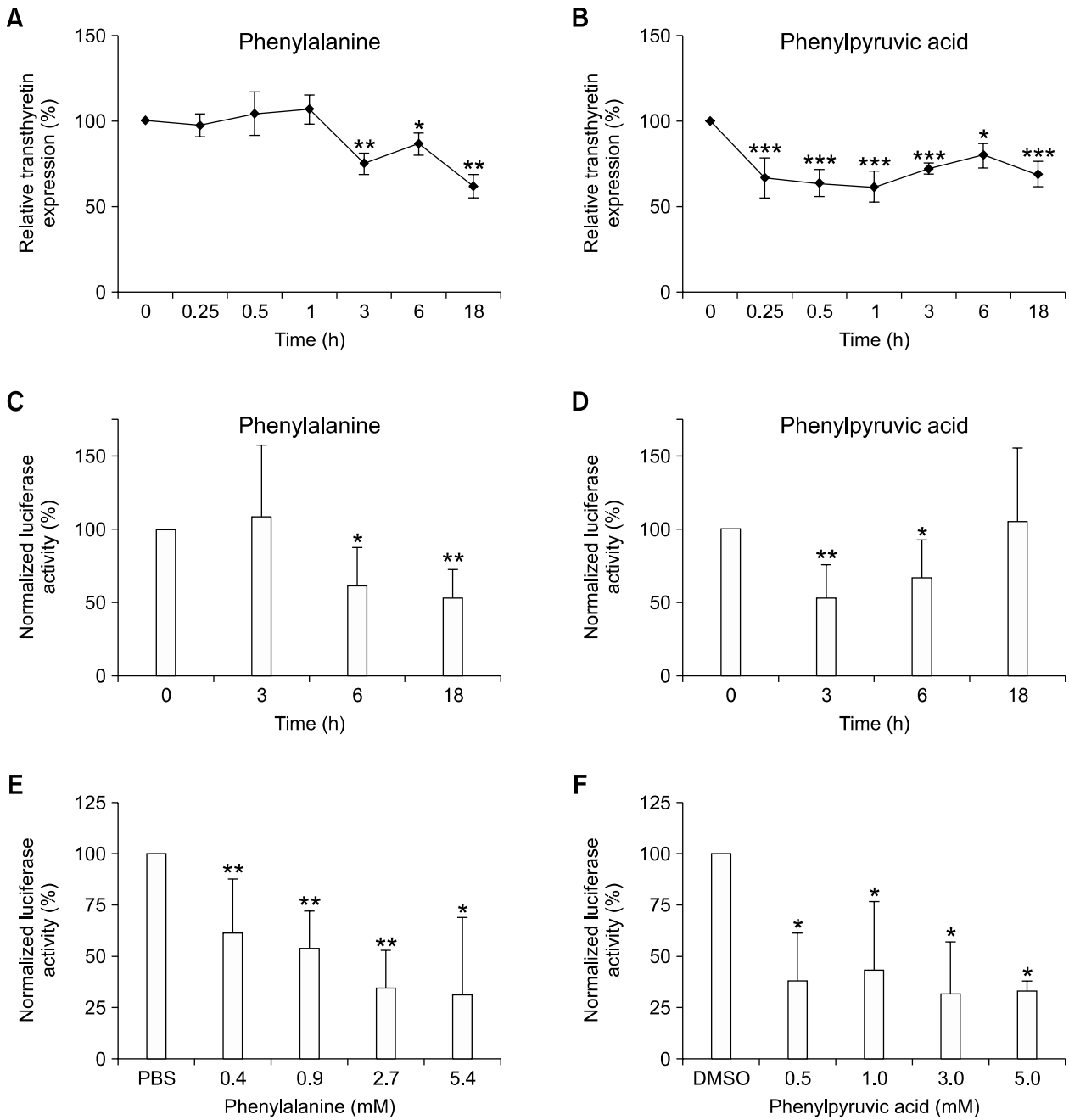


Figure 1. Real-time PCR analysis showing decreased *TTR* gene transcription in HepG2 cells treated with 0.9 mM phenylalanine (A) or 1.2 mM phenylpyruvic acid (B). HepG2 cells were transfected with a reporter plasmid containing the mouse *Ttr* promoter region (C-F). Twenty-four hours after transfection, 0.9 mM phenylalanine (C) or 1.2 mM phenylpyruvic acid (D) were added for the indicated times. To determine whether the effects of phenylalanine and phenylpyruvic acid were dose dependent, various concentrations of phenylalanine (E) or phenylpyruvic acid (F) were added. The cells were lysed, and firefly and *Renilla* luciferase activities were measured. Reporter activity was calculated as the ratio of firefly luciferase activity to *Renilla* luciferase activity in the same sample. Values are expressed as a percentage of the reporter activity relative to that of control cells, and are plotted as the mean \pm SD (n = 4-6). * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs control cells using Student's *t*-test.

not detected in HepG2 cells (data not shown). ERK2 inactivation occurred transiently in HepG2 cells after the addition of phenylalanine or phenylpyruvic acid (Figure 4).

ERK1/2 is activated in the brainstem of phenylketonuria-affected mice

As brainstem transthyretin expression was previously shown to be elevated in homozygous mice,

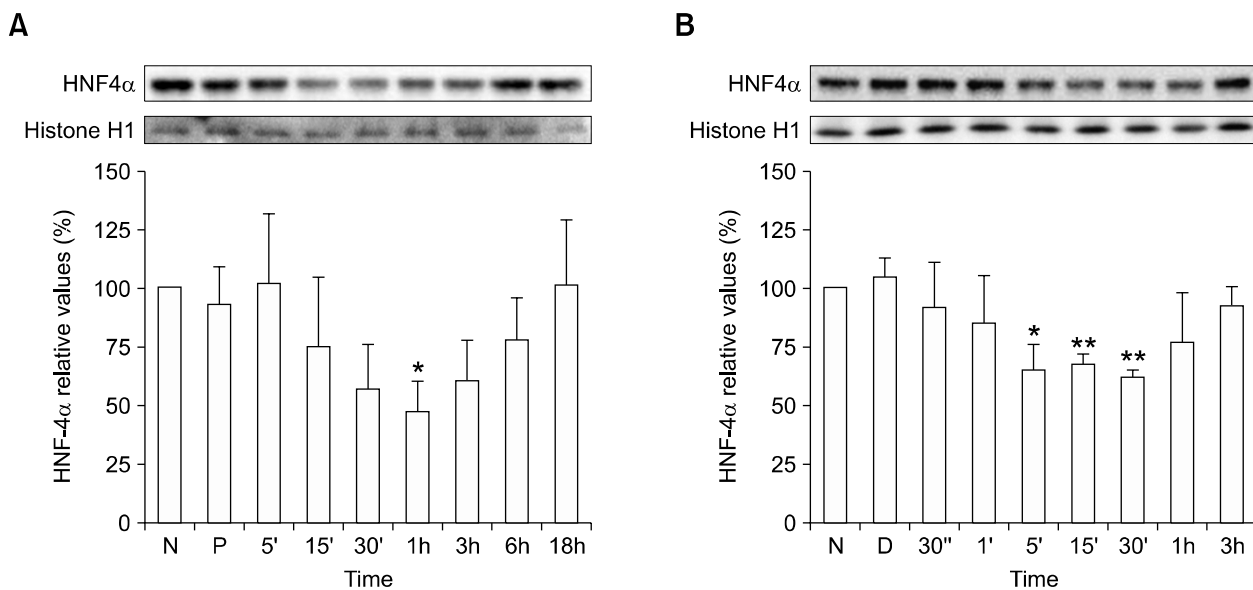


Figure 2. HNF4 α nuclear protein levels in HepG2 cells treated with 0.9 mM phenylalanine (A) or 1.2 mM phenylpyruvic acid (B). Values are expressed as percentages of values for untreated cells (N) and are plotted as the mean \pm SD ($n = 3$). * $P < 0.05$, ** $P < 0.01$ vs (A) PBS-treated control cells (P) or vs (B) DMSO-treated control cells (D) using Student's t -test.

we examined MAPK alterations in brain tissues of the phenylketonuria mouse model (Park *et al.*, 2009). ERK1/2 phosphorylation was elevated in the brainstem of heterozygous and homozygous mice compared with wild-type mice: by $100.0 \pm 30.1\%$ in heterozygous mice and by $207.2 \pm 83.5\%$ in homozygous mice ($P < 0.05$; Figure 5C). Although the level of the phosphorylated form of ERK1/2 increased greatly in the cerebellum of homozygous mice, the increase was not statistically significant (Figure 5B). ERK1/2 phosphorylation levels in the cerebrum were not statistically significant between groups (Figure 5A). JNK/SAPK and p38 kinase phosphorylation levels in the brain were not statistically significant between groups (data not shown).

Phenylalanine-induced elevation of transthyretin expression in PC-12 cells is ERK1/2 dependent

We investigated whether phenylalanine and its metabolite, phenylpyruvic acid, affect ERK1/2 phosphorylation in PC-12 cells. Phosphorylated ERK1/2 protein levels increased transiently 5 min after treatment with 0.9 mM phenylalanine: by $390.4 \pm 118.2\%$ after 5 min ($P < 0.01$) and by $307.0 \pm 142.0\%$ after 15 min ($P < 0.05$; Figure 5D). ERK1/2 phosphorylation in PC-12 cells was not altered by treatment with 1.2 mM phenylpyruvic acid (Figure 5E). Exposure of PC-12 cells to 0.9 mM phenylalanine increased transthyretin expression by $38.6 \pm 19.1\%$ after 7 days ($P < 0.05$;

Figure 5E) (Park *et al.*, 2009). To examine the effect of ERK1/2 phosphorylation on transthyretin expression, PC-12 cells were pretreated with an ERK1/2 inhibitor. After pretreatment with ERK1/2 inhibitor II, the addition of 0.9 mM phenylalanine did not increase transthyretin expression in PC-12 cells (Figure 5E). The selective ERK1/2 inhibitor II decreased transthyretin expression in PC-12 cells regardless of treatment with 0.9 mM phenylalanine or 1.2 mM phenylpyruvic acid (Figure 6A, B). Transthyretin expression did not increase after treatment with 1.2 mM phenylpyruvic acid (Figure 6B).

Phenylalanine and phenylpyruvic acid may affect *TTR* transcription independently

The effect of phenylpyruvic acid on suppression of *TTR* transcription in HepG2 cells was similar to that of phenylalanine, except for a shift in the time course. As phenylalanine may be transformed into phenylpyruvic acid, we hypothesized that phenylpyruvic acid, not phenylalanine, suppresses *TTR* transcription directly in HepG2 cells. We monitored levels of phenylpyruvic acid using GC/MS to determine whether phenylalanine was transformed into phenylpyruvic acid. Phenylpyruvic acid was not detected in phenylalanine-treated HepG2 cells (data not shown). As the PAH enzyme level was normal in HepG2 cells, the phenylalanine that was added may have been metabolized to tyrosine. Phenylpyruvic acid was detected in phenylalanine-

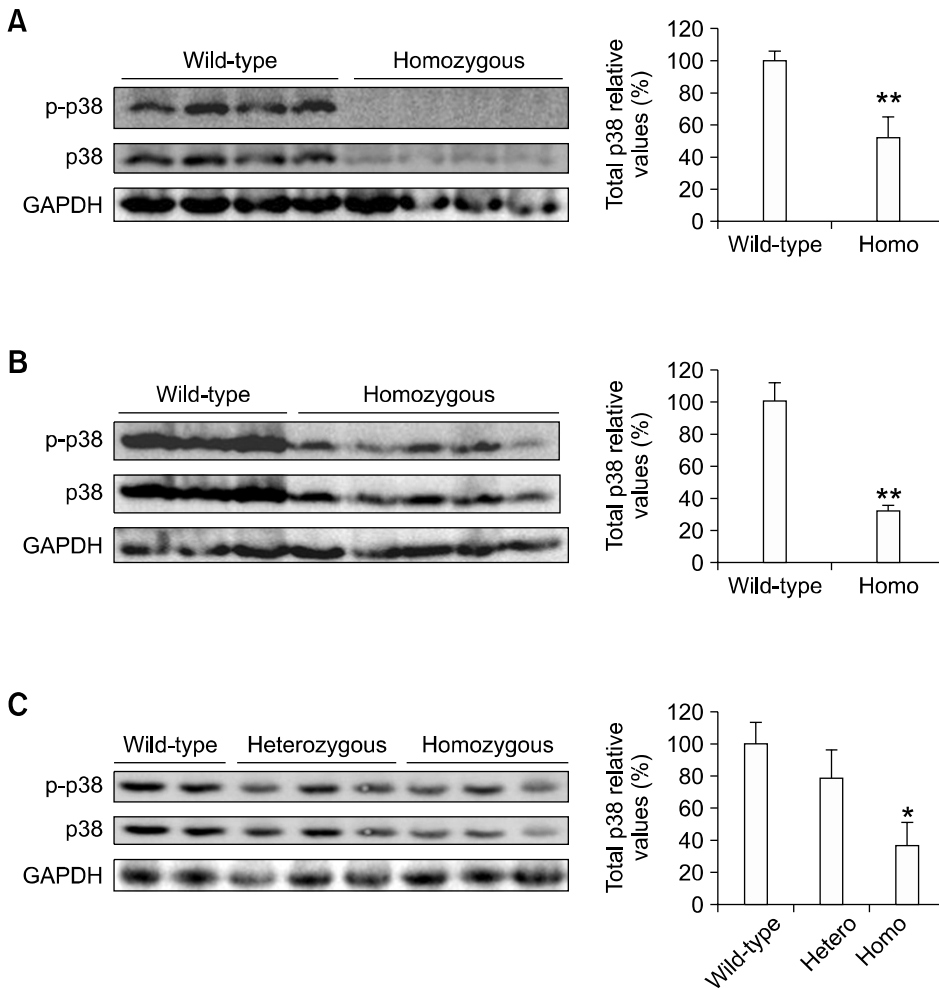


Figure 3. Western blotting was performed to detect p38 kinase expression in the liver of the phenylketonuria mouse model (BTBR-*Pah*^{enu2}): (A) 5-week-old male mice, (B) 30-week-old male mice, and (C) 43-week-old female mice. Levels of p38 kinase protein in liver were quantified using a bio-imaging analyzer. Values are expressed as the mean \pm SD ($n = 2-5$). * $P < 0.05$, ** $P < 0.01$ vs wild-type mice using Student's *t*-test.

treated PC-12 cells, which lack PAH enzyme activity (Figure 7A). However, the level of phenylpyruvic acid in phenylalanine-treated PC-12 cells was low compared with that in phenylpyruvic acid-treated PC-12 cells (Figures 7A and 7B). Phenylpyruvic acid in PC-12 cells reached a maximum concentration after 1 h of 0.9 mM phenylalanine treatment (Figure 7A), which is much longer than that required for phenylalanine to affect MAPK. These findings suggest that phenylalanine and phenylpyruvic acid affect *TTR* transcription independently.

Discussion

Previously, we detected elevated transthyretin expression in the brain and decreased transthyretin expression in the serum and liver of phenylketonuria-affected mice using a *Pah*^{enu2} phenylketonuria mouse model (Park *et al.*, 2009). In addition, the transthyretin level decreased in

HepG2 cells after treatment with 0.9 mM phenylalanine, a concentration typical of that in the plasma of phenylketonuria patients (Bedin *et al.*, 2000; Park *et al.*, 2009). The aim of this study was to investigate the mechanisms of differential transthyretin expression in the brain and liver of phenylketonuria-affected mice using cell lines. We used PC-12 cells as neuronal cells and HepG2 cells as liver cells. A deficiency of the hepatic enzyme, PAH, which converts phenylalanine to tyrosine, results in phenylketonuria. As phenylalanine cannot be oxidized to tyrosine in phenylketonuria, transamination of phenylalanine produces phenylpyruvic acid, which is further metabolized to phenyllactic acid and phenylacetic acid (Sarkissian *et al.*, 2000). We used real-time PCR to examine the effect of phenylalanine and its metabolites on *TTR* transcription in HepG2 cells. Phenyllactic acid and phenylacetic acid did not alter *TTR* transcription significantly (data not shown). Only phenylalanine and phenylpyruvic acid induced a significant decrease in *TTR*

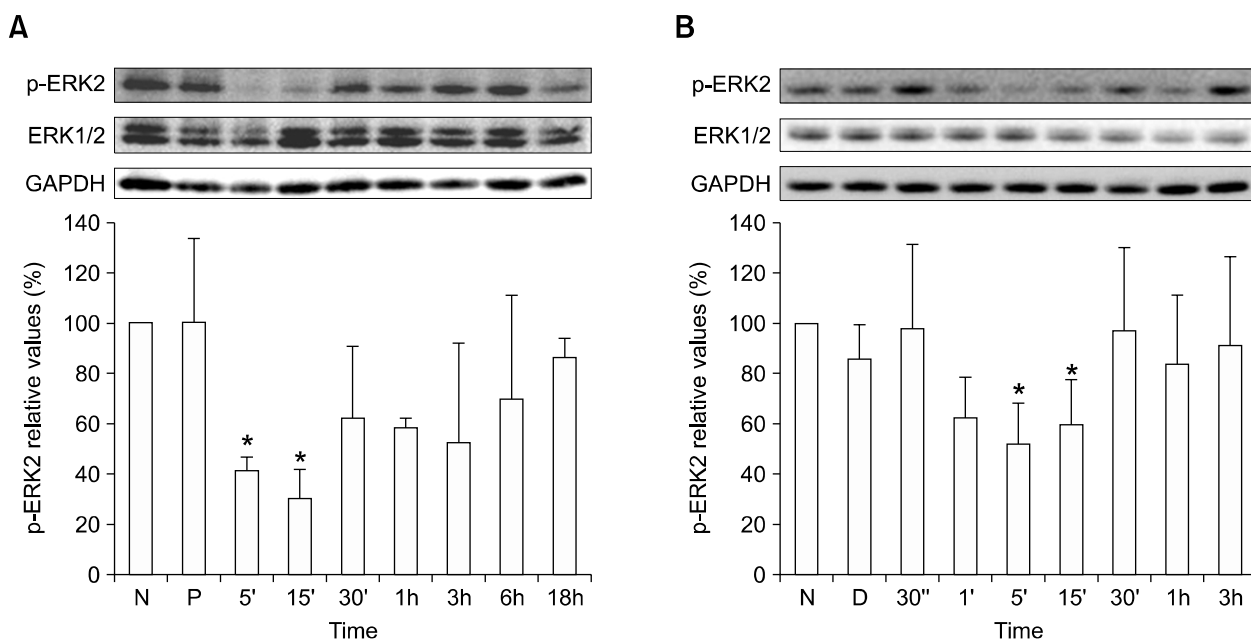


Figure 4. Western blotting analysis of HepG2 cells treated with 0.9 mM phenylalanine or with 1.2 mM phenylpyruvic acid. Whole cell lysates of HepG2 cells treated with 0.9 mM phenylalanine (A) or with 1.2 mM phenylpyruvic acid (B) were used for detection of phosphorylated Erk2 and total Erk1/2. Values are expressed as percentages of values for untreated cells (N) and are plotted as mean \pm SD ($n = 3$). * $P < 0.05$ vs (A) PBS treated control cells (P) or vs (B) DMSO treated control cells (D) using Student's t -test.

transcription. The luciferase analysis with reporter plasmids inserted into the mouse *Ttr* promoter region revealed that modification of *TTR* gene transcription by phenylalanine and phenylpyruvic acid was mediated through the *TTR* promoter region. Of the transcription factors that bind to the *TTR* promoter region, the HNF4 α nuclear protein level decreased. Reporter plasmids inserted into the mouse *Ttr* promoter region were used for luciferase analysis in HepG2 cells. The expression of a mouse gene in a human hepatoma cell line was successful because there is a high level of conservation between the mouse and human genes in the 190-nucleotide stretch just upstream from the cap site, and the general species barrier does not exist for liver-specific gene expression in HepG2 cells (Costa *et al.*, 1989; Friedman *et al.*, 1986).

A key role of HNF4 α in the regulation of *TTR* gene transcription has been reported, and over-expression of HNF4 α protein causes dose-dependent upregulation of *TTR* reporter gene transcription (Wang and Burkner, 2007). In our study, nuclear HNF4 α protein levels decreased in HepG2 cells treated with phenylalanine or phenylpyruvic acid. The phenylalanine- and phenylpyruvic acid-induced decreases in HNF4 α level may have led to a decrease in *TTR* gene transcription in HepG2 cells. As we did not examine the effect of

phenylalanine and its metabolites on the *TTR* enhancer region, this remains to be validated.

Several studies have suggested that there is a link between MAPK and HNF4 α (Hatzis *et al.*, 2006; Li *et al.*, 2006; Xu *et al.*, 2007). Inhibition of p38 kinase activity decreases HNF4 α nuclear protein levels and its phosphorylation status *in vivo* and *in vitro*, which makes the protein less stable (Xu *et al.*, 2007). Awad *et al.* (2000) and Mendelson *et al.* (1996) reported that p38 MAPK is constitutively active in the liver of adult mice, which is in accordance with our results. In our study, the p38 kinase protein level was significantly decreased in the liver of phenylketonuria-affected mice regardless of age or sex. As the total p38 kinase protein level and the level of the phosphorylated form of p38 kinase protein were greatly decreased in the liver of phenylketonuria-affected mice, this may have decreased HNF4 α nuclear protein levels in phenylketonuria-affected mice. Suppression of HNF4 α nuclear protein levels would diminish *TTR* gene transcription because HNF4 α positively regulates *TTR* gene expression (Hatzis *et al.*, 2006). ERK2 activation downregulates HNF4 α expression in HepG2 cells at the transcriptional level via a mechanism that involves the loss of C/EBP α expression and the concomitant disruption of the HNF4 enhancer-promoter complex (Hatzis *et al.*, 2006). ERK2

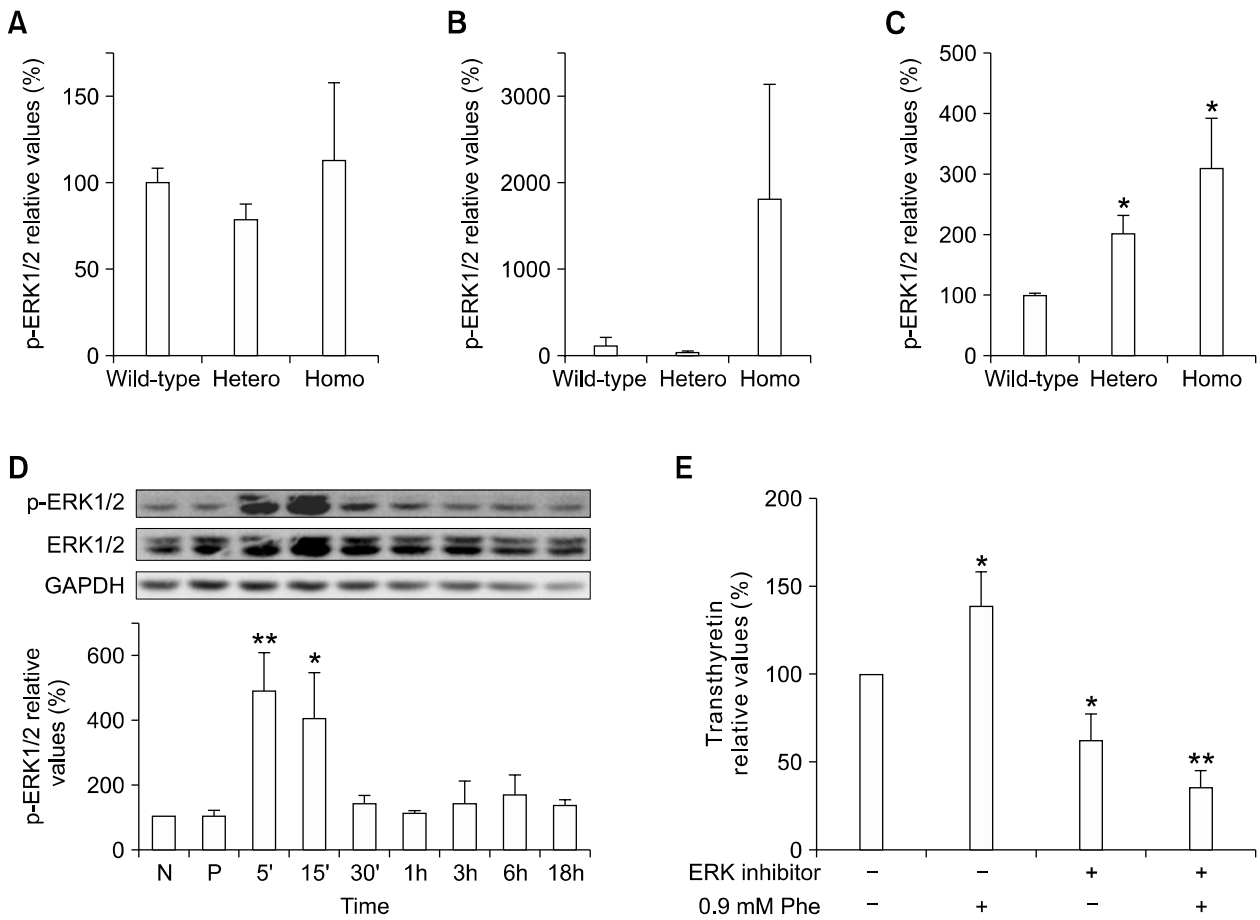


Figure 5. Western blotting was performed to detect ERK1/2 expression levels in the brain of the phenylketonuria mouse model (BTBR-*Pah*^{enu2}). ERK1/2 protein expression in (A) the cerebrum, (B) the cerebellum, and (C) the brain stem was demonstrated and quantified using a bioimaging analyzer. Whole cell lysates of PC-12 cells treated with (D) 0.9 mM phenylalanine were used for detection of phosphorylated ERK1/2 and total ERK1/2. Western blotting was performed to detect transthyretin expression levels in PC-12 cells treated with (E) 0.9 mM phenylalanine after preincubation with ERK inhibitor II. The cells were exposed to phenylalanine for 7 days. Transthyretin levels were normalized to GAPDH levels. Values are expressed as the mean \pm SD ($n = 3-4$). * $P < 0.05$, ** $P < 0.01$ vs wild-type mice or DMSO- and/or PBS-treated controls using Student's *t*-test.

inactivation occurred transiently in HepG2 cells after the addition of phenylalanine or phenylpyruvic acid (Figure 4). As ERK2 activation decreases HNF4 expression, ERK2 inactivation may have increased HNF4 α expression. However, neither phenylalanine treatment nor phenylpyruvic acid treatment altered C/EBP α nuclear protein levels in HepG2 cells (data not shown). In addition, HNF4 α nuclear protein levels decreased after addition of phenylalanine or phenylpyruvic acid (Figure 2). Thus, altered ERK2 activation after addition of phenylalanine or phenylpyruvic acid may not have directly affected HNF4 α expression in our study. Although we also performed electrophoretic mobility shift assays, we did not detect any alterations in the binding affinity of transcription factors, including HNF4 α , in HepG2 cells treated with phenylalanine or phenylpyruvic acid (data not shown). Although phenylalanine and phenylpyruvic

acid did not affect the binding affinity of HNF4 α to the *TTR* promoter region, the decreased amount of HNF4 α *per se* may have suppressed *TTR* gene transcription.

ERK1/2 activation was detected in the brainstem of 43-week-old homozygotes. In our previous study, increased transthyretin protein expression was detected in the brainstem of 30-week-old homozygotes (Park *et al.*, 2009). To examine correlations between phenylketonuria, ERK1/2 activation, and transthyretin expression, PC-12 cells were treated with phenylalanine and phenylpyruvic acid. Although the PC-12 cell line does not represent brain tissue, it was used as one of the neuronal cell lines. Only phenylalanine, not phenylpyruvic acid, activated ERK1/2 in PC-12 cells. Exposure of PC-12 cells to phenylalanine increased transthyretin expression (Park *et al.*, 2009). However, the addition of phenylalanine did

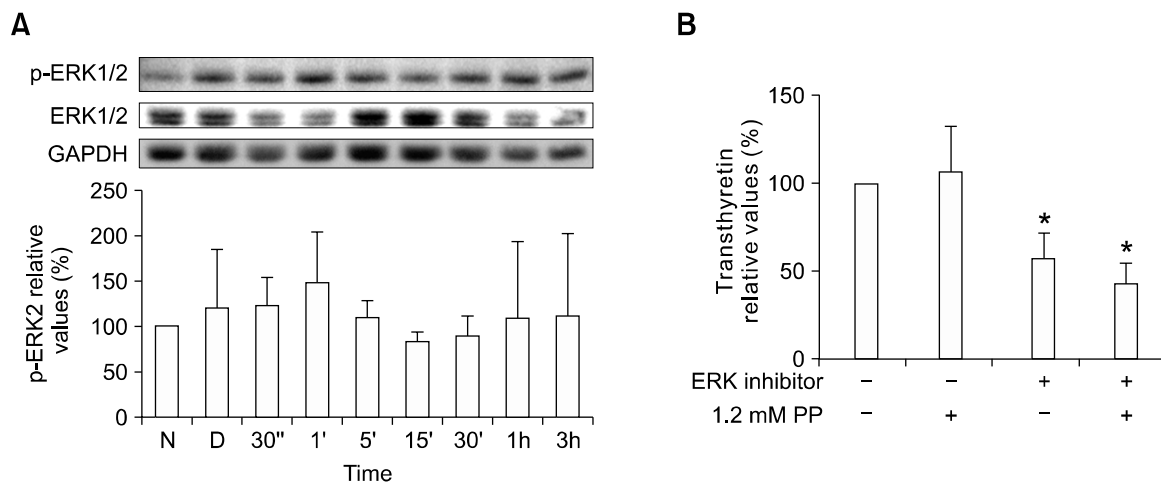


Figure 6. Whole cell lysates of PC-12 cells treated with (A) 1.2 mM phenylpyruvic acid were used for detection of phosphorylated ERK1/2 and total ERK1/2. Western blot analysis was performed to detect transthyretin expression levels in PC-12 cells treated with (B) 1.2 mM phenylpyruvic acid after preincubation with ERK inhibitor II. The cells were exposed to phenylpyruvic acid for 7 days. Transthyretin levels were normalized to GAPDH levels. Values are expressed as the mean \pm SD ($n = 3-4$). * $P < 0.05$ vs DMSO-treated controls using Student's *t*-test.

not increase transthyretin expression in PC-12 cells after pretreatment with ERK1/2 inhibitor II. Thus, the elevated transthyretin expression in the brainstem of phenylketonuria-affected mice was probably the result of ERK1/2 activation induced by high brain phenylalanine levels. Of the transcription factors that bind to the *TTR* promoter region, only C/EBP and AP-1 exist in neuronal cells (Kuo *et al.*, 1990). Thus, ERK1/2 activation by phenylalanine may have affected C/EBP and/or AP-1, which may have affected transthyretin expression in the brainstem of phenylketonuria-affected mice. The effect of phenylalanine and phenylpyruvic acid on

C/EBP and AP-1 in the brain remains to be elucidated. Transthyretin has been associated with familial amyloidotic polyneuropathy (FAP), and ERK1/2 activation is upregulated in human FAP nerves and Schwann cells, whereas p38 and JNK MAPK activation is not detected in human FAP nerves and Schwann cells (Costa *et al.*, 1978; Monteiro *et al.*, 2006). Our data corroborate the results of Monteiro *et al.* (2006).

Serum transthyretin level has been implicated as an indicator of nutritional status in phenylketonuria patients (Shenton *et al.*, 1983; Arnold *et al.*, 2002). As phenylalanine and its metabolite, phenylpyruvic

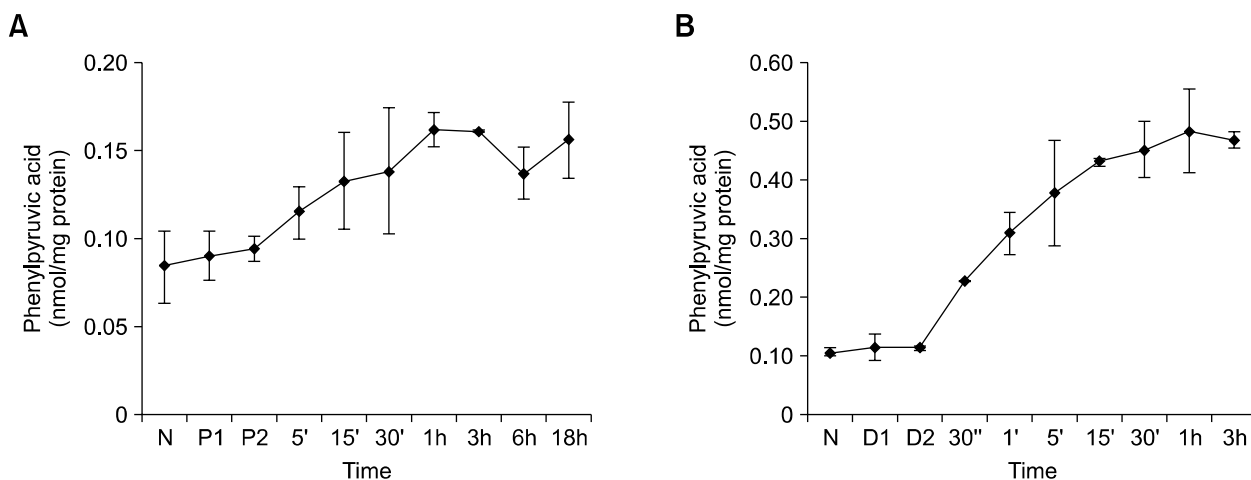


Figure 7. Phenylpyruvic acid levels in PC-12 cells treated with (A) 0.9 mM phenylalanine or (B) 1.2 mM phenylpyruvic acid assessed using GC/MS. Values are expressed as percentages of values for untreated cells (N) and are plotted as the mean \pm SD ($n = 2$). P1 and P2 indicate PBS-treated control cells after 1 h and 6 h, respectively. D1 and D2 indicate DMSO-treated control cells after 1 min and 1 h, respectively.

acid, decrease transthyretin expression in HepG2 cells, the use of serum transthyretin level as a nutritional indicator for phenylketonuria patients may be limited; further studies are necessary to elucidate this problem. In addition, further studies should be performed to validate the function of transthyretin in the phenylketonuria-affected brain.

In conclusion, our findings indicate that phenylalanine and its metabolite, phenylpyruvic acid, affect MAPK activation, resulting in tissue-specific transthyretin expression in a phenylketonuria mouse model. It has been suggested that the presence of cell-specific transcriptional factors would explain cell-specific regulation (Costa *et al.*, 1989). However, the differential regulation of MAPK in specific cells or tissues remains to be elucidated. This may explain tissue-specific gene expression, including that of the *TTR* gene, and the tissue-specific vulnerability of the phenylketonuria mouse model.

Methods

Animals

The animal protocol was approved by the Animal Experiment Review Committee of the Ewha Medical Research Institute, Ewha Womans University, Korea. All animals were treated in accordance with the Animal Care Guidelines of the Ewha Womans University School of Medicine and the National Institutes of Health's Guidelines for Animal Care. The *Pah*^{enu2} phenylketonuria mice were obtained from the Jackson Laboratory (Bar Harbor, ME). The *Pah*^{enu2} phenylketonuria mouse contains a T-to-C transition in exon 7 of the *PAH* gene (McDonald *et al.*, 1997). The genotype of each mouse was confirmed by genotyping. Mice were provided with water and given *ad libitum* access to a diet containing phenylalanine.

Cell culture

HepG2 and PC-12 cells were grown in DMEM (Gibco-BRL, Gaithersburg, MD) supplemented with 100 U/ml of penicillin, 100 µg/ml of streptomycin, and 10% (v/v) of heat-inactivated FBS (Gibco-BRL) at 37°C in a humidified atmosphere containing 5% CO₂. The medium was replaced with serum-free medium 6 h before the experiment. Each cell line was treated with 0.9 mM L-phenylalanine (Sigma-Aldrich, Madison, WI) or 1.2 mM phenylpyruvic acid (Fluka, Milwaukee, WI) in serum-free medium for the intervals indicated in the figures. A phenylalanine concentration of 0.9 mM is typical of that in the plasma of phenylketonuria patients, and a phenylpyruvic acid concentration of 1.2 mM is greater than that in the plasma of phenylketonuria patients (< 1 mM) (Bedin *et al.*, 2000; Berti *et al.*, 2001). For ERK inhibitor trials, cells were preincubated for 1 h with 10 µM of the ERK inhibitor II, FR180204 (Calbiochem, San Diego, CA), dissolved in DMSO.

Real-time PCR

Real-time PCR was performed using the SYBR Green system (Applied Biosystems, Warrington, UK) as previously described (Park *et al.*, 2009). GAPDH was used as the endogenous control gene. The relative between-group expression of the *Ttr* gene was calculated as $2^{-\Delta\Delta Ct}$ (Livak and Schmittgen, 2001).

Reporter plasmid construct

A 356 bp mouse *Ttr* gene promoter containing *Bgl*II and *Hind*III site adapters was subcloned into the corresponding restriction sites of the luciferase reporter pGL3-basic vector (Promega, San Luis Obispo, CA). The subcloned sequences were verified by DNA sequence analyses.

Reporter gene transfection assays: luciferase assay

The luciferase assay was carried out using the Dual-Luciferase Reporter assay system (Promega, San Luis Obispo, CA) as previously described (Wang and Burkner, 2007). Reporter plasmids containing the *Ttr* promoter region were transfected into subconfluent HepG2 cells using FuGENE HD transfection reagent (Roche Applied Science, Indianapolis, IN). To normalize the transfection efficiency, the pRL-cytomegalovirus promoter (*Renilla* luciferase vector, Promega) was cotransfected into each well. Each well of a 24-well culture plate received 1 µg of DNA mixture consisting of firefly reporters and *Renilla* control vector in the ratio of 40:1. After 24 h, each cell was treated with phenylalanine, phenylpyruvic acid, PBS, or DMSO. Cells were lysed using Promega lysis buffer. Firefly and *Renilla* luciferase activities were measured sequentially in the same sample using a Veritas microplate luminometer (Turner BioSystems, Sunnyvale, CA). Normalized luciferase activities were calculated by dividing the firefly luciferase activity by the *Renilla* luciferase activity. Relative luciferase activity is expressed as a percentage of the reporter activity relative to that of control cells.

Western blot analysis

Nuclear extracts and lysates were prepared as previously described (Wadman *et al.*, 1997; Park *et al.*, 2009). Western blotting was performed according to standard protocols using antibodies against HNF1, HNF3α/β, HNF4α, C/EBPα, C/EBPβ, histone H1 (Santa Cruz Biotechnology, CA), phospho-p38 MAP kinase, total-p38 MAP kinase, phospho-ERK1/2, total-ERK1/2 (Cell Signaling Technology, Beverly, MA), transthyretin (Dako, Carpinteria, CA), and GAPDH (AbFrontier, Seoul, Korea) (Ju *et al.*, 2009).

Gas chromatography-mass spectrometric analysis (GC/MS) measurements of phenylpyruvic acid

Cells treated with 0.9 mM phenylalanine or 1.2 mM phenylpyruvic acid were sonicated and acidified to pH 2. The acidified solution was supplemented with NaCl and extracted with ethyl acetate. Tropic acid (Sigma-Aldrich)

was added as an internal standard. The organic phase was separated by centrifugation and transferred to a new tube. The dried samples were mixed with N-methyl-N-trimethylsilyltrifluoroacetamide (Pierce Chemical Co., IL). Finally, the samples were incubated for 30 min at 80°C, and 2 µl of the analyte was injected into an HP-5MS column (Agilent J & W Scientific, Folsom, CA) and analyzed using an Agilent 7890A gas chromatograph coupled with an Agilent 5975C inert MSD mass spectrometer (Agilent Technologies, Santa Clara, CA).

Acknowledgments

This study was supported by a Korean Health 21 R&D Project from the Ministry of Health and Welfare, Republic of Korea (A010384).

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