

Obesity and genetic polymorphism of *ERCC2* and *ERCC4* as modifiers of risk of breast cancer

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Abbreviations: BMI, body mass index; CI, confidence interval; NER, nucleotide excision repair; OR, odd ratio

Abstract

To evaluate the relationship of genetic polymorphisms of *ERCC2* and *ERCC4* genes, both involved in nucleotide excision repair (NER), and the risk of breast cancer, a hospital-based case-control study was conducted in Korea. Histologically confirmed breast cancer cases ($n = 574$) and controls ($n = 502$) with no present or previous history of cancer were recruited from three teaching hospitals in Seoul during 1995-2001. Information on selected characteristics was collected by interviewed questionnaire. *ERCC2* Asp³¹²Asn (G > A) was genotyped by single-base extension assay and *ERCC4* Ser⁸³⁵Ser (T > C) by dynamic allele-specific hybridization system. Although no significant association was observed between the genetic polymorphisms and the risk of breast cancer, women with both *ERCC2* A allele- and *ERCC4* C allele-containing genotypes showed a 2.6-fold risk (95% CI: 1.02-6.48) of breast cancer compared to

women concurrently carrying the *ERCC2* GG and *ERCC4* TT genotypes. The breast cancer risk increased as the number of "at risk" genotypes increased with a borderline significance (P for trend = 0.07). Interactive effect was also observed between *ERCC4* genotype and body mass index (BMI) for the breast cancer risk; the *ERCC4* C allele containing genotypes posed a 1.7-fold (95% CI: 0.96-2.93) breast cancer risk in obese women (BMI > 25 kg/m²) with a borderline significance. Our finding suggests that the combined effect of *ERCC2* Asp³¹²Asn and *ERCC4* Ser⁸³⁵Ser genotypes might be associated with breast cancer risk in Korean women.

Keywords: body mass index; breast neoplasms; DNA repair enzymes; *ERCC4* protein

Introduction

Breast cancer is the second common malignancy diagnosed among Korean women and the incidence is increasing continuously (National Statistical Office of Korea, 1998). Although a substantial proportion of breast cancer cases are explained by well-established risk factors (*i.e.*, later age of first birth, nulliparity and first-degree family history of breast cancer) (Madigan *et al.*, 1995), the reason for the observed worldwide increase in breast cancer incidence is still largely unknown.

Because of the importance of maintaining genomic integrity in carcinogenesis (Seo *et al.*, 2004), genes coding for DNA repair molecules have been proposed as candidates for modifiers of individual susceptibility to breast cancer (Shields *et al.*, 1991). This view is supported by previous epidemiological studies suggesting that a combination of genetic polymorphism of DNA repair genes and exposure to genotoxic agents may contribute to breast cancer development (Patel *et al.*, 1997; Goode *et al.*, 2002; Smith *et al.*, 2003).

The nucleotide excision repair (NER) pathway repairs bulky lesions such as pyrimidine dimers, photo-products, large chemical adducts, and cross-links (Wood *et al.*, 1997). The NER pathway involves at least four steps: (a) damage recognition by a complex of bound protein XPC; (b) unwinding of the DNA by the TFIIH complex that includes *ERCC2*; (c) removal of the damaged single-stranded fragment by molecules including an *ERCC1-ERCC4* complex; and (d) synthesis by DNA polymerase (Friedberg *et al.*, 2001).

The *ERCC2* protein possesses both single-strand DNA-dependent ATPase and 5'-3' DNA helicase ac-

tivities and is thought to participate in DNA unwinding during the NER. To date, several genetic polymorphisms of *ERCC2* gene have been found, *i.e.*, in exon 6 (22541: Arg¹⁵⁶Arg, A>C), exon 10 (23591: Asp³¹²Asn, G>A), and exon 23 (35931: Lys⁷⁵¹Gln, A>C) (Winsey *et al.*, 2000). Although both Asp³¹²Asn and Lys⁷⁵¹Gln are nonsynonymous and related to functional activities (Tang *et al.*, 2002), Asp³¹²Asn locus was selected in this study because Asp³¹²Asn polymorphisms has been shown to affect smoking related lung cancer (Hou *et al.*, 2002; Zhou *et al.*, 2002) and breast cancer risk whereas Lys⁷⁵¹Gln is not associated with breast cancer risk and Asp³¹²Asn and Lys⁷⁵¹Gln in the strong linkage disequilibrium ($P < .0001$) (Justenhoven *et al.*, 2004).

The ERCC4 has an important role of removing the damaged single-stranded fragment by constructing an ERCC1-ERCC4 complex. Several genetic polymorphisms of *ERCC4* gene have been found, including those in 5' noncoding region (g2063: T>A), exon 8 (1224: Arg⁴¹⁵Gln, G>A; 1727: Arg⁵⁴⁶Thr, G>C), and exon 11 (2117: Ile⁷⁰⁶Thr, T>C; 2505: Ser⁸³⁵Ser, T>C; 2624: Glu⁸⁷⁵Gly, A>G) (Fan *et al.*, 1999). A few studies on association between *ERCC4* genotypes and cancer proneness was conducted: Ser⁸³⁵Ser and melanoma skin cancer (Winsey *et al.*, 2000) and Arg⁴¹⁵Gln and breast cancer (Smith *et al.*, 2003). Since the genotype of frequency of Gln/Gln genotype of codon 415 was very rare (3%) and there was no significant association with breast cancer in Caucasian women, Ser⁸³⁵Ser locus of exon 11 was selected in this study.

Our previous studies suggest that *XRCC1* Arg³⁹⁹Gln and *hOGG1* Ser³²⁶Cys genotypes may be associated with the risk of breast cancer (Kim *et al.*, 2002; Choi *et al.*, 2003). Here we extended the study to examine the association between the *ERCC2* Asp³¹²Asn (G>A) and *ERCC4* Ser⁸³⁵Ser (T>C) genotypes and breast cancer risk. These genes and loci were selected on the basis of their functional significance and known allele frequencies (Fan *et al.*, 1999; Winsey *et al.*, 2000; Hou *et al.*, 2002; Tang *et al.*, 2002; Zhou *et al.*, 2002).

Subjects and Methods

Study subjects

The study subjects were recruited from patients admitted to the department of surgery at three teaching hospitals located in Seoul (Seoul National University Hospital, Borame Hospital, and Asan Medical Center) from March 1995 to January 2001 (Lee *et al.*, 2004). Eligible subjects consisted of a series of histologically confirmed incident breast cancer patients ($n = 577$) and non-cancer controls ($n = 507$) from whom blood samples were available. After excluding subjects with previous history of cancer, hysterectomy or oophorectomy, the final study population consisted of 574 cases and 502 controls. The study design was approved by the Committee on Human Research of

Seoul National University Hospital. The study subjects gave informed consent prior to participation in the study. Informed consent was obtained from all participants at the time of blood withdrawal.

Information on demographic characteristics, education, marital status, family history of breast cancer, reproductive factors and menstruation, and life style habits (including smoking and alcohol consumption) was collected using a questionnaire administered by trained interviewers.

Genotyping Methods

DNA was isolated using standard methods from whole blood drawn into 10 ml heparinized tubes and stored in -70°C until use.

The *ERCC2* Asp³¹²Asn genotype was determined by single base extension assay. Polymerase chain reaction (PCR) product was obtained using 500 nM of oligonucleotide primers (P1: 5'-CCC AGC TCA TCT CTC CGC AGG ATC A-3' and P2: 5'-GGG AGG CCG GAA AGG GAC TG-5') in a total volume of 20 μl . The amplification conditions were: initial denaturation at 95°C for 5 min followed by 35 cycles of 30 s at 94°C and 180 s at 72°C .

Primer extension was performed by combining 1 μl of exonuclease I and alkaline phosphatase treated PCR product with 5 μl single base extension kit, 0.15 pmol extension primer (5'-TGG CCA ACC CCG TGC TGC CC-3') and 3 μl water. The reaction mixture was incubated at 94°C for 2 min prior to PCR of 25 cycles of 95°C for 5 s, 50°C for 5 s, and 60°C for 5 s. Aliquots of 1 μl SNaPshot product and 9 μl Hi-Di formamide, were combined in a 96-well 3100 optical microamp plate, which was loaded onto a 3100 DNA sequencer (Applied Biosystems, Foster city, CA). Reactions were electrophoresed on a 36-cm capillary array at 60°C by using POP4 polymer, dye set "E" and Genescan run module "SNP36POP4_default." Electrophoresis data were processed by Genescan Analysis version 3.7 (Applied Biosystems, Foster city, CA).

The *ERCC4* Ser⁸³⁵Ser genotype was determined by dynamic allele-specific hybridization system (DASH, Hybaid). The 61 bp PCR product was obtained from about 10 ng DNA using the following oligonucleotide primers P1: 5'-GGC GAC AGC ACT GGC CAT TAC-3' (20 pmol) and P2: biotin-labeled 5'-ATT ATA CTT CTC TGA CTC GG-3' (4 pmol) (Bioneer: Seoul, Korea) in 20 μl volume PCR reaction. The genotype data could be achieved for 67% of the subjects for *ERCC4* and of 93% the subjects for *ERCC2*. The main reasons for the low success rate were too low amount of DNA and/or unsuccessful PCR amplification.

Statistical analysis

Chi-square test and Fisher's exact test were used to test whether observed genotype data were consistent with Hardy-Weinberg equilibrium and to compare the allele frequencies between cases and controls. Odds ratios (ORs) and 95% confidence intervals (CIs) were

calculated by unconditional logistic regression model. The ORs were adjusted for age, body mass index (BMI), lifetime estrogen exposure, education and family history of breast cancer. To test the effect of combined genotype, we also carried out a likelihood ratio test for the multiple genotypes grouped by the *ERCC2* Asp³¹²Asn and *ERCC4* Ser³³⁵Ser polymorphisms. Lifetime estrogen exposure was calculated for premenopausal women by subtracting the menarcheal age and child number and multiplied by 280/365 from the age at interview, and for postmenopausal women subtracting the menarcheal age and child number \times 280/365 from the age at menopause (Huang *et al.*, 1999). To increase the statistical power, the respective genotypes were divided into two groups in the statistical analyses. All analyses were carried out with the SPSS (version 10.0) statistical software package.

Results and Discussion

The mean age was 47.9 in cases and 46.9 in controls. Higher education (at and over high school; OR = 2.0, 95% CI: 1.47-2.67), BMI (≥ 25 kg/m²; OR = 1.5, 95% CI: 1.07-2.02), and family history of breast cancer (OR = 2.4, 95% CI: 1.34-4.40) were significantly associated with breast cancer risk (Table 1). There was a significant trend of increased risk of the lifetime estrogen exposure increased ($P_{\text{for trend}} < 0.001$, Table 1).

The frequency of *ERCC2* A allele in the present Asian controls (5.3%) was drastically lower than that previously reported in Caucasians (33-44%) (Wensey *et al.*, 2000; Tomescu *et al.*, 2001; Vogel *et al.*, 2001; Hou *et al.*, 2002), whereas the frequency of *ERCC4* C allele (23%) was similar to the frequencies found in UK (23%) (Wensey *et al.*, 2000) and USA (33%) (Fan *et al.*, 1999). When the *ERCC2* and *ERCC4* genotypes were studied separately, no significant overall associations with the risk of breast cancer were seen (Table 2). No association between the *ERCC2* Asp³¹²Asn polymorphisms and breast cancer observed in this study is consistent with a previous study (Tang *et al.*, 2002), but not consistent with a recent larger study (Justenhoven *et al.*, 2004). However, interactive effect was observed between *ERCC4* and BMI for the breast cancer risk: the *ERCC4* C allele containing genotypes posed a 1.7-fold (95% CI: 0.96-2.93) breast cancer risk in obese women (BMI > 25 kg/m²) with a borderline significance. There was no association between *ERCC4* genetic polymorphism and BMI (OR = 1.2, 95% CI: 0.89-1.84, data not shown).

Although there is no previous data available on the potential association between *ERCC2* genetic polymorphism and breast cancer risk, the present negative finding is similar to the result reported by Fraglia *et al.* that the *ERCC2* genotypes did not significantly affect the formation of 4-aminobiphenyl-DNA adducts in breast tissue (Fraglia *et al.*, 2003). Although the odds ratio was just statistically borderline significant, the effects of *ERCC4* C allele containing genotypes on the risk of breast cancer in women with higher BMI might be explained by the fact that a higher BMI may be associated with higher levels of lipophilic aromatic compounds, stored in breast adipose tissue, leading to a continuous exposure to DNA-damaging agents (Gorlewska-Roberts *et al.*, 2002). Deficient

Table 1. Selected characteristics of study subjects.

	Cases (n = 574)	Controls (n = 502)	OR* (95% CI)
Mean age (yr)	47.9 \pm 10.55	46.9 \pm 13.93	$P = 0.180$
BMI			
< 25 kg/m ²	437 (77.1)	408 (81.4)	1.0
≥ 25 kg/m ²	130 (22.9)	93 (18.6)	1.5 (1.07-2.02)
Family history of breast cancer			
No	528 (92.0)	485 (96.6)	1.0
Yes	46 (8.0)	17 (3.4)	2.4 (1.34-4.40)
Education			
Less than high-school	188 (32.9)	211 (42.4)	1.0
At and over high-school	384 (67.1)	287 (57.6)	2.0 (1.47-2.67)
Lifetime estrogen exposure			
< 20 yr	69 (12.1)	136 (27.5)	1.0
20-30 yr	306 (53.5)	215 (43.5)	3.9 (2.67-5.84)
> 30 yr	197 (34.4)	143 (28.9)	4.2 (2.59-6.69)
			$P_{\text{for trend}} < 0.001$

*Adjusted for age, BMI (< 25 kg/m² vs. ≥ 25 kg/m²), lifetime estrogen exposure (< 20 yr, 20-30 yr, ≥ 30 yr), education (less than high school vs. at and over high school), and family history of breast cancer.

Table 2. ERCC2 Asp³¹²Asn, and ERCC4 Ser⁸³⁵Ser and risk of breast cancer development.

	All women			Premenopausal women			Postmenopausal women		
	Cases N (%)	Controls N (%)	OR* (95% CI)	Cases N (%)	Controls N (%)	OR* (95% CI)	Cases N (%)	Controls N (%)	OR* (95% CI)
ERCC2									
GG	475 (90.0)	401 (90.1)	1.0	300 (89.0)	238 (89.1)	1.0	175 (91.6)	160 (91.4)	1.0
GA	50 (9.5)	41 (9.2)	1.2 (0.74-1.85)	35 (10.4)	26 (9.8)	1.3 (0.72-2.30)	15 (7.9)	15 (8.6)	0.9 (0.43-2.03)
AA	3 (0.5)	3 (0.7)	0.8 (0.12-3.23)	2 (0.6)	3 (1.1)	0.4 (0.06-2.26)	1 (0.5)	-	-
GA+AA	53 (10.0)	44 (9.9)	1.1 (0.72-1.75)	37 (11.0)	28 (10.9)	1.1 (0.66-2.01)	16 (8.4)	15 (8.6)	1.0 (0.47-2.16)
Total	528 (100)	445 (100)		337 (100)	267 (100)		191 (100)	175 (100)	
ERCC4									
TT	211 (54.7)	194 (57.7)	1.0	141 (56.2)	110 (57.6)	1.0	70 (51.7)	82 (57.3)	1.0
TC	157 (40.7)	129 (38.4)	1.1 (0.85-1.63)	98 (39.0)	73 (38.2)	1.1 (0.77-1.81)	59 (43.7)	56 (39.2)	1.1 (0.68-1.92)
CC	18 (4.6)	13 (3.9)	1.3 (0.57-2.74)	12 (4.8)	8 (4.2)	1.1 (0.43-2.99)	6 (4.4)	5 (3.5)	1.4 (0.36-5.26)
TC+CC	175 (45.3)	142 (42.3)	1.2 (0.87-1.62)	110 (43.8)	81 (42.4)	1.1 (0.78-1.77)	65 (48.1)	61 (42.7)	1.2 (0.70-1.92)
Total	386 (100)	336 (100)		251 (100)	191 (100)		135 (100)	143 (100)	

*Adjusted for age, BMI (< 25 kg/m² vs. ≥ 25 kg/m²), lifetime estrogen exposure (< 20 yr, 20-30 yr, ≥ 30 yr), education (less than high school vs. at and over high school), and family history of breast cancer.

Table 3. Association between ERCC4 genotype and BMI for the risk of breast cancer.

BMI (ka/m ²)	ERCC4	Cases N (%)	Controls N (%)	OR* (95% CI)
< 25 kg/m ²	TT	169 (44.2)	159 (47.3)	1.0
	CT/CC	128 (33.5)	116 (34.5)	1.1 (0.77-1.52)
≥ 25 ka/m ²	TT	40 (10.5)	35 (10.4)	1.2 (0.72-2.10)
	CT/CC	45 (11.8)	26 (7.7)	1.7 (0.96-2.93)

*Adjusted for age, lifetime estrogen exposure (< 20 yr, 20-30 yr, ≥ 30 yr), education (less than high school vs. at and over high school), and family history of breast cancer.

DNA repair may contribute to the accumulation of damaged DNA unrepaired in both lymphocyte and target tissue.

When the combined genotype effects were examined, women with concurrent presence of ERCC2 A allele and ERCC4 C allele containing genotypes showed a 2.6-fold (95% CI: 1.02-6.48) risk of breast cancer compared to those with ERCC2 GG and ERCC4 TT genotype combination (Table 4). The risk of breast cancer increased with the number of "at risk" genotypes, however it was only borderline significance ($P_{\text{for trend}} = 0.07$). This gene-gene interaction may be explained by that ERCC2 opens up the DNA structure and ERCC4 forms a complex with ERCC1 that incises DNA at the 5' side of a bulky adduct lesion (Friedberg *et al.*, 2001). Genetic variants of ERCC2 and ERCC4 may have a combined effect on breast cancer risk since both ERCC2 and ERCC4 are involved in nucleotide excision repair. In conclusion,

Table 4. Combined effect of ERCC2 and ERCC4 for the risk of breast cancer.

ERCC2 and ERCC4	Cases N (%)	Controls N (%)	OR* (95% CI)
All women			
GG and TT	171 (50.3)	148 (53.0)	1.0
GA/AA or TC/CC	150 (44.1)	123 (44.1)	1.1 (0.75-1.48)
GA/AA and TC/CC	19 (5.6)	8 (2.9)	2.6 (1.02-6.48)
			$P_{\text{for trend}} = 0.07$
Premenopausal			
GG and TT	118 (52.2)	89 (52.4)	1.0
GA/AA or TC/CC	94 (41.6)	76 (44.7)	1.0 (0.62-1.49)
GA/AA and TC/CC	14 (6.2)	5 (2.9)	2.8 (0.83-9.11)
			$P_{\text{for trend}} = 0.08$
Postmenopausal			
GG and TT	53 (46.5)	57 (53.3)	1.0
GA/AA or TC/CC	56 (49.1)	47 (43.9)	1.1 (0.66-2.06)
GA/AA and TC/CC	5 (4.4)	3 (2.8)	2.3 (0.19-10.4)
			$P_{\text{for trend}} = 0.53$

*Adjusted for age, BMI (< 25 kg/m² vs. ≥ 25 kg/m²), lifetime estrogen exposure (< 20 yr, 20-30 yr, ≥ 30 yr), education (less than high school vs. at and over high school), and family history of breast cancer.

our finding suggests that the combined effect of ERCC2 Asp³¹²Asn and ERCC4 Ser⁸³⁵Ser genotypes may be associated with breast cancer risk. However, a larger study combined with other SNP loci of functional significance (e.g., ERCC2 Lys⁷⁵¹Gln, ERCC4 5' noncoding region, etc.) is warranted in the future.

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