ARTICLE

Gene-ontology enrichment analysis in two independent family-based samples highlights biologically plausible processes for autism spectrum disorders

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Recent genome-wide association studies (GWAS) have implicated a range of genes from discrete biological pathways in the aetiology of autism. However, despite the strong influence of genetic factors, association studies have yet to identify statistically robust, replicated major effect genes or SNPs. We apply the principle of the SNP ratio test methodology described by O'Dushlaine *et al* to over 2100 families from the Autism Genome Project (AGP). Using a two-stage design we examine association enrichment in 5955 unique gene-ontology classifications across four groupings based on two phenotypic and two ancestral classifications. Based on estimates from simulation we identify excess of association enrichment across all analyses. We observe enrichment in association for sets of genes involved in diverse biological processes, including pyruvate metabolism, transcription factor activation, cell-signalling and cell-cycle regulation. Both genes and processes that show enrichment have previously been examined in autistic disorders and offer biological plausibility to these findings.

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INTRODUCTION

Autism is a complex neurodevelopmental disorder characterized by impairments of varying severity in the three core areas of communication, social interaction and repetitive behaviour. Population prevalence of autism is approximately 15–20 per 10 000 with all autism spectrum disorders (ASD) estimated at 60 in 10 000 children.^{1,2} The role of genetic factors in the development of autism is undisputed. Heritability has been estimated as high as 91–93% using a multi-threshold liability model.³ However, despite the strong influence of genetic factors, autism linkage studies and association studies of common SNPs have not identified any genes of major effect. Recent genome-wide association studies (GWAS) have implicated a number of genes from discrete biological pathways in the aetiology of autism.^{4–6} In a recent study by the AGP using these data, we identified genome-wide significant association with *MACROD2*.⁷ However, we did not observe strong marker-wise associations within the cadherin gene region (*CDH9*, *CDH10*) or the *TAS2R1*, *SEMA5A* region that were highlighted in the work of Wang and co-workers,⁴ Ma and co-workers⁵ and Weiss and co-workers.⁶ In addition to identifying genome-wide significant association, it can be hypothesized that additional true vulnerability loci may exist within the nominal to modest range of statistical significance and confer risk to the disorder.⁸ A milieu of nominal to modestly associated risk variation fits with a polygenic model of disease and presents additional challenges for the

identification of patterns of association within expected experimental noise.⁹

One promising approach is to examine association enrichment within 'pathways' or groups of genes. The underlying hypothesis of association enrichment analysis is that functional polymorphisms that exist within a group of biologically inter-related genes are in essence 'disrupting' the normal functioning of the biological process of the pathway. Consequently, one can consider the biological process, rather than the individual gene or SNP, in the development of the disease/disorder. By examining the ratio of association signals within a group of genes, we can determine whether there is enrichment of the signal above that expected by chance. This strategy decreases the multiple-testing burden that accompanies GWAS, and can have increased power to observe association.

A number of pathway-based methodologies have been developed to examine gene enrichment in association data (reviewed by Wang *et al*¹⁰). These include gene ranking algorithms,¹¹ gene-enrichment algorithms, for example, ALIGATOR (Association List Go AnnoTatOR)⁹ and SNP enrichment approaches such as the SNP ratio test (SRT).¹² The SRT provides a formal test of whether markers within pre-defined pathways show enrichment in association signal over that expected by chance alone. For case-control data, the basic algorithm underpinning the SRT is to first calculate the ratio of the number of nominally associated SNP markers within a pathway to the total number of markers within the pathway. Significance is assigned

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through a case-randomisation permutation routine, which takes account of the linkage disequilibrium between markers.

To apply the SRT to family-based data, we are unable to perform standard case randomisation; therefore, a pseudo-sibling model is generated from the alleles that are not transmitted to the proband. A proband-randomisation procedure is performed within the family, whereby the affection status of the offspring (case and pseudo-sibling) is permuted. This method allows retention of the linkage disequilibrium structure within the families and retains the advantages of the transmission disequilibrium test (TDT) design for the family-based association. In this study, we chose the SRT over other approaches for a number of reasons. Firstly, as the SRT retains all of the markers from the association analysis, it is sensitive to more than one true association signal per gene, and therefore gains information in the presence of allelic heterogeneity. Secondly, the SRT's use of multiple association signals across a gene as opposed to a single maximum signal limits potential genotyping artefact effects. Genotyping error at a single point may highlight a gene erroneously in a maximum signal design where this becomes the only observation. However, taking the ratio of all signals across a gene restricts the impact of single points of error as they are more likely to be diluted across the gene. Thirdly, the SRT also controls for gene size and linkage disequilibrium effects by permuting case-ness independently of genotype, consequently maintaining the same recombination patterns. Approaches that do not apply a gene-wise correction to GWAS data can show inflated signals for pathways containing larger genes. This is often the case in brain expressed pathways that are enriched for larger genes such as cell-surface receptors and can lead to misinterpretation of any association enrichment. Finally, as the SRT uses an SNP-wise association statistic over a gene-wise association statistic, we have sufficient observations to examine pathways that may contain fewer genes. Thereby, we are able to examine discrete 'niche' pathways as well as larger, more diverse gene sets for enrichment in the GWAS.

For this study, we use gene-set lists derived from the gene-ontology (GO) (<http://www.geneontology.org>) database to examine whether association enrichment is present in a cohort of individuals from the Autism Genome Project (AGP) with a diagnosis of autistic disorder.

SUBJECTS AND METHODS

Subjects

The individuals examined in this study were collected as part of the AGP Consortium genome analysis project. The AGP represents more than 50 centres in North America and Europe. Subjects with known karyotypic abnormalities, fragile X mutations or other known genetic disorders were excluded. Diagnostic and ancestral definitions were as previously reported by this group.⁷ Briefly, families are grouped into two nested diagnostic classes (Strict and Spectrum) based on proband diagnostic measures. To qualify for the Strict class, affected individuals met the criteria for autism on both primary diagnostic instruments: the Autism Diagnostic Interview-Revised (ADI-R¹³) and Autism Diagnostic Observation Schedule (ADOS¹⁴). ADI-R-based diagnostic classification of subjects as ASD followed the criteria published by Risi and co-workers.¹⁵ Specifically, individuals who almost met the ADI criteria for autism were classified as ASD if: (1) they met the criteria on social and either communication or repetitive behaviour domains; or (2) met the criteria on the social domain and were within two points of criteria for communication, or met the criteria on the communication domain and were within two points of social criteria, or within one point on both social and communication domains. The Spectrum class included all individuals who met the Strict criteria and those individuals who were classified as ASD or autism on both the ADI-R and ADOS or who were not evaluated on one of the instruments, but were diagnosed with autism on the other instrument. A summary of the sample sizes for the discovery and replication datasets for each Diagnostic|Ancestry subset is shown in Table 1.

Table 1 Sample size for the discovery and replication samples for each diagnostic/ancestral subset

Diagnosis Ancestry	Discovery (n)	Replication (n)	Combined (n)
Total	1248	874	2122
Spectrum/All	1248	874	2122
Spectrum/European	1169	704	1873
Strict/All	775	591	1366
Strict/European	726	477	1203

Each sample is made up of parent-proband trios with at least a Spectrum diagnosis of ASD (Spectrum/All).

Individuals were selected from the total if they were determined to be of European ancestry (Spectrum/European) or if they show a more restrictive diagnosis of Autism (Strict/All) or if they met both restrictive ancestral and diagnostic thresholds (Strict/European).

As described elsewhere,⁷ ancestry for these individuals was determined for the proband by using 5239 widely spaced, independent SNPs that had a genotype completion rate of $\geq 99.9\%$. The software used was Spectral-GEM,¹⁶ which estimated five significant dimensions of ancestry. Subsequent clustering on dimensions of ancestry identified nine clusters; five clusters were used to describe European ancestry and the remaining clusters best reflect Asian, African (East/West) and Latin American origins. The *All ancestry* class included all individuals including those who met the *European ancestry* criteria.

Genotyping and association analysis (TDT)

The discovery sample were genotyped using the Illumina Infinium 1M-single SNP microarray, the replication sample were genotyped on either the Illumina Infinium 1M-single SNP microarray or the Illumina 1M-duo microarray. All quality control (QC) procedures were maintained across datasets; in addition, QC marker sets from both the discovery and replication datasets were matched and only those markers meeting QC for both the discovery and replication datasets were carried forward to analysis. Additional QC details are described elsewhere.⁷ A total of 856 932 SNPs passed QC on both the discovery and replication sample. TDT statistics were generated using PLINK v.1.07.¹⁷

Pedigree SRT

The pedigree SRT (pedSRT) is a modification to the SRT described by O'Dushlaine and co-workers,¹² which is applicable to family-based data. Briefly, the SRT tests the ratio of the number of associated SNPs to the total number of SNPs in a pre-defined set of genes. A marker is considered 'associated' if the association statistic is observed below a given threshold. The threshold used is arbitrary, but is set by default at an unadjusted $P \leq 0.05$. The significance of the ratio is determined through permutation using an empirical P -value derived from the proportion of the ratios for the permuted datasets that are greater than or equal to the observed ratio.¹² We performed 10 000 permuted GWAS analyses for each of the diagnostic, ancestry strata for both the discovery and replication datasets. The pedSRT determines association using the TDT,¹⁸ as implemented in PLINK.¹⁷ In a case-control model, permutation is performed using case randomisation. In the TDT design, case randomisation is performed by creating a pseudo-sibling. The pseudo-sibling is created from the non-transmitted alleles from the parents. Within each permutation cycle either the proband or pseudo-sibling is considered the 'case'. Alternate case randomisation for the TDT is implemented in PLINK, using the alternate phenotype routine.

It is important to note that to reduce type-I error in the SRT due to inflation of the original association signal, for each permutation 'associated' SNPs are assigned according to their rank in the dataset.¹² In short, the numbers of SNPs (T) that meet the 'associated' threshold are calculated from the primary dataset. For each permuted dataset, the top T SNPs are termed 'associated'.

All SNP ratio statistics were calculated using custom scripts in STATA version 10 (Stata Corp., College Station, TX, USA).

Gene tagging

Individual SNP codes from the Illumina 1M Infinium SNP array platform were updated to reflect build 130 of dbSNP. SNPs were assigned to genes using gene

criteria from the dbSNP/NCBI criteria; namely, if the SNP resides within the locus containing the gene transcript including 2 kb 5' and 500 bp 3' of the transcript. The gene assignment protocol was performed using the NCBI criteria and facilitated using the file b130_SNPContigLocusId_36_3.bcp available at ftp://ftp.ncbi.nih.gov/snp/organisms/human_9606/database/organism_data/.

Gene-set selection

Gene sets were described using the GO database (<http://www.geneontology.org>).¹⁹ Gene lists were obtained from the OBO format 1.2 database release available from <http://www.geneontology.org/GO.downloads.ontology.shtml> (build release date 15 December 2009). GO terms are structured in a semihierarchical relationship within the cellular component, molecular function and biological process nodes. Daughter ontology terms are more specialized and parent ontology terms are less specialized. However, unlike a hierarchy, a term may have more than one parent term.

Parent terms were populated by their daughter terms to describe a composite list of genes for each term. SNP ratios were calculated on GO terms with greater than 20 SNPs, but less than 2000 SNPs, and greater than 1 gene, but no more than 1000 genes. A total of 6853 GO terms met these criteria. To account for identity of terms, we merged those GO terms containing identical gene lists; in total the list of unique terms is 5955.

Simulation of GO terms

As mentioned above, the GO terms used in this study can show considerable overlap owing to term redundancy, biological overlap and the hierarchical nature of the database. Simulations were performed to calculate the null distribution and subsequent expectancy for the total number of associated GO terms at a given threshold in a single study given the GO terms used.

We performed 1000 pedSRT permutations on a case-randomized sample derived from 1248 families from the discovery dataset. A GWAS TDT was performed on each dataset followed by pedSRT, using 10 000 additional permutations on the 5995 GO terms. For each of the 1000 original permutations, the proportion of the 5955 GO terms that met a significance threshold of $P \leq 0.05$ in the subsequent 10 000 was calculated. The mean proportion across the 1000 permutations was used to predict the expected number of associated GO terms in a dataset.

Pathway enrichment map generation

Visual representation of overlap in enriched GO terms was performed using the EnrichmentMap (<http://baderlab.org/Software/EnrichmentMap>)²⁰ plugin for Cytoscape 2.8.0 (<http://www.cytoscape.org/>)²¹. Consistent with the author's recommendations for use with the GO database, nodes were joined if the overlap coefficient was ≥ 0.5 .

RESULTS

Across all analysis in the discovery dataset, 1035 unique GO terms show association enrichment at $\text{SRT-}P\text{-value} \leq 0.05$. Examination of those GO terms that show strong enrichment ($\text{SRT-}P\text{-value} < 0.001$) highlights diverse processes such as the regulation of cell division (mitosis and meiosis), ribosome processing and apoptosis. A visual representation of enriched pathways is shown in Supplementary Figure 1. A summary of the total number of GO terms that show enrichment at $\text{SRT-}P\text{-value} \leq 0.05$ is given in Table 2. On the basis of simulated data, 4.46% (SD=0.8%) of the 5995 unique but non-independent pathways are expected to be associated at $\text{SRT-}P \leq 0.05$ level. Given this level, we would expect 267 GO terms to be associated per experiment. To provide a greater distinction of potentially important GO terms, we examined the overlap of enriched GO terms in an independent replication dataset. On the basis of 4.46% of GO terms showing enrichment, we would expect to observe replication for 12 of the 5995 pathways. All individual discovery samples show more GO terms associated than would be expected by chance (see Expected 1; Table 2). Moreover, the overlap between the discovery and replication

Table 2 Summary of enriched GO terms and overlap in the discovery and replication sample

	Observed enriched GO		Expected discovery GO terms	Expected replicated replication	Expected replication replication
	terms in discovery sample	terms in discovery sample			
Diagnosis Ancestry		(1)		(2)	(3)
Spectrum All	389	267	38	12	17
Spectrum European	392	267	34	12	17
Strict All	329	267	22	12	15
Strict European	354	267	24	12	16

(1) Expected associated GO terms given 4.46% of 5995 pathways showing enrichment; (2) expected replication given 4.46% of 5995 unique pathways showing enrichment in simulated dataset; (3) expected replication given proportion of associated pathways observed in the Discovery and an expected Replication-set enrichment of 4.46%.

sample also show enrichment over what would be expected by chance (see Expected replication 2; Table 2). When we use a more cautious interpretation based on the total number of observed associated GO terms in the discovery data and the predicted replication of 4.46% we would expect to replicate is between 15 and 17 pathways (see Expected 3; Table 2). Under this model, we still show enriched replication for each 'Diagnosis|Ancestry' groupings. Overall, compared to simulated data we observe between 1.5- and 3.2-fold enrichment in the overlap of pathways in the discovery and replication dataset above what can be expected by chance.

A summary of the replicated pathways, summary statistics, gene number and genes tagged in this analysis is shown in Tables 4–7 (full lists of replicated pathways can be found in Supplementary Tables 1A–C). A total of 88 unique GO terms were shown to be replicated within analytic groupings (see Supplementary Table 2), 22 GO terms were replicated within two of the analytic groupings and four GO terms were replicated within three of the analytic groupings (see Table 3). Replication was only considered within strata, such that, for example, GO terms identified in the discovery Strict|European analyses were examined in the Strict|European replication dataset. The four GO terms that show enrichment across three groupings are as follows: GO:0006090, GO:0032872, GO:0032874 and GO:0042156, involved in pyruvate metabolism, regulation of the mitogen-activated protein kinase (MAPK) cascade and zinc-mediated transcriptional activation. A visual representation of replicated enriched pathways is shown in Supplementary Figure 2.

DISCUSSION

The interpretation of GWAS data purely on the strength of association data is challenging where the distribution of association is close to or barely exceeding what is expected by the number of tests. In the absence of clear association enrichment across the entire dataset, interpretation has relied on rank order or via the application of suboptimal significance thresholds that juggle type-I and type-II error. The principle of association enrichment approaches is to discover whether within this milieu of data there are underlying patterns to the association. In these approaches, we ask whether SNPs that are linked to genes of common function show greater proportion of nominal association than expected by chance. Although a modest association signal at an individual SNP within a gene may not warrant further investigation, the cumulative association of SNPs within a gene family may offer insight into the biology of the disorder.

Gene enrichment approaches have been primarily developed to aid interpretation of data from microarray expression studies. In this

Table 3 GO terms showing replicated enrichment in two or more analytic groupings

GO term	GO name	Tagged genes	Groupings
GO:0042156	Zinc-mediated transcriptional activator activity	3	All and <i>Spectrum</i> /European
GO:0006090	Pyruvate metabolic process	39	European and Strict/All
GO:0032872	Regulation of stress-activated MAPK cascade	10	European and Strict/All
GO:0032874	Positive regulation of stress-activated MAPK cascade	9	European and Strict/All
GO:0004758	Serine C-palmitoyltransferase activity	3	European
GO:0017059	Serine C-palmitoyltransferase complex	5	European
GO:0032494	Response to peptidoglycan	6	European
GO:0043330	Response to exogenous dsRNA	8	European
GO:0045070	Positive regulation of viral genome replication	4	European
GO:0006730	One-carbon metabolic process	98	Spectrum
GO:0008276	Protein methyltransferase activity	47	Spectrum
GO:0010712	Regulation of collagen metabolic process	11	Spectrum
GO:0016278	Lysine <i>N</i> -methyltransferase activity	31	Spectrum
GO:0031489	Myosin V binding	2	Spectrum
GO:0032059	Bleb	3	Spectrum
GO:0032400	Melanosome localization	4	Spectrum
GO:0032402	Melanosome transport	9	Spectrum
GO:0032755	Positive regulation of interleukin-6 production	18	Spectrum
GO:0042054	Histone methyltransferase activity	38	Spectrum
GO:0044246	Regulation of multicellular organismal metabolic process	14	Spectrum
GO:0051648	Vesicle localization	5	Spectrum
GO:0060346	Bone trabecula formation	2	Spectrum
GO:0070382	Exocytic vesicle	4	Spectrum
GO:0005876	Spindle microtubule	27	Strict
GO:0015232	Haeme transporter activity	3	Strict
GO:0046888	Negative regulation of hormone secretion	25	Strict

Tagged gene lists for each of the GO term are summarized in Supplementary Table 3.

Table 4 Top 10 association enrichments of pedSRT for overlapping GO terms for analyses of families of all ancestries with a proband with a Spectrum diagnosis

GO term	GO category	GO name	Identity	Total SNPs	Discovery		Replication		Total genes	Tagged genes
					Ratio	P	Ratio	P		
GO:0006730	Biological process	One-carbon metabolic process		1701	0.1053	0.0001	0.0665	0.0437	107	98
GO:0042156	Molecular function	Zinc-mediated transcriptional activator activity		37	0.6818	0.0001	0.2333	0.0201	3	3
GO:0032059	Cellular component	Bleb		64	0.3333	0.0004	0.1636	0.0284	3	3
GO:0000318	Molecular function	Protein–methionine–R-oxide reductase activity		44	0.4194	0.0048	0.2571	0.0354	2	2
GO:0050705	Biological process	Regulation of interleukin-1 alpha secretion	GO:0050717	56	0.2444	0.0085	0.1915	0.0225	3	3
GO:0032650	Biological process	Regulation of interleukin-1 alpha production	GO:0032730	58	0.2340	0.0091	0.1837	0.0238	4	4
GO:0007567	Biological process	Parturition		124	0.1376	0.0152	0.1273	0.0244	7	7
GO:0031489	Molecular function	Myosin V binding		56	0.2174	0.0167	0.3023	0.0020	2	2
GO:0032402	Biological process	Melanosome transport	GO:0051904	118	0.1456	0.0189	0.1683	0.0083	9	9
GO:0046628	Biological process	Positive regulation of insulin receptor signalling pathway		43	0.1944	0.0194	0.1944	0.0180	3	3

Ratio refers to the ratio of associated to non-associated SNPs per GO term. *P* refers to the empirical significance generated from 10 000 permutations.

context, each gene is tagged by either one or a small number of probes regardless of gene size. However, when applying these technologies to SNP-based data, we do not measure gene-wise variation or gene-wise association; instead, we can potentially examine multiple points of association at any given gene using many tagging SNPs. This brings additional challenges and bias. When applying association

enrichment, we must account for and correct for these potential bias in these data. Firstly, when examining larger genes, we utilize more SNP markers to tag the variation than for smaller genes. If we choose a maximum association signal approach per gene, we observe by chance an inflated signal for the larger genes. By calculating the ratio of associated to not associated SNPs, we can adjust each GO term

Table 5 Top 10 association enrichments of pedSRT for overlapping GO terms for analyses of families of European ancestries with a proband with a Spectrum diagnosis

GO term	GO category	GO name	Identity	Discovery		Replication		Total genes	Tagged genes
				Total SNPs	Ratio	P	Ratio	P	
GO:0042156	Molecular function	Zinc-mediated transcriptional activator activity	37	0.6087	0.0001	0.2333	0.0195	3	3
GO:0006730	Biological process	One-carbon metabolic process	1701	0.0890	0.0017	0.0698	0.0297	107	98
GO:0009566	Biological process	Fertilization	609	0.1362	0.0044	0.0973	0.0398	44	39
GO:0032872	Biological process	Regulation of stress-activated MAPK cascade	122	0.1619	0.0059	0.1091	0.0476	10	10
GO:0006090	Biological process	Pyruvate metabolic process	589	0.1113	0.0065	0.1219	0.0013	40	39
GO:0042788	Cellular component	Polysomal ribosome	22	0.3750	0.0070	0.5714	0.0006	3	3
GO:0031235	Cellular component	Intrinsic to internal side of plasma membrane	147	0.2049	0.0074	0.1667	0.0241	3	3
GO:0032874	Biological process	Positive regulation of stress-activated MAPK cascade	116	0.1600	0.0074	0.1154	0.0412	9	9
GO:0032494	Biological process	Response to peptidoglycan	78	0.2000	0.0080	0.2000	0.0079	6	6
GO:0045070	Biological process	Positive regulation of viral genome replication	50	0.2500	0.0090	0.1905	0.0252	4	4

Ratio refers to the ratio of associated to non-associated SNPs per GO term. *P* refers to the empirical significance generated from 10 000 permutations.

Table 6 Top 10 association enrichments of pedSRT for overlapping GO terms for analyses of families of all ancestries with a proband with a Strict diagnosis

				Discovery			Replication			
GO term	GO category	GO name	Identity	Total SNPs	Ratio	P	Ratio	P	Total genes	Tagged genes
GO:0031146	Biological process	SCF-dependent proteasomal ubiquitin-dependent protein catabolic process		21	0.9091	0.0001	0.5000	0.0009	2	2
GO:0031571	Biological process	G1/S DNA damage checkpoint		43	0.3030	0.0007	0.1944	0.0219	4	4
GO:0015232	Molecular function	Haeme transporter activity		29	0.2083	0.0011	0.1600	0.0070	3	3
GO:0030276	Molecular function	Clathrin binding		489	0.1425	0.0016	0.1063	0.0231	8	7
GO:0004738	Molecular function	Pyruvate dehydrogenase activity	GO:0004739	22	0.1579	0.0088	0.1000	0.0393	3	3
GO:0016139	Biological process	Glycoside catabolic process		35	0.2963	0.0091	0.2069	0.0282	3	3
GO:0000244	Biological process	Assembly of spliceosomal tri-snRNP		34	0.2593	0.0193	0.2593	0.0184	4	4
GO:0032642	Biological process	Regulation of chemokine production		139	0.1301	0.0301	0.1301	0.0306	15	14
GO:0033630	Biological process	Positive regulation of cell adhesion mediated by integrin		67	0.1964	0.0327	0.2182	0.0253	3	2
GO:0032872	Biological process	Regulation of stress-activated MAPK cascade		122	0.1193	0.0333	0.1509	0.0065	10	10

Ratio refers to the ratio of associated to non-associated SNPs per GO term. *P* refers to the empirical significance generated from 10 000 permutations.

to the total number of SNPs examined per GO term. Secondly, where multiple markers tag a gene, one might observe multiple strong association signals due to strong linkage disequilibrium between the associated markers. To reduce this effect, we calculate significance of the data through permutation. Permutation is performed by case randomisation within families where a pseudo-control sibling is created from the alleles that are not transmitted to the proband. By using the non-transmitted alleles, we retain the linkage disequilibrium structure across the genome, thereby retaining linkage-disequilibrium-related inflation in the original association signal.

We have applied the SRT to family-based data from the AGP to identify 88 gene sets from the GO database that show a replicated enrichment for association signal. Of the overlapping GO terms, we observe enrichment in sets involved in diverse biological processes, including pyruvate metabolism, transcription factor activation, cell-signalling and cell-cycle regulation.

One of the strongest findings from the discovery and replication findings was observed across the 'Strict diagnosis|All ancestries' grouping for the GO term GO:0031146; SCF-dependent proteasomal ubiquitin-dependent protein catabolic process (Discovery SRT-*P*=0.0001; Replication SRT-*P*=0.0009). GO:0031146 is described by

only two genes (*FBXO31* and *FBXO6*). Both genes are members of the F-box protein family, which are involved in a variety of molecular and cellular functions, including protein degradation, synapse formation and circadian rhythm.²² *FBXO6* has also been suggested as a putative biomarker for autism,²³ as one of 13 genes highlighted in the work of Nishimura and Brown²⁴ who show differential expression at this gene in the lymphoblastoid cell lines from individuals with both the *FMR1* mutation and autism compared with typically developing controls.

Those GO terms that show replication across multiple diagnostic and ancestral groups are also worth noting as they are robust to differences in sampling used in our analyses. Four replicated GO terms were observed in three analytic groupings (see Table 3). These include GO:0006090, GO:0032872, GO:0032874 and GO:0042156. GO:0006090 (pyruvate metabolic process) describes a group of 39-tagged genes (see Supplementary Table 3) covered by 589 SNPs. These genes are involved in the biological processes connecting the chemical reactions and pathways involving pyruvate. Pyruvate metabolism is a component of the energy metabolism pathway, which has received considerable attention with respect to autism. The biological plausibility of the pyruvate metabolic process association enrichment is supported by numerous studies showing evidence of aberration in

Table 7 Top 10 association enrichments of pedSRT for overlapping GO terms for analyses of families of European ancestries with a proband with a Strict diagnosis

GO term	GO category	GO name	Identity	Discovery			Replication		Total genes	Tagged genes
				Total SNPs	Ratio	P	Ratio	P		
GO:0034142	Biological process	Toll-like receptor 4 signalling pathway	GO:0070427	22	0.8333	0.0001	0.5714	0.0012	2	2
GO:0033083	Biological process	Regulation of immature T-cell proliferation		31	0.4762	0.0002	0.2917	0.0161	3	3
GO:0040036	Biological process	Regulation of fibroblast growth factor receptor signalling pathway		133	0.3300	0.0004	0.1982	0.0235	7	7
GO:0070423	Biological process	Nucleotide-binding oligomerization domain containing signalling pathway	GO:0070431	41	0.3667	0.0011	0.2424	0.0272	4	4
GO:0004758	Molecular function	Serine C-palmitoyltransferase activity	GO:0016454	145	0.1885	0.0032	0.1328	0.0308	3	3
GO:0070555	Biological process	Response to interleukin-1		608	0.1115	0.0035	0.0975	0.0112	17	17
GO:0045879	Biological process	Negative regulation of smoothened signalling pathway		111	0.3059	0.0051	0.2472	0.0197	5	5
GO:0017059	Cellular component	Serine C-palmitoyltransferase complex	GO:0031211	164	0.1631	0.0060	0.1310	0.0256	5	5
GO:0070391	Biological process	Response to lipoteichoic acid		41	0.2813	0.0074	0.1714	0.0437	4	4
GO:0010830	Biological process	Regulation of myotube differentiation	GO:0010832	152	0.1783	0.0093	0.1343	0.0404	3	3

Ratio refers to the ratio of associated to non-associated SNPs per GO term. *P* refers to the empirical significance generated from 10 000 permutations.

pyruvate levels in individuals with autism.²⁵ The GO term GO:0042156 (zinc-mediated transcriptional activator activity) describes a group of three genes tagged by 37 SNPs (*MTF1*, *RNF4* and *ZNF384*). One of the constituent genes, *MTF1*, human metal-regulatory transcription-factor-1, has previously warranted investigation as putative candidate gene for autistic disorder under an environmental exposure model of autism.²⁶ Finally, GO:0032872 (regulation of stress-activated MAPK cascade) and GO:0032874 (positive regulation of stress-activated MAPK cascade), which differ by a single gene (see Supplementary Table 3), describe 10 and 9 genes, and 122 and 116 SNPs, respectively. These pathways are involved in increasing the signalling of the stress-related MAPK signalling pathway. Stress-activated MAPKs are thought to have a critical role in modulating inflammation, DNA damage response, apoptosis in cancer²⁷ and negative regulation of cell-cycle progression.^{28,29} Cell-cycle progression and DNA damage response are also highlighted in enriched replicated GO terms in these analyses, for example, GO:0032404 (mismatch repair complex binding) and GO:0031571 (G1/S DNA damage checkpoint).

In a recent study by this group, we explored enrichment in GO terms for rare deleted CNVs.³⁰ Using individuals from the Discovery Group, we identified 24 enriched GO terms that show enrichment in rare CNV at FDR $q < 0.05$ that highlighted five biological domains: namely cell proliferation, cell projection and motility, MHC-I, GTPase/RAS signalling and kinase activation/regulation. We do not observe any overlap between the 88 gene sets showing replicated enrichment in the GWAS data with the 24 significant GO terms identified for rare structural variation. However, we do observe some overlap for GO terms enriched only in the discovery dataset. These include overlap in 'cell migration', 'cell motility', 'cell morphogenesis' and GO terms identified as having a role in protein kinase regulation.

We can take some encouragement that highlighted pathways are supported in the autism literature. We have emphasized biological plausibility of some of these pathways with autism and ASD. However, one major caveat when interpreting these data is whether this overlapping evidence reflects the considerable literature surrounding autism research and is therefore coincident, or is biologically meaningful concordance.

Pathway approaches, such as the SRT and pedSRT, can also be applied to research questions using candidate gene list. Candidate

genes rely on the selection of genes and markers based on previous knowledge of biology, function and position of the gene or marker. The pathway approach in the form used in this manuscript applies a 'hypothesis-free' design, in which we examine all GO terms regardless of putative role. In a recent autism GWAS described by Wang *et al*,⁴ the authors applied a hypothesis-testing candidate gene approach using their own methodology¹¹ to examine whether a group of cadherin and neurexin genes showed enrichment in their association data. The authors conclude that there was association enrichment for both a group of cadherin, and cadherin plus neurexin genes ($P=0.02$, 0.004, respectively). We applied our approach to these gene lists in our data (data not shown). Using the pedSRT, which differs in statistical method and gene-to-SNP assignment to that of Wang and co-workers, we do not observe significant association signal enrichment in either the discovery or replication dataset for any of the analytic groupings.

To further explore potential overlap of our data and other GWAS, we examined whether previously implicated genes from recent autism GWAS were present in the GO terms identified in this study. None of the genes that overlap with the top-associated SNPs from previous GWAS described by Wang and co-workers⁴ (*CDH22*, *CTNNA3*, *DMD*, *FEZF2*, *LOC100132914*, *LRRC1* and *SYT17*) and Weiss and co-workers⁶ (*ACTN2*, *ADA*, *CENPC1*, *CRIM1*, *CTNNA3*, *CUGBP2*, *GAS2*, *IQGAP2*, *JARID2*, *SGCD* and *XG*) appeared in the 88 unique GO terms showing overlap in these analyses. Moreover, we do not observe overlap with those genes highlighted by the authors as residing close to their maximal association peaks, namely *SEMA5A*, *TAS2R1* and *CDH9*, *CDH10*.

The GO database is continuously updated as evidence is gathered on gene biology. The build of the database used in these analyses contains information on 17 703 genes, compared with less than 5000 for databases such as KEGG. However, not all genes are tagged to GO terms. This is exemplified by the *MACROD2* gene, which contained SNPs showing the strongest association signal from our previous GWAS analyses.⁷ Over time more information will be gathered on the biological role and interactions between these genes to further annotate these terms.

In addition to single gene effects such as *MACROD2*, data presented in this analysis may offer some additional insight into biological processes, within which genetic risk for autism may lie. This can include hypothesis-free gene lists such as those in the GO dataset, or

more hypothesis-driven candidate gene lists highlighting previous linkage, association or biology. The application of pedSRT to our GWAS data has highlighted biological processes previously implicated in autism and offers impetus to re-examine these processes based on evidence from genome-wide investigation. Association enrichment analysis provides additional evidence from GWAS data to identify genetic risk variants and genes and prioritize biological processes for further research into areas such as biomarker discovery, gene–gene interaction analyses and identification of putative drug targets.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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AUTHOR CONTRIBUTIONS

EMK and COD developed the principle of the SRT experiments. RJLA designed the study, developed and implemented the pedSRT methodology and wrote the manuscript. EMK, COD, BLY, MG and LG aided in manuscript preparation. RJLA, EMK, COD, BY, EP, JDB and JSS discussed research strategies and data through the ‘pathway-based analysis working group’. Additional intellectual support and guidance was provided through the AGP including BD, ADP, EHC, PS, JTG, CK, KW, HH and EM.

APPENDIX

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