

## The role of the *hedgehog/patched* signaling pathway in epithelial stem cell proliferation: from fly to human

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### ABSTRACT

The *hedgehog-patched* (*hh-ptc*) intercellular signaling pathway has recently been shown to control the proliferation of epithelial stem cells in both *Drosophila* and vertebrates. Mutant and ectopic expression analyses in *Drosophila* suggest that the HH protein diffuses from the signaling cells to promote the proliferation of nearby ovarian somatic stem cells by antagonizing the suppression of its receptor PTC towards the CI transcription factor in the stem cells. Consequently, the transcription of CI-dependent genes leads to stem cell proliferation. This regulatory pathway appears to function also in vertebrates, where defects in *ptc* cause basal cell carcinoma, tumors of epidermal stem cell origin. Basal cell carcinoma can also be induced by ectopic expression of *Sonic hedgehog* (*shh*) or *Glil*, the vertebrate homolog of *ci*. These studies suggest the conservation of the *hh* signaling pathway in controlling epithelial stem cell divisions among different organisms.

**Key words:** *Stem cell, epithelia, hedgehog, patched, Drosophila, vertebrate.*

### INTRODUCTION

In the famous screen of Nüsslein-Volhard and Wieschaus for genes governing

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*Drosophila* embryonic patterning, *hedgehog* (*hh*) and *patched* (*ptc*) were initially shown to be involved in setting up the segmental boundary pattern of the fly embryo[1]. The *hh* gene encodes a secreted ligand that can bind to its receptor complex composed of the transmembrane proteins on the surface of the target cells encoded by *ptc* and *smoothened* (*smo*) genes[2]. This binding releases the suppression of PTC towards a transcription factor encoded by *cubitus interruptus* (*ci*, a homolog of the mammalian *Gli1* gene) through an intracellular signal transduction cascade, thus allowing several important genes including *wingless* (*wg*, a homolog of the mammalian *Wnt* genes), *decapentaplegic* (*dpp*, a member of the TGF $\beta$  family), and *ptc* itself to be transcribed. The activation of these genes then specifies the fate of the target cells.

Studies in the past several years have revealed that the *hh/ptc* signaling pathway is conserved in a number of postembryonic developmental processes in *Drosophila* and embryonic development in vertebrates. For example, during *Drosophila* metamorphosis, *hh* and *ptc* act to control cell fate decisions for patterning the central nervous system[3], legs, wings, (reviewed in[4, 5]) and eyes[6]. In vertebrates, the *hh/ptc* pathway is involved in neurogenesis and limb patterning[7, 8]. More recently, the *hh/ptc* pathway has also been shown to play a crucial role in regulating stem cell division and differentiation in adult *Drosophila* and mammalian epithelia[9-12], even though they appear to act on a different set of downstream genes. The *hh/ptc* signaling cascade in non-stem cells, especially with regard to components downstream of *ptc* inside the target cells has been reviewed in detail by Ruiz i Altaba[13]. This paper reviews the novel function of *hh/ptc* in controlling stem cell divisions, with emphasis on the unique features of the *hh/ptc* pathway in this process.

## ***hh/ptc* pathway controls epithelial cell proliferation and differentiation in *Drosophila* oogenesis**

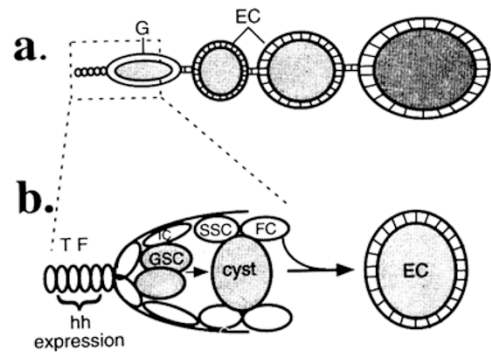
*Drosophila* oogenesis begins at the apical tip of the ovary in 15-17 functionally equivalent structures called the ovarioles (Fig 1a). Within each ovariole, oogenesis starts at the apical tip in a specialized structure called the germarium in which germline stem cells produce germline cysts. Each cyst then becomes enveloped by a layer of somatic epithelial cells called the follicle cells to form an egg chamber, which leaves the germarium and moves towards the oviduct as it develops further. To encapsulate new germline cysts, the somatic stem cells in the middle region of the germarium must undergo active divisions to generate a continuous supply of follicle cells[14] (Fig 1b). Recent work by Forbes et al. [9, 10] revealed that the division and differentiation of the somatic stem cells and their daughter cells are controlled by a group of non-mitotic somatic cells at the very tip of the germarium called the terminal filament cells through the *hh/ptc* signaling pathway.

Forbes et al. [9, 10] first demonstrated a requirement for *hh* in regulating somatic cell proliferation and differentiation during oogenesis by showing that *hh* is specif-

ically expressed in the terminal filament cells. A *hh* enhancer trap line specifically stains the terminal filament; so did anti-HH antibodies. Functional evidence for *hh* requirement comes from examining the effect of removing or ectopically expressing *hh* in the germarium. When females homozygous for a temperature-sensitive *hh* mutation were raised at a permissive temperature to adulthood and then shifted to a restrictive temperature to remove *hh* activity in the ovary, this shift caused the severe underproliferation of follicle cells. In contrast, when *hh* was ectopically expressed as driven by a *hsp70* promoter, this overexpression resulted in an increase in the number of follicle cells. These experiments were the first to establish that the *hh* signaling pathway plays a role in oogenesis and epithelial cell proliferation.

**Fig 1.a.** A schematic drawing of the *Drosophila* ovariole, which is composed of a string of egg chambers (EC) produced by the germarium (G).

**b.** A magnified view of the apical half of the germarium as boxed in panel a. *hh* is expressed in terminal filament cells (TF), while *ptc* is expressed in inner sheath cells (IC), somatic stem cells (SSC), and follicle cells (FC). GSC: germline stem cells; Cyst: germline cyst. The germline is shaded.



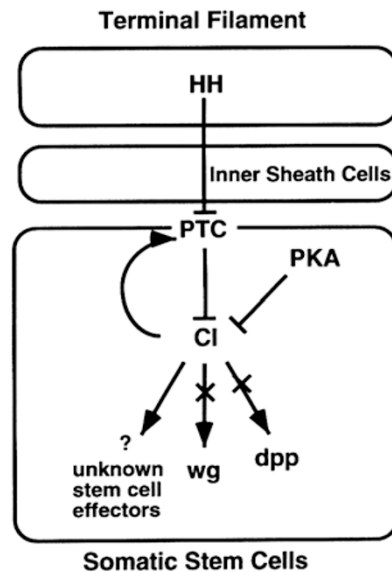
Given that the epithelium is a stem cell-derived tissue, *hh* may control the process by regulating stem cell division. This appears to be true since timed ectopic *hh* expression experiments revealed that *hh* acts on somatic stem cells and their immediate daughters[9].

Since *hh* was known to interact with *ptc* in other *Drosophila* developmental processes, the function of *ptc* and other segment polarity genes in oogenesis were also tested[10]. Both the *ptc-LacZ* reporter and the anti-PTC antibody staining experiments showed that *ptc* indeed is expressed in a pattern complementary to the *hh* expression pattern in the ovary. Loss of *ptc* function in clones of somatic stem cells and/or their daughter cells caused these cells to overproliferate, mimicking the effects of *hh*-overexpression. This suggests that *ptc* acts as a receptor molecule for the *hh* signal in controlling somatic stem cell division during oogenesis (Fig 2). Consistent with this, ectopic expression of *ptc* in the ovary showed no discernible effect on oogenesis, since the PTC protein is already present in the somatic stem cells and their immediate daughter cells without ectopic expression. These experiments provide substantial evidence that the functional relationship between *hh* and *ptc* is

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conserved in the epithelial stem cell system in the *Drosophila* ovary. These results also suggest that downstream targets of the *ptc* signal are insensitive to the concentration of PTC protein as long as it is above the threshold level, yet complete lack of PTC would fail to repress downstream target genes.

**Fig 2.** A simplified diagram of the *hh/ptc* signaling pathway in regulating epithelial stem cell division in the *Drosophila* ovary. This pathway appears to function also in vertebrate epithelia.



Perhaps the most unexpected aspect of the *hh/ptc* pathway in the *Drosophila* epithelial stem cell system is the finding that *dpp* and *wg* are not the downstream effectors of the pathway as is the case in other developmental processes examined so far [10, Fig 2]. Although the *ci* expression suggests that it is under the control of the *hh/ptc* pathway, its target genes no longer include *wg* and *dpp*. It has been well documented in the *Drosophila* embryo and imaginal disc systems that *wg* and *dpp* are the effector genes of the *hh/ptc* signaling pathway, serving as the direct targets of the *CI* transcription factor. The finding that the epithelial stem cells do not require *wg* and *dpp* [10, H.L., unpublished results] suggests that there must be other effector genes more specific or relevant to the stem cell system. Thus, studies by Forbes et al. have shown not only the involvement of the *hh/ptc* pathway in controlling stem cell proliferation but also the divergence of downstream effector genes in this process. This role of the *hh/ptc* pathway in *Drosophila* epithelial stem cell division parallels its role in mammalian epithelial development as revealed by recent findings in vertebrate systems.

## *hh/patched* as regulators of cell proliferation in mammalian epithelia

The *hh/ptc* pathway has been characterized in vertebrate systems by isolating homologs, examining their expression profiles and manipulating their expression experimentally to examine the developmental consequences. For example, ectopic expression of *Sonic hedgehog* (*Shh*), a chick homolog of *hh*, caused mirror-like du-

plications in limb structures that paralleled the mutant phenotype in *Drosophila*[8]. For another example, *ptc* in mice was shown to be expressed in tissues tightly linked to those expressing *Shh*[7]. Mice deficient in *Shh* had severe defects in embryonic development such as neurogenesis, axial determination, skeleton formation and limb development[15]. These results, together with the apparent similarity at the amino acid sequence level, suggest structural and functional conservation of the *hh/ptc* pathway in governing differentiation and patterning during mammalian embryonic development.

Recently, a growing body of evidence shows that the *Shh/ptc* pathway also controls cell proliferation in vertebrate epithelia, a role similar to its function in controlling epithelial stem cell division during *Drosophila* oogenesis. Two recent reports[11, 12] have shown that the human homolog of *ptc*, 96% and 40% amino acid identity to the mouse and *Drosophila* *Ptc* proteins respectively, is mutated in individuals with an autosomal dominant disorder called Nevoid Basal Cell Carcinoma Syndrome (NBCCS). Consistent with the multiple developmental phenotypes of *Drosophila* mutants, defects in human *ptc* include a number of developmental anomalies which define its role in embryonic patterning. In addition, NBCCS individuals are prone to develop basal cell carcinomas (BCCs), as well as the induction of several other classes of tumors, suggesting *ptc* plays an important role in controlling cell proliferation in the affected tissues. The BCCs originate from pluripotent basal skin cells and tumors are associated with loss of heterozygosity at the human *ptc* locus. Since pluripotent basal skin cells by definition should be epidermal stem cells, the overproliferation of these cells in BCCs in *ptc*-deficient patients implies a role for *ptc* as a negative regulator of epidermal stem cell division as shown in *Drosophila*.

Further support for the *hh/ptc* pathway in regulating basal cell division was provided by overexpressing *Shh* in transgenic mice bearing a *Shh* transgene driven by the keratin 14 promoter. This ectopic expression was sufficient to induce BCC-like skin lesions in mice, thereby mimicking the loss of *ptc* function[16].

It is not surprising that several downstream targets of the *hh/ptc* signaling pathway in *Drosophila* are themselves agents in tumorigenesis when mutated in vertebrates. Perhaps the most illustrative case is the role *Gli1*, a vertebrate homolog of *ci*, in incidences of various cancers including BCCs (reviewed in[13]). In all *Drosophila* developmental systems investigated, the transcription factor *ci* has been shown to be the downstream target of *ptc* signaling. In human, *Gli1* was identified as a proto-oncogene causing incidences of glioblastomas (reviewed in[17]). Ectopic expression of *Gli1* in frog embryos has recently been shown to cause the induction of BCCs, suggesting that *Gli1* is potentially under the control of the *hh/ptc* signal in regulating basal stem cell division. This hypothesis is consistent with the observation that in every case of human skin tumors with *Shh* expression, *Gli1* is always expressed[18].

In addition to its role in BCCs, *ptc* has been shown to be mutated in 2 out of 14 sporadic medulloblastomas and in 2 out of 7 incidences of breast carcinomas ex-

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amined[19]. Also, activation of *Wnt-2* and *Wnt-3* has been correlated with tumor formation in the mouse mammary gland as shown by expression profiles and by mutant analysis[20]. Given that breast carcinomas and mammary gland tumors are of epithelial origin, it is likely that *ptc* and *Wnt* genes are involved in controlling epithelial stem cell divisions in this organ. Particularly, the involvement of *Wnt-2* and *Wnt-3* genes in this process suggests that other *wg* homologs in *Drosophila*, instead of *wg* itself, may be involved in regulating ovarian epithelial stem cell division. One future direction may be to examine which of these *Wnt* genes are actual targets of *ptc* regulation in *Drosophila* and mammals.

Although the use of *Drosophila* to dissect the molecular mechanisms of tumorigenesis is hindered by the short life cycle of the fly, screens for recessive mutations that cause overproliferation have been successfully conducted[21]. Genetic methods have been developed to screen specifically for tumor suppressors in the fly using Flp/FRT mediated genetic mosaics[22]. Given the known conservation between *Drosophila* and mammalian *hh/ptc* pathways as well as many other developmental pathways, the fly remains an effective model system for exploring the nature of stem cell division and overproliferation.

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## REFERENCES

- [1] Nusslein-Volhard C, Wieschaus E. Mutations affecting segment number and polarity in *Drosophila*. Nature 1980; **287**:795-801.
- [2] Alcedo J, Ayzenzon M, Von Ohlen T, Noll M, Hooper JE. The *Drosophila smoothed* gene encodes a seven-pass membrane protein, a putative receptor for the *hedgehog* signal. Cell 1996; **86**:221-32.
- [3] Bhat KM. The *patched* signaling pathway mediates repression of *gooseberry* allowing neuroblast specification by *wingless* during *Drosophila* neurogenesis. Development 1996; **122**:2921-32.
- [4] Perrimon N. Hedgehog and beyond. Cell 1995; **80**:517-20.
- [5] Ingham P. Signaling by *hedgehog* family proteins in *Drosophila* and vertebrate development. Current Opinions in Genetics Dev 1995; **5**:528-34.
- [6] Heberlein U, Moses K. Mechanisms of *Drosophila* retinal morphogenesis: the virtues of being progressive. Cell 1995; **81**:987-90.
- [7] Goodrich LV, Johnson RL, Milenkovic L, McMahon JA, Scott MP. Conservation of the *hedgehog/patched* signaling pathway from flies to mice: induction of a mouse *patched* gene by Hedgehog. Genes Dev 1996; **10**:301-12.
- [8] Riddle RD, Johnson RL, Laufer E, Tabin C. Sonic *hedgehog* mediates the polarizing activity of the limb. Cell 1993; **75**:1401-16.
- [9] Forbes AJ, Lin H, Ingham PW, Spradling AC. *hedgehog* is required for the proliferation and specification of ovarian somatic cells prior to egg chamber formation in *Drosophila*. Development

- 1996; **122**:1125-35.
- [10] Forbes AJ, Spradling AC, Ingham PW, Lin H. The role of segment polarity genes during early oogenesis in *Drosophila*. *Development* 1996; **122**:3283-94.
- [11] Johnson RL, Rothman AL, Xie J, et al. Human homolog of *patched*, a candidate gene for the basal cell nevus syndrome. *Science* 1996; **272**:1668-71.
- [12] Hahn H, Wicking C, Zaphiropoulos PG, et al. Mutations of the human homolog of *Drosophila patched* in the nevoid basal cell carcinoma syndrome. *Cell* 1996; **85**:841-51.
- [13] Ruiz i Altaba A. Catching a Gli-mpse of Hedgehog. *Cell* 1997; **90**:193-6.
- [14] Margolis J, Spradling A. Identification and behavior of epithelial stem cells in the *Drosophila* ovary. *Development* 1995; **121**:3797-807.
- [15] Chiang C, Litingtung Y, Lee E, et al. Cyclopia and defective axial patterning in mice lacking *Sonic hedgehog* gene function. *Nature* 1996; **383**:407-13.
- [16] Oro AE, Higgins KM, Hu Z, Bonifas JM, Epstein E Jr., Scott MP. Basal cell carcinomas in mice overexpressing *Sonic hedgehog*. *Science* 1997; **276**:817-21.
- [17] Collins VP. Gene amplification in human gliomas. *Glia* 1995; **15**:289-96.
- [18] Dahmane N, Lee J, Robins P, Heller P, Ruizi Altaba A. Activation of the transcription factor *Gli1* and the *Sonic hedgehog* signalling pathway in skin tumours. *Nature* 1997; **389**:876-81.
- [19] Xie J, Johnson RL, Zhang X, et al. Mutations of the *PATCHED* gene in several types of sporadic extracutaneous tumors. *Cancer Res* 1997; **57**:2369-72.
- [20] Nusse R, Varmus HE. Wnt genes. *Cell* 1992; **69**:1073-87.
- [21] Gateff E, Mechler BM. Tumor-suppressor genes of *Drosophila melanogaster*. *Crit Rev Oncog* 1989; **1**:221-45.
- [22] Xu T, Wang W, Zhang S, Stewart RA, Yu W. Identifying tumor suppressors in genetic mosaics: the *Drosophila lats* gene encodes a putative protein kinase. *Development* 1995; **121**:1053-63.

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