

Cloning and sequencing of mouse GABA transporter complementary DNA

TAM ANTHONY C.W ., LI HE GUO^{*1}, DOMINIC MAN KIT LAM.

Hong Kong Institute of Biotechnology New Territories,
Hong Kong

* Shanghai Institute of Cell Biology, Chinese Academy of
Science

ABSTRACT

A cDNA encoding the mouse GABA transporter has been isolated and sequenced. The results show that the mouse GABA transporter cDNA differs from that of the rat by 60 base pairs at the open reading frame region but the deduced amino acid sequences of the two cDNAs are identical and both composed of 599 amino acids. However, the amino acid sequence is different from the sequence deduced from a recently published mouse GABA transporter cDNA.

Key words: *GABA transporter, cloning, sequencing, mouse*

INTRODUCTION

The GABA transporter is a membraneous protein responsible for the cellular uptake or re-uptake of the neurotransmitter, γ -aminobutyric acid, from the synaptic cleft and hence for the termination of its synaptic activity[1,2]. Complementary DNAs encoding the rat and human GABA transporters have been cloned and sequenced[3,4]. The rat cDNA has been expressed in *Xenopus* oocytes which then exhibited GABA accumulating property[5]. From the sequencing data, it was deduced that the rat cDNA encoded a protein of 599 amino acids with probably 11 to 13 transmembrane regions. The human GABA transporter was also deduced to be made up of the same number of amino acids and the two amino acids sequences

1. Corresponding author

Cloning and sequencing of mouse GABA transporter cDNA

showed high degree of homology[4]. Independently, we also obtained a cDNA fragment encoding the human GABA transporter from a human brain substantia nigra cDNA library by polymerase chain reaction(PCR) technique[3]. With this cDNA fragment as probe, a cDNA clone encoding the mouse GABA transporter was isolated from a mouse brain stem cDNA library and the cDNA was sequenced. While this work was in progress, the sequence of a mouse GABA transporter cDNA and the exon-intron structure of the mouse GABA gene were published by Liu et al.[4] A comparison of the published sequence and our sequencing data showed a considerable degree of discrepancy despite that both we and the other group used similar techniques and materials in obtaining our respective cDNAs. We report here the isolation and sequencing of our mouse GABA transporter cDNA. A comparison of this sequence with that of the rat cDNA and the published mouse cDNA is made and the significance discussed.

MATERIALS AND METHODS

An oligo dT primed mouse brain cDNA library in Lambda Uni-ZapTMXR obtained from Stratagene was used in this study. The library contained approximately 2.0×10^6 primary plaques with an average insert size of 1.5 Kb. A cDNA fragment of approximately 1.8 Kb in length encoding the human GABA transporter cloned by Lam et al.[3] was ^{32}P -labeled by nick translation and used as probe to screen the library by the Stratagene protocol. After screening and plaque purification, pBluescript plasmids with positive inserts were excised from the selected Lambda vectors and transformed into *E.coli* strain BB4. Plasmids purified from the transformants were cut by EcoR I and Xho I to release the inserts. Two oligonucleotides, primer I and primer II, which corresponded respectively to the 5' and 3' ends of the reading frame of the human GABA transporter cDNA and were originally designed by Lam et al.[3] for PCR experiments, were used to identify inserts with complete reading frame by Southern hybridization. The selected cDNA clone was then mapped by restriction enzymes. Overlapping restriction fragments obtained were cloned into plasmid pTZ18U and pTZ19U for dideoxy sequencing (Pharmacia T7 Sequencing Kit and Gene-ATAQ Sequencing Kit). Inserts too long to be completely sequenced once were shortened by Nuclease Ba131 before subcloning and sequencing. Sequencing data were analyzed using PC / Gene software on an IBM compatible personal computer

RESULT AND DISCUSSION

After screening 7.5×10^6 recombinant plaques plated out from the mouse brain cDNA library, five recombinants that gave comparatively stronger hybridization signals were selected out of some thirty positive plaques. Following phagemid excision and reinfection, the pBluescript derivatives were purified and the inserts excised. All inserts hybridized with primer II but only one hybridized with primer I, indicating that this was the only inserts with a complete reading frame of the GABA transporter. This insert, designated as MGAT, was the longest among the five and was about 4.2 Kb in length. It must be added here that the ease of obtaining the target cDNA indicates that the mouse brain is rich in GABA transporter mRNA.

Ten restriction enzymes were used to map MGAT. The result is shown in Fig 1. This map is very much different from that of the rat GABA transporter cDNA

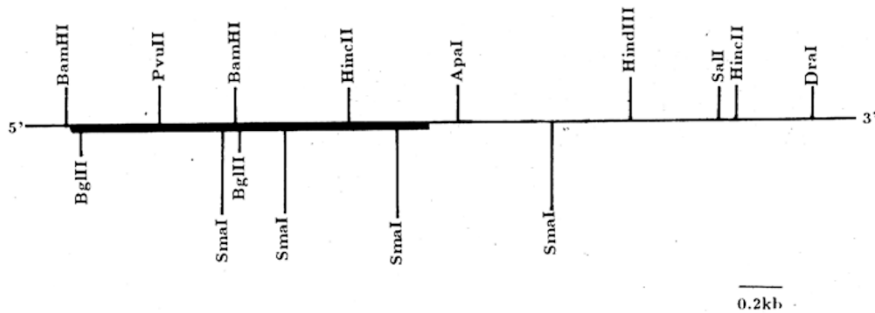


Fig 1. Physical map of MGAT, a cDNA clone encoding the GABA transporter from mouse brain. The open reading frame is marked by a bar. MGAT-1 was ligated to the pbluescript vector through an EcoR I adaptor at the 5' end and an Xho I adaptor at the 3' end.

(data not shown). Sequencing results shows that the cDNA consists of a long 3' noncoding region of 2.3 Kb and a complete open reading frame of 1797 bp. The sequence of the 5' region including the open reading frame was compared with the sequences of the rat i.e. composed of 599 amino acids. The reading frames of the two cDNAs differs from each other by 60bp. However, with the exception of one bp, all of the differences occur at the third position of a codon and result in no variation in the two amino acid sequences. The difference at nucleotide 1308 (G versus C) which is at the first position of a codon for leucine also results in no amino acid divergence. On the other hand, the mouse GABA transporter amino acid sequence as deduced from the published cDNA sequence of Liu et al.[4] is amino acid shorter i.e. Pro 213 is missing. In addition, it differs from the other two GABA transporters[1, 3, 5] by 7 amino acids, despite that there is only a 14 bp difference between the open reading frames of the two mouse cDNA sequences.

The human GABA transporter differs from the rat GABA transporter by 17 amino acids, but all except one of those residues are located at the N- and C-terminal regions of the proteins[5]. In a region starting from Asp 40 to Phe 512 (473 A.A. in length), the two amino acids sequences are almost identical with only leu 179 in rat replaced by a methionine in human. The conservation of the GABA transporter in rat and human indicated that the conformation of the protein is important to its functioning. Therefore it is within our expectation to found that the mouse GABA transporter (MGAT) amino acid sequence is identical to that of the rat. We are surprised to learn that the mouse GABA transporter amino acid sequence

540 CTGTACAACTCCTTCAACCAGCCCTGCCATGGAAAACAGTGTGACAAACCCGTGGAACACATGACCCGCTGCTTCTCCAACACTACAGCCTGGTC
 -----T-----G-----C-----T-----A-----
 180 LeuTyrAsnSerPheThrThrLeuProTrpLysGlnCysAspAsnProTrpAsnThrAspArgCysPheSerAsnTyrSerLeuVal
 -----T-----A-----A-----A-----G-----A-----
 630 AATACCACCAATGACCAGCCCGTGGTGGAGTTCTGGGAGCGCAACATGCACCAGATGACAGATGGACTGGACAAGCCAGGACAGATC
 -----T-----A-----A-----A-----G-----A-----
 210 AsnThrThrAsnMetThrSerAlaValValGluPheTrpGluArgAsnMetHisGlnMetThrAspGlyLeuAspLysProGlyGlnIle
 -----T-----A-----A-----A-----G-----A-----
 720 CGTGGCCTCTGGCCATCACACTGGCCATTGCCCTGGGTGCTCGTGTATTTCTGCATCTGGAAGGTGTGGTTGGACTGGAAGGTTGGTC
 -----G-----A-----T-----T-----A-----A-----
 240 ArgTrpProLeuAlaIleThrLeuAlaIleAlaTrpValLeuValTyrPheCysIleTrpLysGlyValGlyTrpThrGlyLysValVal
 -----T-----A-----A-----A-----G-----A-----
 810 TACTTCTCAGCCACGTAACCCCTACATCATGCTTATCATCTGTCTTCCGTGGAGTGACGCTTCCCGGGCCCAAGGAGGGGATCCCTCTTC
 -----A-----C-----A-----T-----A-----T-----
 270 TyrPheSerAlaThrTyrProTyrIleMetLeuIleIleLeuPhePheArgGlyValThrLeuProGlyAlaLysGluGlyIleLeuPhe
 -----A-----C-----A-----T-----A-----T-----
 900 TACATCACACCAACTTCGAAAGCTGTCTGATTCAGAGGTGGCTTGACGGCCGCCACCCAGATCTTCTCTCCTACGGGCTGGGCCCTG
 -----G-----A-----T-----T-----A-----A-----
 300 TyrIleThrProAsnPheArgLysLeuSerAspSerGluValTrpLeuAspAlaAlaThrGlnIlePhePheSerTyrGlyLeuGlyLeu
 -----A-----T-----T-----T-----A-----A-----
 990 GGGTCCCTGATTGCTCTGGGAAGCTACAACCTTTCCACAACAATGTGTACAGGGACTCCATCATCGTTTGTGTCATCAACTCCTGCACC
 -----T-----T-----T-----T-----A-----T-----
 330 GlySerLeuIleAlaLeuGlySerTyrAsnSerPheHisAsnAsnValTyrArgAspSerIleIleValCysCysIleAsnSerCysThr
 -----T-----T-----T-----T-----A-----T-----
 1080 AGCATGTTGCCGGATTCTGTCATCTTCTCCATCGTGGGCTTCATGGCTCATGTCCACCAAGAGTCCATAGCTGATGTGGCAGCCTCAGGC
 -----C-----C-----C-----C-----A-----C-----
 360 SerMetPheAlaGlyPheValIlePheSerIleValGlyPheMetAlaHisValThrLysArgSerIleAlaAspValAlaAlaSerGly
 -----T-----T-----T-----T-----A-----T-----

Cloning and sequencing of mouse GABA transporter cDNA

1170
 CCGGGGTGGCATTCTTGGCGTACCCTGAGGCTGTGACACAGTACCACATCTCTCCCTCTGGGCTATCCTCTTCTTCTCCATGCTGCTG
 --T--A-----A-----C-----
 390
 ProGlyLeuAlaPheLeuAlaTyrProGluAlaValThrGlnLeuProIleSerProLeuTrpAlaIleLeuPhePheSerMetLeuLeu

 1260
 ATGCTGGCATTGACAGCCAGTTCTGTACCGTGGAGGGTTTCACTACCTGCCCTGGTGAGCAGTACCCACAGACTTCTCCGCAATCGCCCGT
 -----G-----T-----C-----
 420
 MetLeuGlyIleAspSerGlnPheCysThrValGluGlyPheIleThrAlaLeuValAspGluTyrProArgLeuLeuArgAsnArgArg

 1350
 GAACTCTTCAITGCTGCCGTGTGCATCGTGTCTACCTGATTGGCCTGTCTAACATCACCCAGGGTGGCAATTATGTCTTCAAACCTGTTT
 -----C-----
 450
 GluLeuPheIleAlaAlaValCysIleValSerTyrLeuIleGlyLeuSerAsnIleThrGlnGlyIleTyrValPheLysLeuPhe

 1440
 GATTATTACTCTGCCAGCGCATGAGCTTGCTGTTCCTGGTTTTCTTCGAGTGTGTCTCCATTTCCCTGGTTTTTATGGTGTCAACCCGGTTC
 -----T-----C-----T-----
 480
 AspTyrTyrSerAlaSerGlyMetSerLeuLeuPheLeuValPhePheGluCysValSerIleSerTrpPheTyrGlyValAsnArgPhe

 1530
 TATGACAACATCCAGGAGATGGTTCGCTCCAGGCCCTGCACTGGTGGAAAGTGTGTGGTCCCTTTTTTTCACACCCATCATTTGTGGCGGGC

 510
 TyrAspAsnIleGlnGluMetValGlySerArgProCysIleTrpTrpLysLeuCysTrpSerPhePheThrProIleIleValAlaGly

 1620
 GTGTTTCTCTCAGTGTGCAGATGACACCCTCACCATGGGAAGCTATGTTTCCCCAAGTGGGCCAGGGCGTGGGCTCATG
 -----C-----
 540
 ValPheLeuPheSerAlaValGlnMetThrProLeuThrMetGlySerTyrValPheProLysTrpGlyGlnGlyValGlyTrpLeuMet

 1710
 GCTCTGCTCCATGGTGTCTATCCCCGGGTACATGGCTTACATGTCTCCCTCACCCCTGAAGGGCTCCCTGAAGCAGCGTCTCCAGGTCATG
 -----A-----T-----
 570
 AlaLeuSerSerMetValLeuIleProGlyTyrMetAlaTyrMetPheLeuThrLeuLysGlySerLeuLysGlnArgLeuGlnValMet

```

ATTAGCCAGTGAAGATATTGTGGCCCTGAGAAATGGCCCTGAGCAGCCGCGAGGCTGGCAGCTCAGCCAGCAAGGAGGCTACATCTAG 1800
-----C-----T-----
IleGlnProSerGluAspIleValArgProGluAsnGlyProGluGlnProGlnAlaGlySerSerAlaSerLysGluAlaTyrIle 599
-----
GGGTGCAGCCCCCCCATCACCCCTACACTGGCACTCTGGACTGGCTGTACCCACACCCCTTGAAAGACTGAAGATACTCTGTCTCCACC 1890
-----T-----C-----A-----
TACCTCAGGGGCGAGTCCAGACACCATCACCATGCAGAGAGGG
-----G-----G-----

```

Fig 2. Nucleotide and deduced amino acid sequences of the mouse GABA transporter cDNA, MGAT. Aligned with the MGAT sequence is the rat GABA transporter cDNA sequence as reported by Guastella et. al. and the mouse GABA transporter cDNA sequence as reported by Liu et al. Dashes in the sequences represent nucleotides identical to that of MGAT. The first nucleotide at the start site is designated as position one. Below is the amino acid sequence deduced from MGAT and aligned with it is the mouse GABA transporter amino acids sequence as reported by Liu et al. Dashes in the last sequence represent amino acid residues identical to that deduced from MGAT.

Cloning and sequencing of mouse GABA transporter cDNA

as published by Liu et al.[4] differs from that of the rat by such extend and that all discrepancies occur at the central conserved region of the peptides. A proline residue, which is present at position 213 of the GABA transporter sequence of rat but not at that of the published mouse sequence, is known to be rigid and when occurs on a polypeptide is likely to create a rigid bend. Two bulky tryptophan residues in rat are replaced by two comparatively smaller residues in mouse (Cys 212 and Ile 285) and leu 118 in rat is replaced by a tryptophan residue in mouse. All these would result in conformational differences between the two transporters.

As we worked on our sequencing of MGAT, we encountered problems due to band compressions and occurrence of secondary structure in the templates. These problems were subsequently solved by various means[8]. We are sure that the discrepancies in the sequencing results as reported by us and by Liu et al.[4] were not due to our misreadings of the sequence. As the presence of a variety of GABA transporter subtypes has been suggested[7], we do not rule out that we and the other group were dealing with different subtypes of the GABA transporter in mouse. We have now isolated a fragment and are in the process of sequencing both the exons and introns regions. By comparing the exon-intron structure of our genomic fragment with that as reported by Liu et al. [4], we will be able to tell whether there exist two GABA transporter genes in mouse.

REFERENCES

- [1] Guastella J, Nelson N, Nelson H, et al. Cloning and expression of a rat brain GABA transporter. *Science* 1990; **249**:1303-6.
- [2] Kanner BI, Schuldiner S. Mechanism of transport and storage of neurotransmitters. *CRC Crit. Rev. Biochem.* 1987; **22**:1-38.
- [3] Lam DMK, Fei J, Zhang XY, et al. Molecular cloning and structure of the human (GABATHG) GABA transporter gene. *Molecular Brain Research* 1993; **19**:227-32.
- [4] Liu QR, Mandiyan S, Nelson H, Nelson N. A family of genes encoding neurotransmitter transporters. *PNAS* 1992; **89**:6639-43.
- [5] Nelson H, Mandiyan S, Nelson N. Cloning of the human GABA transporter. *FEBS Letter* 1990; **269**:181-4.
- [6] Radian R, Bendahan A, Kanner BI. Purification and identification of the functional sodium- and chloride-coupled-aminobutyric acid transport glycoprotein from rat brain. *J Biol Chem.* 1986; **261**:15437-41.
- [7] Wood JD, Sidhu HS. A comparative study and partial characterization of multi-uptake systems for -aminobutyric acid. *J Neurochem.* 1987; **49**:1202-8.
- [8] The T7 Sequencing™ Kit from Pharmacia was used for routine sequencing. The band compressions problems were solved either by substituting 7-deaza dGTP and 7-deaza DAPT for dGTP and dATP in the sequencing mixes (supplied as Deaza G?A T7 Sequencing™ Mixes by Pharmacia) or by sequencing the templates from both ends. The problems due to secondary structures were solved by using the Gene-ATAQ™ Sequencing Kit from Pharmacia which employs high temperature and Taq DNA polymerase in the sequencing reactions.

Received 3-1-1994 Revised 19-3-1994 Accepted 8-4-1994