

# C-terminal domains of histone demethylase JMJ14 interact with a pair of NAC transcription factors to mediate specific chromatin association

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**Jumonji C (JmjC) domain-containing protein 14 (JMJ14) is an H3K4-specific histone demethylase that has important roles in RNA-mediated gene silencing and flowering time regulation in *Arabidopsis*. However, how JMJ14 is recruited to its target genes remains unclear. Here, we show that the C-terminal FYRN (FY-rich N terminus) and FYRC (FY-rich C terminus) domains of JMJ14 are required for RNA silencing and flowering time regulation. Chromatin binding of JMJ14 is lost upon deletion of its FYRN and FYRC domains, and H3K4me3 is increased. FYRN and FYRC domains interact with a pair of NAC (NAM, ATAF, CUC) domain-containing transcription factors, NAC050 and NAC052. Genome-wide chromatin immunoprecipitation analysis revealed that JMJ14 and NAC050/052 share a set of common target genes with CTTGNNNNCAAG consensus sequences. Mutations in either *NAC052* or *NAC050* impair RNA-mediated gene silencing. Together, our findings demonstrate an important role of FYRN and FYRC domains in targeting JMJ14 through direct interaction with NAC050/052 proteins, which reveals a novel mechanism of histone demethylase recruitment.**

**Keywords:** JMJ14; histone demethylase; NAC050; NAC052; transgene silencing

*Cell Discovery* (2015) 1, 15003; doi:10.1038/celldisc.2015.3; published online 28 April 2015

## Introduction

Histone lysine methylation has important roles in diverse biological functions including heterochromatin formation, transcriptional gene silencing, transcriptional activation, DNA repair, and DNA recombination [1–3]. Histone methylation can be removed by Jumonji C (JmjC) domain-containing proteins (JMJs) and LSD1 family proteins [4]. In *Arabidopsis*, some JmjC domain-containing proteins have been shown to

remove methyl groups from different lysine residues of histone H3, thereby participating in controlling transcriptional repression and DNA methylation for genome stability, and in regulation of plant development [5–16]. JmjC domain-containing protein 14 (JMJ14) has been demonstrated to demethylate tri-, di-, and mono-methylated H3K4 [9–11]. JMJ14 contributes to flowering time regulation through repression of the florigen *FLOWERING LOCUS T (FT)*, the floral integrators *APETALA1 (API)*, *SUPPRESSOR OF CO OVEREXPRESSION 1 (SOC1)*, and *LFY* [10]. Additional lines of evidence indicate that JMJ14 is also involved in transgene silencing that acts on cell-to-cell movement of an RNA silencing signal, reduction of transgene transcription, maintenance of endogenous transposon silencing, and DRM2-mediated CHH DNA methylation [12, 14, 16]. *jmj14*

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Received 11 January 2015; accepted 19 January 2015

mutants suppress post-transcriptional gene silencing at a variety of targets, including JAP3 loci carrying a pSUC2-driven *PHYTOENE DESATURASE* (*PDS*) inverted repeat transgene and a 35S-driven  $\beta$ -glucuronidase transgene (L1) undergoing sense transgene silencing (S-post-transcriptional gene silencing) [12, 16]. In *jmj14* mutants, transcription levels of transgene loci were downregulated, whereas several known endogenous targets of JMJ14 are upregulated [16].

JMJ14 belongs to the KDM5/JARID1 subfamily of JmjC proteins, in which the plant and animal counterparts contain distinct domains: animal and plant proteins contain JmjN, JmjC, and C5HC2 zinc-finger domains, while the AT-rich interaction domain (ARID) and plant homeodomain (PHD) domains existing in members of yeast and animals are missing in most of the plant proteins [17]. There are several PHD domain subtypes in mammals and yeast, among which the first PHD domain of Jarid1C in human and the second PHD domain of Lid2 in yeast were reported to recognize methylated H3K9 [18, 19]. In *Arabidopsis*, all active KDM5 members, including JMJ14, JMJ15, JMJ16 and JMJ18–lack PHD domains, but contain an F/Y-rich N terminus (FYRN) and an F/Y-rich C terminus (FYRC) domains in the C terminus [2, 10, 17, 20]. FYRN and FYRC domains are rich in phenylalanine/tyrosine residues, which are found in many chromatin-related proteins, such as mixed lineage leukemia 1 and mixed lineage leukemia 2, two histone H3K4 methyltransferases in human [21, 22]/nuclear interactor of ARF and MDM2, a growth inhibitory protein involved in maintaining chromosomal stability [23], ATX1, an H3K4 methyltransferase involved in plant development and stress responses [24, 25], and AtMBD9, a methyl-CpG binding domain-containing protein involved in plant development [26].

In this study, we find that FYRN and FYRC domains are essential for JMJ14 function. These two domains have critical role in JMJ14 binding to its genomic targets through interacting with transcription factors NAC050 and NAC052. Thus, our findings reveal a novel mechanism of targeting a histone demethylase to its functional loci and show the importance of transcription factors in epigenetic regulation.

## Results

### *FYRN and FYRC domains are critical for the biological functions of JMJ14*

To analyze whether the FYRN and FYRC domains of JMJ14 are important for its regulation in flowering

time and transcriptional regulation, we transformed *jmj14-1* mutants with translational fusion constructs consisting of either *JMJ14* genomic DNA (JMJ14-HA) or a truncated *JMJ14* without FYRN and FYRC domains (JMJ14 $\Delta$ FYR-HA), each containing an Influenza Hemagglutinin (HA) epitope tags and driven by the native *JMJ14* promoter. We selected two transgenic lines from each constructs for further analysis, in which the expression levels of *JMJ14* were similar to that in wild-type Columbia (Col; Supplementary Figure S1). JMJ14-HA plants could rescue the early-flowering phenotype of *jmj14*, whereas the JMJ14 $\Delta$ FYR-HA plants could not (Figure 1a). Moreover, the expression level of *FT* in these lines was also in line with their flowering time (Figure 1b).

Jawohl:AtSuc2:PDS (JAP) plants contain a transgene that triggers inverted repeat post-transcriptional gene silencing of the endogenous *PDS* gene and show a JMJ14-dependent photobleaching phenotype [12]. We crossed different JMJ14 transgenic lines to JAP *jmj14-1*, in which the photobleaching phenotype of JAP plants was suppressed by *JMJ14* mutation. The result showed that JMJ14-HA but not JMJ14 $\Delta$ FYR-HA can derepress the photobleaching phenotype of *jmj14-1* (Figure 1c). Taken together, these evidence demonstrate that FYRN and FYRC domains are critical for JMJ14 in regulation of flowering time and transgene silencing.

### *FYRN and FYRC domains are required for genome-wide JMJ14 targeting*

To gain a broader understanding of how FYRN and FYRC domains affect JMJ14 function, we performed chromatin immunoprecipitation (ChIP) followed by sequencing (ChIP-seq) with anti-H3K4me3 antibody in *jmj14-1*, JMJ14-HA *jmj14*, JMJ14 $\Delta$ FYR-HA *jmj14*, and wild-type Col. We identified 221 genes in *jmj14* and 262 genes in JMJ14 $\Delta$ FYR-HA *jmj14* that showed H3K4me3 hypermethylation, among which 177 genes are common between the two data sets (Figure 2a). This indicates that JMJ14 $\Delta$ FYR-HA cannot rescue the H3K4me3 hypermethylation phenotype in the *jmj14* mutant, while H3K4me3 levels of these 177 genes were completely recovered in JMJ14-HA *jmj14* to wild-type Col level (Figure 2b). Interestingly, JMJ14 $\Delta$ FYR-HA showed normal H3K4me3 demethylation activity when overexpressed (Supplementary Figure S2). These results suggest that FYRN and FYRC domains may be essential for recruiting JMJ14 to its endogenous target genes as opposed to being required for enzymatic activity, *per se*.

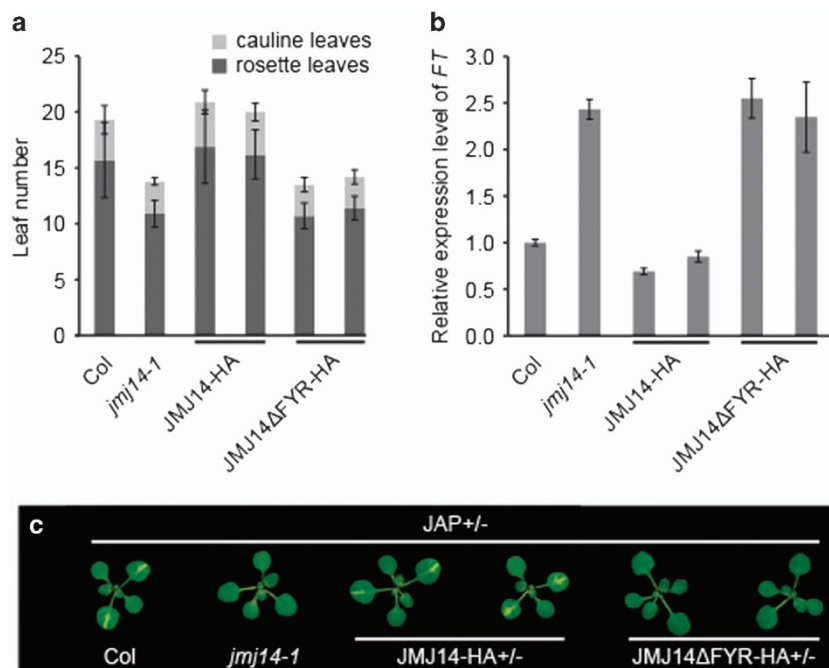
To further determine whether the FYRN and FYRC domains mediate the global targeting of JMJ14, we performed ChIP-seq with anti-HA in both the JMJ14-HA *jmj14* and JMJ14 $\Delta$ FYR-HA *jmj14* lines. By analyzing the ChIP-seq results in JMJ14-HA *jmj14* transgenic plants, we identified 761 target genes of JMJ14. Strikingly, the binding signals on JMJ14 target genes were completely lost in JMJ14 $\Delta$ FYR-HA *jmj14*, which is reminiscent of *jmj14* mutant (Figure 2c). Altogether, these results show that FYRN and FYRC domains are essential for proper targeting of JMJ14 to chromatin.

In *Arabidopsis*, genome-wide distribution patterns of H3K4me3 are found to be exclusively genic and predominantly localized at transcriptional starting sites [10]. Interestingly, we found that JMJ14 occupied the whole gene bodies in a pattern strikingly different from that of H3K4me3 in all the target genes. When more transgenic lines were used for ChIP-quantitative PCR (qPCR) analysis, all target genes tested were validated (Supplementary Figure S3). For example, *At1g21290*, which codes a *copia*-like retrotransposon, is one of the JMJ14 targets (Figures 2d and e). In wild-

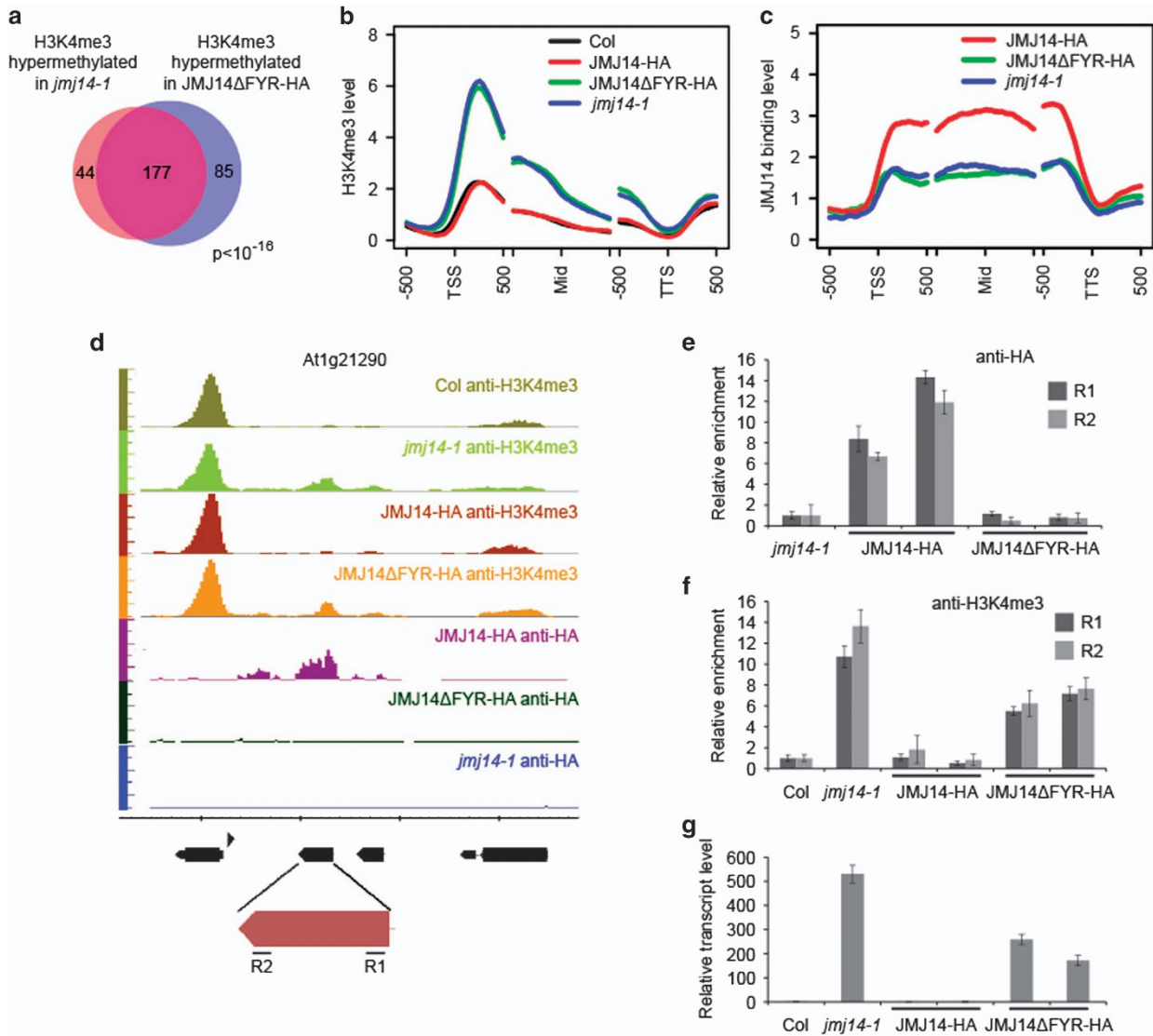
type and JMJ14-HA *jmj14-1* plants, the H3K4me3 at *At1g21290* was very low, indicating that this transposable element was silenced in normal condition (Figures 2d and f). Hypermethylation of H3K4me3 coupled with increase of gene expression was observed in both *jmj14-1* and JMJ14 $\Delta$ FYR-HA *jmj14-1* plants compared with that in Col and JMJ14-HA *jmj14-1* plants (Figures 2d and g). These results imply that FYRN and FYRC domains are essential for JMJ14 recruitment to remove H3K4me3 and repress transcription.

#### JMJ14 interacts with NAC050 and NAC052 through its FYRN and FYRC domains

To identify potential interacting proteins of JMJ14, a yeast two-hybrid screening was performed using the full-length JMJ14 as the bait. From this screening, we identified a pair of putative transcription factors, NAC050 and NAC052 as the top hits. Independently, we created a 3  $\times$  flag tagged version of JMJ14 driven by its endogenous promoter, and used this line to perform immunoprecipitation followed by mass spectrometry. NAC050 and NAC052 were among the



**Figure 1** The FYR (FYRN+FYRC) domain is important for the biological function of JMJ14. **(a)** The flowering times of Columbia (Col), *jmj14-1* and different JMJ14 (Jumonji C (JmjC) domain-containing protein 14) complementary lines under long day condition (16 h light, 8 h dark) at 23 °C. Flowering time was assessed by counting the number of rosette and cauline leaves when the plants flowered. **(b)** The expression level of flowering locus T (*FT*) in different JMJ14 complementary lines compared with Col and *jmj14-1*. **(c)** The JAP/*jmj14-1* homozygous plants were crossed with *jmj14-1* and different JMJ14 complementary lines. The leaf phenotypes of F1 plants were observed. The JAP<sup>+/-</sup> plants under Col background were used as controls. JMJ14-HA and JMJ14 $\Delta$ FYR-HA indicates the  $P_{JMj14}::JMj14-HA$  *jmj14-1* and  $P_{JMj14}::JMj14\Delta FYR-HA$  *jmj14-1* transgenic lines, respectively.

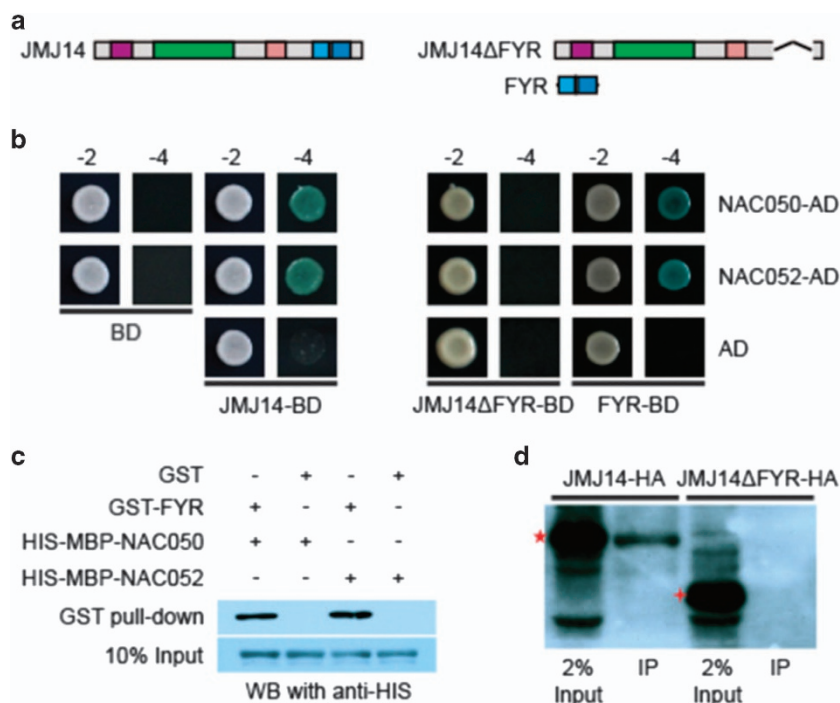


**Figure 2** JMJ14 (Jumonji C (JmjC) domain-containing protein 14) lost its targets on chromosome in the absent of FYR (FYRN +FYRC) domain. (a) The target genes with twofold increase of H3K4me3 in *JMJ14ΔFYR-HA* *jmj14-1* transgenic plants are significantly overlapped with those in *jmj14-1*. (b) The H3K4me3 pattern in Columbia (Col), *JMJ14-HA* *jmj14-1*, *JMJ14ΔFYR-HA* *jmj14-1*, and *jmj14-1* plants. TSS, Mid and TTS refers to transcription start site, middle of gene, and transcription termination site, respectively. Genes used for analysis were the common 177 hypermethylated genes of *jmj14-1* and *JMJ14ΔFYR-HA* *jmj14-1*. (c) The JMJ14 binding level on the direct target genes of JMJ14 in *JMJ14-HA* *jmj14-1*, *JMJ14ΔFYR-HA* *jmj14-1*, and *jmj14-1* plants. The tag counts were normalized in each bin according to the total number of reads. (d) The anti-HA and anti-H3K4me3 chromatin immunoprecipitation sequencing (ChIP-seq) results for *At1g21290* locus on genome browser. (e) The anti-HA ChIP-qPCR validation for *At1g21290* locus. (f) The anti-H3K4me3 ChIP-qPCR validation for *At1g21290* locus. (g) Detection of *At1g21290* transcripts by RT-quantitative PCR. Two independent lines of each transgenic plants were used in the qPCR. R1 and R2 indicate the 5' and 3' regions of the gene showed in d.

most abundant interacting proteins (Supplementary Table S1). NAC050 and NAC052 belong to a NAC (NAM, ATAF, CUC) transcription factor superfamily, which is specific to plants. There are >100 predicted members of NAC family containing the conserved NAC domain in *Arabidopsis* [27, 28]. NAC050 and NAC052 are encoded by tandem genes

*At3g10480* and *At3g10490*, respectively. Sequence alignment revealed that NAC050 and NAC052 share high homology, with 88% identity (Supplementary Figure S4). In order to dissect the nature of the interaction, we co-expressed full-length coding sequence (CDS) of *NAC050* or *NAC052* fused with *GAL4-AD* and *JMJ14-GAL4-BD* into yeast AH109 strains, and





**Figure 3** JMJ14 (Jumonji C (JmjC) domain-containing protein 14) interacts with NAC050 and NAC052 through its FYR (FYRN + FYRC) domain *in vitro* and *in vivo*. (a) The schematic representation of constructs used for yeast two hybrids. (b) The full-length JMJ14 interacts with NAC050 and NAC052 in yeast (left). Deletion of FYR domain completely abolishes the interaction between JMJ14 and NAC050/NAC052 while FYR domain alone could interact with NAC050/NAC052 in yeast (right). ‘-2’ indicates SD/-Trp-Leu medium, ‘-4’ indicates SD/-Trp-Leu-His-Ade medium with X- $\alpha$ -Gal. (c) Recombinant His-maltose binding protein-NAC050/052 interact with glutathione S-transferase (GST)-FYR directly but not GST *in vitro*. (d) JMJ14, but not JMJ14ΔFYR, was immunoprecipitated by anti-NAC detected by western blot (WB). JMJ14-HA refers to the  $P_{JMJ14}::JMJ14-HA$  *jmj14-1* transgenic lines and JMJ14ΔFYR-HA refers to the  $P_{JMJ14}::JMJ14\Delta FYR-HA$  *jmj14-1* transgenic lines. The red star and the red cross indicate JMJ14-HA and JMJ14ΔFYR-HA fusion proteins, respectively, detected by anti-HA.

found these combinations activated the ADE2, HIS3, and lacZ reporter genes, further confirming our previous results (Figures 3a and b). To determine which part of JMJ14 mediates the interaction between JMJ14 and NAC050/NAC052, we generated a series of constructs with deletions of different domains of *JMJ14*, including JMJ14ΔJmjN, JmjN, JMJ14ΔJmjC, JmjC, JMJ14ΔZnF, ZnF, JMJ14ΔFYR, and FYR-only fused with *GAL4-BD*, respectively (Figure 3b and Supplementary Figure S5A). We found that the interaction between JMJ14 and NAC050/NAC052 was completely abolished after deletion of FYRN and FYRC domains (Figure 3b), whereas deletion of other domains had little effect (Supplementary Figure S5B). We further proved that the FYRN and FYRC domains were sufficient to mediate the interaction between JMJ14 and NAC050/NAC052 in yeast (Figure 3b), indicating that the FYRN and FYRC domains are responsible for the interaction. Interestingly, we found that only FYRN or FYRC

domain alone could not interact with NAC050 and NAC052 (Supplementary Figure S5B), suggesting that FYRN and FYRC domains function as an integral. To further verify this interaction, we performed pull-down assays using GST (glutathione S-transferase)-tagged JMJ14 FYRN and FYRC domains as ‘bait’ and maltose binding protein-His-tagged NAC050 or NAC052 as ‘prey’. GST pull-down and immunoblotting assays showed that FYRN and FYRC domains of JMJ14 indeed interact directly with NAC050 and NAC052 *in vitro* (Figure 3c).

In *Arabidopsis*, both NAC050 and NAC052 localized to the nucleus, as with JMJ14 (Supplementary Figure S6A). Moreover, these two proteins and JMJ14 also shared similar expression pattern by promoter::GUS analysis (Supplementary Figure S6B–D), suggesting that JMJ14 and NAC050/052 may act together *in vivo*. We further confirmed the interaction *in vivo* by co-immunoprecipitation assays. Antibodies that could recognize both NAC050 and NAC052 were

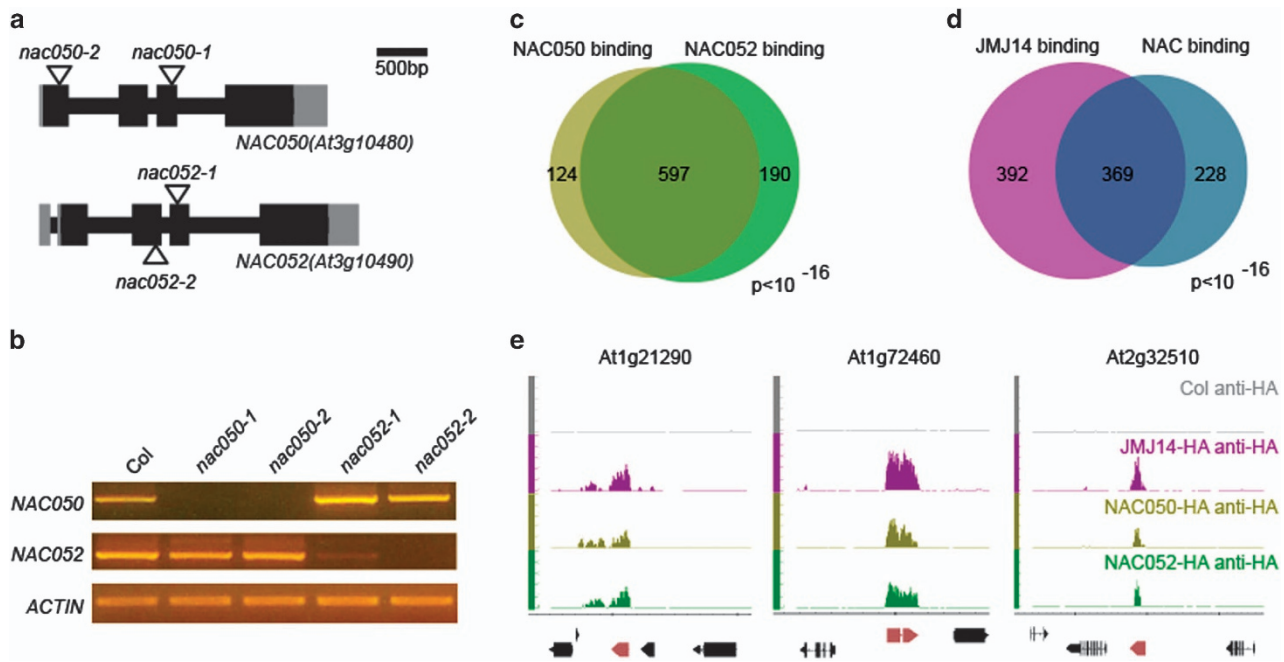
used to immunoprecipitate proteins from lysate of 12-day-old seedlings containing either JMJ14-HA or JMJ14ΔFYR-HA, respectively. Only the full-length JMJ14-HA could be immunoprecipitated but not the truncated JMJ14 without FYRN and FYRC domains (JMJ14ΔFYR-HA; Figure 3d), confirming that JMJ14 interacts with NAC050/052 in *Arabidopsis* and such interaction is dependent on the FYRN and FYRC domains.

*NAC050 and NAC052 share widespread common targets with JMJ14 in direct binding and gene regulation*

NAC family proteins have been shown to have important roles in almost every aspects of plant growth and development, including vegetative and floral development, root formation, auxin signaling, and biotic and abiotic stress response [29]. To determine the biological functions of NAC050 and NAC052 and their relationship with JMJ14, we identified the T-DNA insertion mutants from ABRC and WiscDsLox T-DNA Collection (Figure 4a), in which

*nac050-1*, *nac050-2*, and *nac052-2* are null alleles, whereas *nac052-1* represents a weak allele with partial reduction of full-length cDNA (Figure 4b).

As NAC050 and NAC052 interact with JMJ14, we hypothesized that NAC050 and NAC052 may have common targets with JMJ14 in the genome. We therefore performed ChIP-seq analysis using anti-HA antibody in *P<sub>NAC050</sub>::NAC050-HA nac050-1* and *P<sub>NAC052</sub>::NAC052-HA nac052-2*, respectively. The wild-type Col plants that do not contain HA tags were used as a negative control. We identified 721 and 787 target genes of NAC050 and NAC052, respectively, and more than three quarters (597) are common targets (Figure 4c), among which, >60% (369 out of 597) are also bound by JMJ14 (Figure 4d). Moreover, the binding signals of NAC050 and NAC052 distributed across the whole gene body of their targets as a similar pattern as that of JMJ14 (Supplementary Figure S7). The common targets of JMJ14, NAC050, and NAC052 were predominantly enriched in negative regulation of molecular function and protein



**Figure 4** NAC050 and NAC052 share common targets in direct binding with JMJ14 (Jumonji C (JmjC) domain-containing protein 14). **(a)** Gene structure of *NAC050* and *NAC052* and the T-DNA insertion sites of *nac050* and *nac052* mutants. Black bars, gray bars and black lines indicate coding exons, UTRs and introns, respectively. T-DNA insertions are indicated by triangles. Scale bar, 500 bp. **(b)** Detection of full-length CDS of *NAC050* and *NAC052* in the *nac050* and *nac052* mutants. *Actin* was used as an internal control. **(c)** The targets of NAC050 and NAC052, identified by chromatin immunoprecipitation sequencing (ChIP-seq), are significantly overlapped. **(d)** Nearly half of JMJ14’s targets are co-localized by both NAC050 and NAC052. **(e)** Three target genes showed co-localized ChIP-seq signal of JMJ14, NAC050, and NAC052 on genome browser. JMJ14-HA, NAC050-HA, and NAC052-HA indicates the *P<sub>JMJ14</sub>::JMJ14-HA jmj14-1*, *P<sub>NAC050</sub>::NAC050-HA nac050-1*, and *P<sub>NAC052</sub>::NAC052-HA nac052-2* transgenic lines, respectively.

phosphorylation in terms of biological processes (Supplementary Figure S8), indicating that they may be involved in signal transduction. For example, the binding signals of JMJ14, NAC050, and NAC052 exhibit significant overlap on target genes *At1g72460* and *At2g32510*, which encode two kinase proteins; the previously identified transposable elements *At1g21290* showed a similar pattern as well (Figure 4e). The significant overlapping of target genes suggests that NAC050 and NAC052 may function redundantly and act together with JMJ14 to collaboratively regulate a large number of genes in *Arabidopsis*.

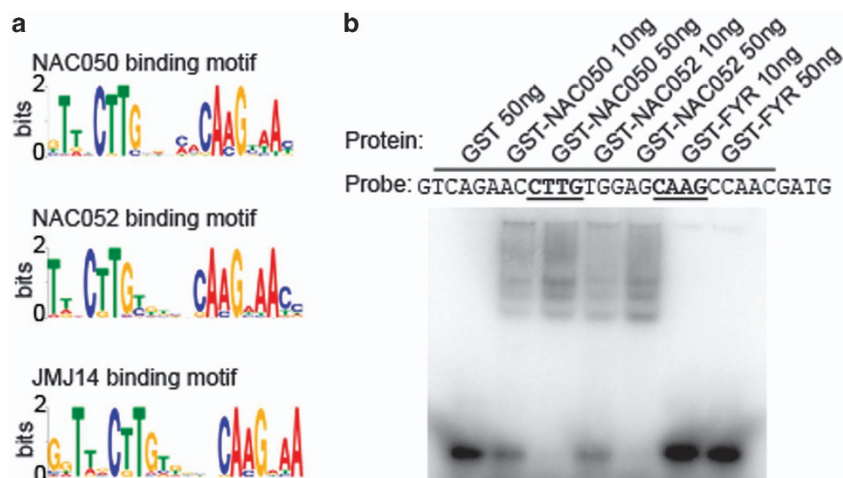
#### *NAC050 and NAC052 bind a specific palindromic DNA motif*

Previous reports showed that NAC transcription factors can be recruited by cis-elements on their target genes [30, 31]. Therefore, we speculated that NAC050 and NAC052 could also bind specific DNA sequences. To identify the DNA motif that was bound by NAC050 and NAC052, we analyzed the DNA sequences around the binding peaks of NAC050 and NAC052 within the genome using MEME-ChIP programs [32]. A palindromic sequence CTTGN NNNNCAAG intensively exists in both NAC050 and NAC052 binding peaks (Figure 5a), which is also enriched in JMJ14 binding peaks (Figure 5a). To determine the binding ability of NAC050 and NAC052 *in vitro*, we synthesized a 30-bp DNA probe that contains the CTTGNNNNNCAAG core sequence in *At1g72460*, one of the common targets of NAC050,

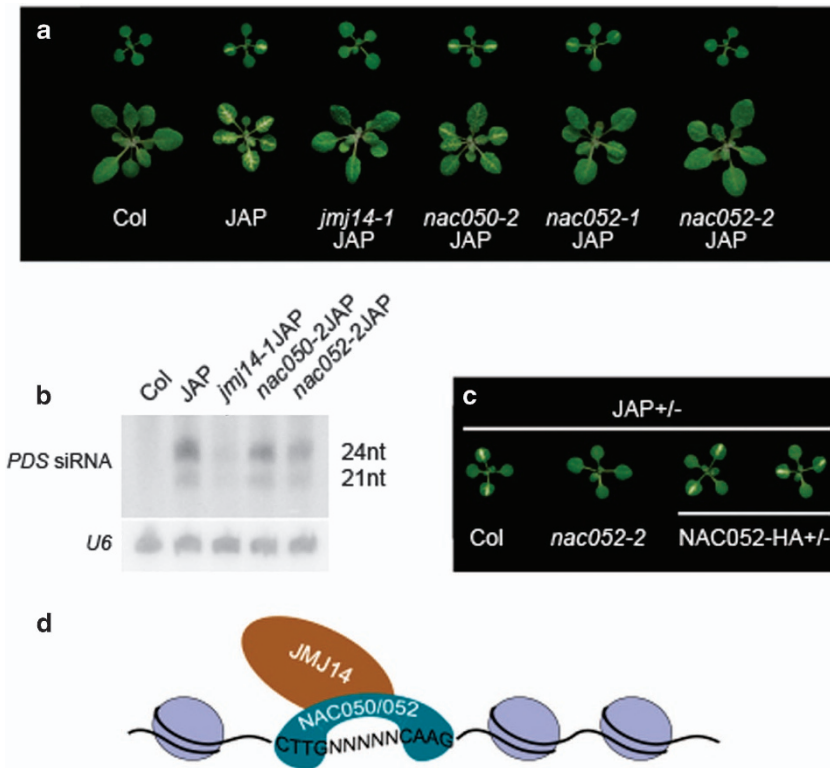
NAC052, and JMJ14, for electrophoretic mobility shift assays. In contrast to GST-FYR and GST alone, which did not bind this sequence, both NAC050 and NAC052 proteins displayed strong binding affinity to this sequence (Figure 5b). To further verify the specificity of the binding motifs, we generated a series of mutated version of the probe (P) (Supplementary Figure S9A) and performed electrophoretic mobility shift assay. We found that mutation of either CTTG and/or CAAG core sequences reduced the binding significantly (Supplementary Figure S9B). In addition, we found that the DNA sequences rather than the inverted repeated signature is important for the binding (Supplementary Figures S9A and B). We also found that the linker length between CTTG and CAAG is also very critical for NAC050 and NAC052 binding (Supplementary Figures S9A and B). These findings suggest that NAC050 and NAC052 could directly bind the ‘CTTGNNNNNCAAG’ motif, which may be necessary to facilitate the recruitment of JMJ14 to its targets.

#### *NAC052 and NAC050 are involved in RNA silencing*

To further test whether NAC050 and NAC052 function with JMJ14, we crossed *nac050* and *nac052* with JAP lines. In parallel with the genetic relation between *jmj14* and JAP lines, mutation of *NAC052*, and to a lesser extent *NAC050*, indeed suppress the photobleaching phenotype of JAP (Figure 6a). Consistent with reduced photobleaching, *JAPnac052-2* and *JAPnac050-2* also displayed lower levels of *PDS* small



**Figure 5** Identification of the DNA motif bound by NAC050 and NAC052. (a) A palindromic DNA motif CTTGNNNNNCAAG identified in the binding regions of NAC050, NAC052, and JMJ14. (b) The binding assay using different amount of GST (glutathione S-transferase)-fused recombinant proteins showed significant binding affinity of NAC050 and NAC052 but not FYR (FYRN+FYRC) domain of JMJ14 (Jumonji C (JmjC) domain-containing protein 14) to the probe.



**Figure 6** Photobleaching suppression of *nac052* and to a less extent *nac050* in transgenic lines. **(a)** The leaf photobleaching phenotype of *nac050* and *nac052* compared with *jmj14-1* under JAP background. The upper panel shows the 12-day plants and the bottom panel shows the 3-week plants. **(b)** Detection of *PDS* small interfering RNAs (siRNAs) in JAP lines under different mutant background. **(c)** The JAP/*nac052-2* homozygous plants were crossed with *nac052-2* and NAC052 complementary lines. The leaf phenotypes of F1 plants were observed. The JAP<sup>+/+</sup> plants under Columbia (Col) background were used as controls. NAC052-HA refers to the *P<sub>NAC052</sub>::NAC052-HA nac052-2* transgenic lines. **(d)** The model for JMJ14 targeting mediated by NAC050/052.

interfering RNAs (Figure 6b). All these phenotypes are similar to those seen in *jmj14* mutant. To further confirm this observation, we then crossed two *P<sub>NAC052</sub>::NAC052-HA nac052-2* lines into JAP*nac052-2* and found that NAC052-HA could fully rescue the phenotype of JAP*nac052-2* (Figure 6c). These results confirmed that the loss of photobleaching phenotype in JAP*nac052* was caused by the mutation of *NAC052*. Thus, we concluded that NAC050 and NAC052 cooperate with JMJ14 to positively regulate RNA silencing.

## Discussion

JmjC-domain-containing proteins have been found to have important roles in growth and development in both animals and plants. However, how these proteins are recruited to their target genes is poorly understood. In animals, two modes were reported for chromatin-associated protein targeting: namely

mediated by a specific domain of such protein or coordinated with other chromatin-associated proteins. For example, two H3K36 histone demethylases, lysine demethylase 2A (KDM2A) and KDM2B [33, 34], are recruited to non-methylated CpG islands by their zinc-finger CxxC (ZF-CxxC) domain [35–38]. Jarid1a, one of the H3K4 demethylases in KDM5 subgroup, is recruited to different target genes by associating with polycomb repressive complex 2, a histone H3K27 methyltransferase containing complex [39, 40], or G9a, a histone H3K9 methyltransferase [41, 42].

JMJ14 belongs to KDM5/JARID1 subfamily and possesses H3K4 demethylase activity [9–11, 17]. It has been shown to be involved in transgene silencing and flowering time regulation [9–12, 16]. In this work, we demonstrate that the FYRN and FYRC domains of JMJ14 have crucial roles in transcriptional gene silencing and flowering time regulation. Deletion of FYRN and FYRC domains causes mistargeting



of JMJ14 over several hundred of genes and H3K4me3 hypermethylation mimicking *jmj14* mutants.

We also reveal that JMJ14 interacts with two putative transcription factors, NAC050 and NAC052, through its FYRN and FYRC domains. Although FYRN and FYRC were identified as two domains separately, our results showed these two domains function only when they are both present. This is consistent with the previous structural evidence that FYRN and FYRC form a single domain with an  $\alpha+\beta$  fold [23]. By genome-wide analysis, we identified a significant palindromic DNA motif CTTGNNN NNCAAG in JMJ14, NAC050, and NAC052 binding regions. We prove that NAC050 and NAC052 but not FYRN and FYRC domains of JMJ14 bind to this DNA motif directly. Therefore, we propose that NAC050 and NAC052 bind to their targets by recognizing the palindromic DNA motifs and recruit JMJ14 through the interaction with FYRN and FYRC domains (Figure 6d). Besides, the overlapped binding peaks of JMJ14 and NAC050/052, we also calculated the density of the CTTGNNNNNCAAG motif in the binding regions of JMJ14 alone and the common binding regions of NAC050 and NAC052. According to our analysis, the motif is still dominantly enriched in the non-overlapped regions, which is present in  $\sim 85\%$  of the non-overlapped JMJ14 binding regions and  $73\%$  of non-overlapped NAC050/052 binding regions (Supplementary Figure S9C). This result may suggest that NAC050/052 first formed a stable interaction by motif matching before recruitment of JMJ14 to the targeted genome, and then left after JMJ14 is ready for action.

Mutations in *NAC050* and *NAC052* derepress transgene silencing similar with that of *jmj14*, which further prove that NAC050/052 act together with JMJ14 *in vivo*. We also noticed that JAP*nac052* showed more reduced photobleaching phenotype than that of JAP*nac050*. This is consistent with more reduction of *PDS* small interfering RNA accumulation in JAP*nac052* than that in JAP*nac050*, which suggest that NAC052 has predominant role and such level of reduction is sufficient to suppress the photobleaching phenotype. Therefore, we uncovered a novel targeting mechanism that uncharacterized domains of JMJ14 associate with a pair of NAC transcription factors to promote transcriptional repression. Recently, it has been shown that a pair of transcription factors Pax3 and Pax9 bind to satellite repeats sequences and recruit H3K9 methyltransferase Suv39h to establish H3K9me3 and silence the heterochromatin in mouse [43], suggesting a conserved function of

transcription factors in silencing in both animals and plants.

In addition, Wang *et al.* [44] showed that JMJ14 acts together with polycomb repressive complex 1-like complex that is responsible for the maintenance of H3K27me3. In *Arabidopsis*, H3K4me3 and H3K27me3 are known to be mutually exclusive. Our finding that JMJ14 recruitment by NAC050 and NAC052 to specific sequences leads to removal of H3K4me3 suggests that JMJ14 is the key regulator in establishing H3K27me3 repressive domain in some of the Polycomb targets. It is interesting to dissect the role of JMJ14 in regulating Polycomb-mediated gene silencing at genome-scale in the future.

## Materials and Methods

### Plant materials

All *Arabidopsis* plants were grown at 23 °C under long day condition (16 h light, 8 h dark). JAP transgenic lines were obtained from Dr David Baulcombe. The mutants alleles used are as follows: *jmj14-1* (SALK\_135712), *nac050-1* (SAIL\_841\_F01), *nac050-2* (SALK\_026244C), *nac052-1* (SALK\_056304C), and *nac052-2* (WiscDsLoxHs027\_03D).

### Yeast two-hybrid assays

For yeast two-hybrid screening, full-length CDS of *JMJ14* was cloned into pGBKT7 vector using primers CX5024 and CX5025. A mixture of RNAs from 10-day-old seedlings and flowers of *Arabidopsis* were used to construct the cDNA library. Library construction and screening were performed with Matchmaker Library Construction & Screening Kits (Clontech 630445, Japan). The full-length CDS of *NAC050* and *NAC052* were cloned into pGADT7 vector using primers CX8980 and CX8981 (for *NAC050*) or CX8984 and CX8985 (for *NAC052*). The truncated *JMJ14* with different domain deletions were cloned using a QuickChange kit (Stratagene, Santa Clara, CA, USA) with primers CX8988 and CX8989 (for *JMJ14ΔJmjN*), CX8990 and CX8991 (for *JMJ14ΔJmjC*), CX8992 and CX8993 (for *JMJ14ΔZnF*), or CX8994 and CX8995 (for *JMJ14ΔFYR*). The different domains of *JMJ14* were cloned using primers HX2032 and HX2033 (for *JmjN*), HX2034 and HX2035 (for *JmjC*), HX2036 and HX2037 (for *ZnF*), HX2038 and HX2041 (for *FYR*), HX2038 and HX2039 (for FYRN), and HX2040 and HX2041 (for FYRC). The bait and prey vectors were cotransformed into yeast AH109 strain following the manufacturer's handbook (Clontech Yeast Protocols Handbook). The cotransformed yeast clones were first cultured in SD/-Leu/-Trp liquid medium. The same amount of yeast in liquid medium was plated on SD/-Leu/-Trp and SD/-Ade/-His/-Leu/-Trp medium containing 20  $\mu\text{g ml}^{-1}$  X- $\alpha$ -Gal respectively. Primer sequences can be found in Supplementary Table S2.

### GST pull-down

The full-length CDS of *NAC050* and *NAC052* were amplified with primers HX2046 and CX9579 (for *NAC050*) or HX2047 and CX9581 (for *NAC052*) and inserted into the expression

vector pMAL-C2-maltose binding protein (XF510). The FYR domain of JMJ14 was amplified with primers HX0406 and CX5012 and inserted into the expression vector pMAL-C2-GST (XF760). The fragments were inserted into pMAL-C2-maltose binding protein and pMAL-C2-GST by a ligation-independent cloning method as previously described [45]. All the recombinant proteins were expressed and purified from *Escherichia coli* strain BL21 RIL (BL21 CP, Stratagene). For pull-down assays, bait proteins were bound to glutathione beads and blocked overnight with 5% bovine serum albumin in pull-down buffer (50 mM Tris pH 7.5, 300 mM NaCl, and 0.1% CA-630). Then the complexes were incubated with target proteins for 45 min, washed eight times with pull-down buffer. Eluates were analyzed by western blotting using anti-His antibodies (Abmart M20001, Shanghai, China). Primer sequences can be found in Supplementary Table S2.

#### Co-immunoprecipitation assay

One gram of 12-day-old seedlings of *Arabidopsis* were ground to powder in liquid N<sub>2</sub> and suspended in 1 ml of PEN-140 buffer (140 mM NaCl, 2.7 mM KCl, 25 mM Na<sub>2</sub>HPO<sub>4</sub>, 1.5 mM KH<sub>2</sub>PO<sub>4</sub>, 0.01 mM EDTA, and 0.05% CA-630). The supernatant was incubated 2 h with 40 μl Dynabeads Protein G (Invitrogen 10004D, Waltham, MA, USA) conjugated with antibodies that could recognize both NAC050 and NAC052 (Supplementary Figure S10). The beads were washed five times with PEN-400 buffer (400 mM NaCl, 2.7 mM KCl, 25 mM Na<sub>2</sub>HPO<sub>4</sub>, 1.5 mM KH<sub>2</sub>PO<sub>4</sub>, 0.01 mM EDTA, and 0.05% CA-630) and then analyzed by western blotting using anti-HA antibodies (Sigma H6908, St Louis, MO, USA).

#### ChIP assay

ChIP assays were performed using 1 g of 12-day-old seedlings as previously described with minor modifications [10]. For anti-HA ChIP, chromatin was fragmented by micrococcal nuclease (NEB M0247S) in the micrococcal nuclease digestion buffer (50 mM Tris pH 8.0, 0.32 M sucrose, 4 mM MgCl<sub>2</sub>, and 1 mM CaCl<sub>2</sub>) instead of sonication before nuclei lysis. Antibodies used in ChIP were anti-H3K4me3 (Millipore 07-473, Darmstadt, Germany), anti-H3 (Abcam ab1791, Cambridge, MA, USA) and anti-HA (Sigma H6908). Primers for ChIP-qPCR can be found in Supplementary Table S2.

#### ChIP-seq analysis

ChIPed DNA was ligated with Illumina single-end genome sequencing adapters, size fractionated to obtain 300-bp fragments, PCR amplified and sequenced according to standard protocols (single-end 36 cycles). The reads were aligned to *Arabidopsis thaliana* genome build TAIR10 by Bowtie 2 [46] using default parameters with local alignment model. Duplicated reads and low-mapping quality reads were identified and removed with SAMtools [47]. Enriched intervals were identified by MACS [48] with default parameters. Density maps of reads for visualization were calculated by SAMtools as well as programs in University of California, Santa Cruz, genome browser based on counts of the 200-bp extension of sequencing reads in the 3' direction (as described previously in Ernst *et al.* [49]) after total reads normalization.

#### Electrophoretic mobility shift assay

The full-length CDS of NAC050 and NAC052 were amplified with primers HX2046 and CX9579 (for NAC050) or HX2047 and CX9581 (for NAC052). The FYR domain of JMJ14 was amplified with primers HX0406 and CX5012. The fragments were inserted into the expression vector pMAL-C2-GST (XF760) by a ligation-independent-cloning method as previously described [45]. All the recombinant proteins were expressed and purified from *E. coli* strain BL21 RIL (BL21 CP, Stratagene). Ten or 50 ng of purified proteins were incubated with 0.3 pM of each <sup>32</sup>P-labeled double-stranded DNA oligos in binding buffer (25 mM Tris-HCl, 100 mM NaCl, 2.5 mM MgCl<sub>2</sub>, 0.1% CA-630, 10% glycerol, and 0.5 mM Dithiothreitol) on ice for 1 h. The binding reaction mixture was loaded onto a 6% non-denaturing polyacrylamide gel at 80 V for 1 h. The DNA was detected by the radioactive signal. Primer sequences can be found in Supplementary Table S2. The probes sequences are listed in Supplementary Figure S9A.

#### Transgene

The full-length *JMJ14* genomic fragment with 1.5 kb of promoter was amplified using primers CX5033 and CX3599. The truncated *JMJ14* without FYRN and FYRC (FYR) domains was amplified using primers CX5033 and CX7385. The fragments were then cloned into pENTR/D-TOPO (Invitrogen) and introduced by LR reaction into pEarleyGate301 (pEG301) vector with a C-terminal HA tag (*P<sub>JMJ14</sub>::JMJ14-HA* and *P<sub>JMJ14</sub>::JMJ14<sub>ΔFYR</sub>-HA*) [50]. The full-length CDS of *NAC050* and *NAC052* were introduced into pEarleyGate102 (pEG102) vector with a C-terminal CFP-HA tag (*P<sub>35S</sub>::NAC050-CFP-HA* and *P<sub>35S</sub>::NAC052-CFP-HA*) using primers CX8188 and CX8189 (for *NAC050*) or CX8191 and CX8194 (for *NAC052*). *P<sub>35S</sub>::JMJ14-YFP-HA* was constructed as previously described [10]. For GUS staining vectors, ~0.8 kb of 3' untranslated regions of *JMJ14*, *NAC050*, and *NAC052* were cloned and introduced into pCambia1300-GFPUS (XF1764) after *NcoI* digestion. The full-length genomic regions with native promoter of *JMJ14*, *NAC050*, and *NAC052* were then amplified and constructed into pCambia1300-GFPUS-UTR. The following primers are used: CX9272 and CX9273 for *NAC050* genomic region, CX9274 and CX9275 for *NAC050* 3'UTR, CX9276 and CX9277 for *NAC052* genomic region, CX9278 and CX9279 for *NAC052* 3'UTR, CX9280 and CX9281 for *JMJ14* genomic region, and CX9282 and CX9283 for *JMJ14* 3'UTR. For *NAC050* and *NAC052* complementary vectors, ~0.8 kb of 3'UTR of *NAC050* and *NAC052* were cloned and introduced into pCambia1300-HA (XF0791) after *PstI* digestion. The full-length genomic sequences with native promoter of *NAC050* and *NAC052* were then amplified and constructed into pCambia1300-HA-UTR by *EcoRI* and *BamHI* double digestion. The primers used are as follows: CX9272 and CX9273 for *NAC050* genomic region, HX2078 and HX2079 for *NAC050* 3'UTR, CX9276 and CX9277 for *NAC052* genomic region, and HX2080 and HX2081 for *NAC052* 3'UTR. The constructs were introduced into *Agrobacterium tumefaciens* cells (strain EHA105) and then stably transformed into *Arabidopsis* using the floral dip method [51]. Primer sequences can be found in Supplementary Table S2.

### Flowering time assessment

All plants were grown in soil side by side at 23 °C under long day (16 h light and 8 h dark) conditions. Flowering time was assessed by counting the number of rosette and cauline leaves when the plants flowered. At least 15 plants were counted for each line.

### Affinity purification and mass spectrometry

Purification of JM14-3 × Flag was performed as described in Law *et al.* [52] with minor modifications. In brief, 15 g of flower tissue collected from transgenic T4 plants, or from Col plants as a negative control, were ground with a mortar and pestle in liquid nitrogen and suspended in 45 ml of lysis buffer (50 mM Tris (pH 7.6), 150 mM NaCl, 5 mM MgCl<sub>2</sub>, 10% glycerol, 0.1% NP-40, 0.5 mM DTT, 1 mg ml<sup>-1</sup> pepstatin, 1 mM Phenylmethylsulfonyl fluoride, and 1 protease inhibitor cocktail tablet (Roche, 14696200, Basel, Switzerland)). Following centrifugation to remove insoluble plant debris, lysate was incubated with 125 µl of Dynabeads (M-270 Epoxy, Invitrogen, 143.01) that had been conjugated with Flag antibody (Sigma F 3165) according to the manufacturer's instructions. After incubation at 4 °C with rotation for 2.5 h, the Flag beads were washed seven times with lysis buffer. Proteins were then eluted from the Flag beads by competition with 150 µl of 100 µg ml<sup>-1</sup> of 3 × Flag peptide (Sigma, F 4799) five times at room temperature. Mass spectrometry and analysis was performed exactly as described in Law *et al.* [52].

### RNA gel blot and transcription level analysis

Total RNA was extracted from 12-day-old seedlings using TRNzol reagent (Tiangen, Beijing, China). RNAs (15 µg per lane) were separated in an agarose gel containing 1% formaldehyde, blotted onto Hybond N<sup>+</sup> membrane (GE Healthcare RPN119B, Little Chalfont, UK), and probed with *JMJ14* cDNA, which was amplified using primers CX5024 and CX5025. *PDS* small interfering RNA gel blot was performed exactly as previous report [53]. Reverse transcription reactions were performed using TransScript II First-Strand cDNA Synthesis SuperMIX (TransGen AH301-02, Beijing, China). qPCR was performed using a CFX96 Real-time PCR instrument (Bio-Rad, Hercules, CA, USA) with RealSYBR Mixture (CWBIO and CW0760). *UBC (At5g25760)* was used as the internal control of Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Primers for RT-qPCR or RT-PCR are listed in Supplementary Table S2.

### Histochemical GUS staining

Histochemical GUS staining was performed according to the standard procedure [54]. Twelve-day-old seedlings of pCambia1300-GFP-GUS transgenic lines were incubated in GUS staining buffer (50 mM sodium phosphate buffer (pH 7.2), 10 mM EDTA, 0.2% Triton X-100, and 2 mM potassium ferrocyanide/ferricyanide) with 2 mM X-glucuronide at 37 °C. After staining, the samples were cleared and stored in 70% ethanol for microscopic analysis.

### In vivo histone demethylation assay

The truncated *JMJ14* without FYRN and FYRC (FYR) domains was amplified using primers CX3598 and CX7385. The fragments were then cloned into pENTR/D-TOPO (Invitrogen) and introduced by LR reaction into *pEarley-Gate101 (pEG101)* vector with a C-terminal YFP-HA tag (*P<sub>35S</sub>::JM14ΔFYR-YFP-HA*). Tobacco (*Nicotiana benthamiana*) leaves was infiltrated with *A. tumefaciens* EHA105 strains containing *P<sub>35S</sub>::JM14ΔFYR-YFP-HA* vector to overexpress JM14ΔFYR-YFP-HA. The demethylation assay was carried out as previously described [10]. Immunolabeling was performed using histone methylation-specific antibodies (H3K4me3: Millipore 07-473, 1:100; H3K4me2: Millipore 07-030, 1:500; and H3K4me1: Millipore 07-436, 1:100). At least 25 pairs of transfected or non-transfected nuclei were observed in the same field of view. Quantification was done using ImageJ (National Institutes of Health, Bethesda, MD, USA) software by comparing Alexa Fluor 555-staining density of nuclei overexpressing JM14ΔFYR-YFP-HA to that of the local neighboring wild-type nuclei.

### Data deposition

The data discussed in this publication have been deposited in NCBI's Gene Expression Omnibus [55] and are accessible through GEO Series accession number GSE60084 (<http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE60084>).

### Acknowledgements

We thank the Arabidopsis Biological Resource Center for providing T-DNA insertion lines, Dr David Baulcombe for JAP plants. This work was supported by the National Basic Research Program of China (grant nos. 2011CB915401 to X Cao and 2013CB967300 to Xia Cui), the National Natural Science Foundation of China (grant nos. 31271363 to X Cui and 31330020 to X Cao), Genetically Modified Breeding Major Projects (grant no. 2014ZX08010-002 to X Cao), the State Key Laboratory of Plant Genomics, and the post-doctoral fellowship to (BZ and Xiekui Cui). Work in the Jacobsen laboratory was supported by National Institutes of Health grant GM60398. SEJ is an investigator of the Howard Hughes Medical Institute.

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