

Identification of a natural product-like STAT3 dimerization inhibitor by structure-based virtual screening

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STAT3 regulates a variety of genes involved with cell proliferation, differentiation, apoptosis, angiogenesis, metastasis, inflammation, and immunity. The purpose of this study was to apply molecular docking techniques to identify STAT3 inhibitors from a database of over 90 000 natural product and natural product-like compounds. The virtual screening campaign furnished 14 hit compounds, from which compound 1 emerged as a top candidate. Compound 1 inhibited STAT3 DNA-binding activity *in vitro* and attenuated STAT3-directed transcription *in cellulo* with selectivity over STAT1 and with comparable potency to the well-known STAT3 inhibitor S3I-201. Furthermore, compound 1 inhibited STAT3 dimerization and decreased STAT3 phosphorylation in cells without affecting STAT1 dimerization and phosphorylation. Compound 1 also exhibited selective anti-proliferative activity against cancer cells over normal cells *in vitro*. Molecular docking analysis suggested that compound 1 might putatively function as an inhibitor of STAT3 dimerization by binding to the SH2 domain. This study also validates the use of *in silico* techniques to identify inhibitors of protein–protein interactions, which are typically considered difficult to target with small molecules.

Cell Death and Disease (2014) 5, e1293; doi:10.1038/cddis.2014.250; published online 13 June 2014

Signal transducer and activator of transcription (STAT) proteins are a family of transcription factors that mediate gene expression in response to cytokines and growth factors.¹ STAT3 regulates a variety of genes involved in cell proliferation, differentiation, apoptosis, angiogenesis, metastasis, inflammation, and immunity.^{2–4} Dysregulated STAT3 activity has been implicated in the development of a variety of solid and hematological tumors, including leukemia, lymphomas, and head and neck cancer.^{5–7} Additionally, elevated STAT3 levels have been associated with poor prognosis of certain cancers.⁸ The central role of aberrant STAT3 signaling in tumorigenesis has rendered STAT3, and, to a lesser extent, STAT5, as an attractive target in anticancer therapy,^{9,10} in addition to autoimmune and inflammatory applications.^{11–14}

The structure of STAT3 is a characteristic of the STAT family, and includes an N-terminal coiled–coiled domain, a C-terminal transactivation domain, a DNA-binding domain, a Src homology 2 (SH2) domain. The N-terminal coiled–coiled domain is involved in protein–protein interactions for the formation of multiple types of dimer complexes,¹⁵ and also contains a lysine (Lys140) residue for methylation by histone methyl transferase SET9, which is a negative regulatory event.¹⁶ On the other hand, the C terminus contains a conserved tyrosine (Tyr705) residue that is essential for

STAT3 activation.¹⁵ In the classical STAT3 activation pathway, growth factors and cytokines induce the tyrosine kinase activities of JAK or Src receptors, resulting in specific tyrosine phosphorylation of receptor chains.¹⁷ Latent STAT3 is recruited to the activated receptor via phosphotyrosine-SH2 recognition, and is phosphorylated at Tyr705 by JAK/Src. STAT3 monomers dimerize through reciprocal phosphotyrosine-SH2 interactions, and activated STAT3 dimers translocate into the nucleus where they bind to DNA response elements in the promoters of targeted genes, leading to gene transcription.

In normal cells, the activation of STAT3 is tightly regulated by a number of signaling mechanisms.⁸ Some protein tyrosine phosphatases (PTPs), such as the ubiquitously expressed SHP-2, contain SH2 domains that can potentially recognize the phosphotyrosine elements of any of the members of the STAT3 signaling pathway.¹⁸ In addition, the constitutively expressed protein inhibitor of activated STAT3 binds specifically to STAT3 and blocks its ability to bind to DNA and activate gene transcription.¹⁹ Finally, suppressors of cytokine signaling (SOCS) possess SH2 domains that recognize phosphotyrosine elements of JAKs, cytokine receptor chains and gp130.¹⁸ The versatile SOCS3, whose transcription is activated by STAT3 in a negative feedback mechanism, can

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Abbreviations: STAT, signal transducer and activator of transcription; SH2, Src homology 2; PTPs, protein tyrosine phosphatases; PIAS3, protein inhibitor of activated STAT3; SOCS, suppressors of cytokine signaling; EGFR, human epidermal growth factor receptor; EGF, human epidermal growth factor; IFN- α , interferon alpha protein; IFN- γ , interferon gamma protein; IL-6, interleukin 6; DMSO, dimethyl sulfoxide; VSL, virtual library screening; BPMP, biased probability Monte Carlo

Received 12.1.14; revised 03.5.14; accepted 06.5.14; Edited by A Stephanou

block STAT3 signaling by the direct inhibition of JAK activity, by competing with STAT3 for phosphotyrosine residues on the receptor chains, or by binding to signaling proteins and triggering their proteasomal degradation.²⁰

In contrast, constitutive STAT3 activation is a hallmark of a variety of human cancers. Though a large amount of evidence has been accumulated linking the pharmacological or genetic control of constitutively active STAT3 with tumor phenotype and progression *in vivo*,^{21,22} the precise mechanism of STAT3-induced tumor formation has yet to be completely established. Increased expression of human epidermal growth factor receptor (EGFR), Src or Src family of tyrosine kinases, or mutations in JAK proteins leading to hyperactive activity are factors that have been proposed to account for constitutively phosphorylated STAT3.^{23,24} The levels of STAT3-activating ligands such as TGF α and IL-6 were also increased in the serum or tumors of patients with various types of human cancers.⁶ Alternatively, the disruption of epigenetic control of STAT3 regulators or the reduced expression of STAT3 antagonists, such as PTP, PIAS, or SOCS proteins may also promote excessive STAT3 activity, enhancing tumor growth.³

Molecules that can inhibit STAT3 activity represent a potential avenue for chemotherapeutic intervention.²⁵ Popular approaches include oligopeptides/peptidomimetics²⁶ or G-quadruplex nucleic acids^{27,28} that target SH2 domains and disrupt STAT3 dimerization, or 'decoy' oligonucleotides that sequester active STAT3 and block the authentic STAT3-DNA interaction.²⁹ More recently, there has been an increased interest in the development of small-molecule inhibitors of STAT3 activity. The amidosalicylic acid S3I-201 inhibits STAT3 dimerization and blocks STAT3 DNA-binding activity and STAT3-dependent transcription *in vitro*.²² Alternatively, platinum(IV) compounds have shown potent activity against STAT3 DNA-binding activity, STAT3-driven gene expression, and cancer growth, via binding to the STAT3 DNA-binding domain.³⁰ Recently our group discovered the first example of a substitutionally-inert, Group 9 organometallic compound as a direct inhibitor of STAT3 dimerization with potent anti-tumor activities in an *in vivo* mouse xenograft model of melanoma.³¹ However, despite these promising studies, no compound discovered as a STAT3 inhibitor *a priori* has yet been approved for clinical use.

One challenge that has to be overcome by STAT3 inhibitors is the high degree of homology between the structures of STAT3 and STAT1, particularly in their SH2 domains. Bluysen and co-workers³² have recently used comparative *in silico* docking to study the binding specificity of STAT inhibitors statin and fludarabine, and have concluded that ligands targeting only the highly conserved phosphotyrosine binding pocket of the SH2 domain (i.e. statin) will lack selectivity towards STATs, as STAT1 and STAT3 have identical active residues at this site. A number of compounds thought to be primarily STAT3 inhibitors, such as resveratrol,³³ have also been found to inhibit STAT1 activity. Although STAT3 and STAT1 are highly similar in terms of both protein and target DNA sequences, they have different physiological effects. Deficiency in STAT1 leads to attenuation of interferon (IFN) responsiveness, increasing the susceptibility of mice to viral or

bacterial infections. Moreover, activated STAT1 may behave as a tumor suppressor,³⁴ though further studies are needed to elucidate this relationship. This suggests that selectivity for STAT3 over STAT1 may be a desirable characteristic for the development of novel anticancer therapeutics.

Structure-based virtual screening has emerged as an efficient strategy in drug discovery complementing conventional high-throughput techniques.^{35–39} By weeding out inactive non-binders *in silico*, the numbers of compounds to be synthesized and tested *in vitro* can be dramatically reduced. Meanwhile, natural products have historically represented an importance source of chemical scaffolds and bioactive substructures for the medicinal chemist.⁴⁰ A number of natural products including curcumin, cucurbitacin, honokiol, guggulsterone, resveratrol, berbamine, and flavopiridol have been reported to exhibit STAT3 inhibitory activity.¹⁴ Encouraged by these ideas, we sought to apply molecular docking techniques to identify STAT3 inhibitors from a database of natural product and natural product-like compounds, and to evaluate the STAT3 inhibitory activity of the hit compounds *in vitro*.

Results

Virtual screening campaign and hit validation. The X-ray structure of the STAT3 homodimer bound to DNA (PDB: 1BG1)⁴¹ was used as the molecular model for our investigations. For the molecular docking, DNA was removed and only one of the monomers was used. The search area for screening was restricted to the SH2 domain of the STAT3 monomer. Over 90 000 compounds from a chemical library of natural products and natural product-like molecules were screened *in silico*. Fourteen compounds (Figure 1) were screened in a preliminary ELISA to evaluate their ability to inhibit STAT3 DNA-binding activity *in vitro* (Figure 2). From these results, the benzofuran derivative compound **1** emerged as a top candidate.

Inhibition of STAT3 DNA-binding. A dose-response experiment was performed to determine the potency of compound **1** at inhibiting the DNA-binding activity of STAT3. In the assay, HepG2 extracts containing EGF-activated STAT3 were incubated with increasing concentrations of compound **1** or S3I-201. Encouragingly, a dose-dependent reduction in the DNA-binding activity of STAT3 was observed in the presence of compound **1**. The potency of compound **1** ($IC_{50} = ca. 15 \mu M$) was comparable to that of the positive control compound S3I-201 ($IC_{50} = ca. 10 \mu M$) determined under the same conditions (Figure 3). We also performed a parallel experiment with the homologous STAT1 protein to evaluate the selectivity of compound **1**. No inhibition in STAT1 DNA-binding was observed when COS-7 nuclear extracts containing IFN- γ -induced activated STAT1 were incubated in the presence of 30 μM of compound **1** and S3I-201 (Supplementary Figure S2). The selectivity of compound **1** (and S3I-201) for STAT3 over STAT1 was confirmed in HeLa nuclear extracts stimulated with IFN- α , which phosphorylates STAT3 and STAT1 with comparable efficiency (Supplementary Figure S3). These results suggest that compound **1** is able to block the binding of activated STAT3 to its consensus DNA sequence in isolated

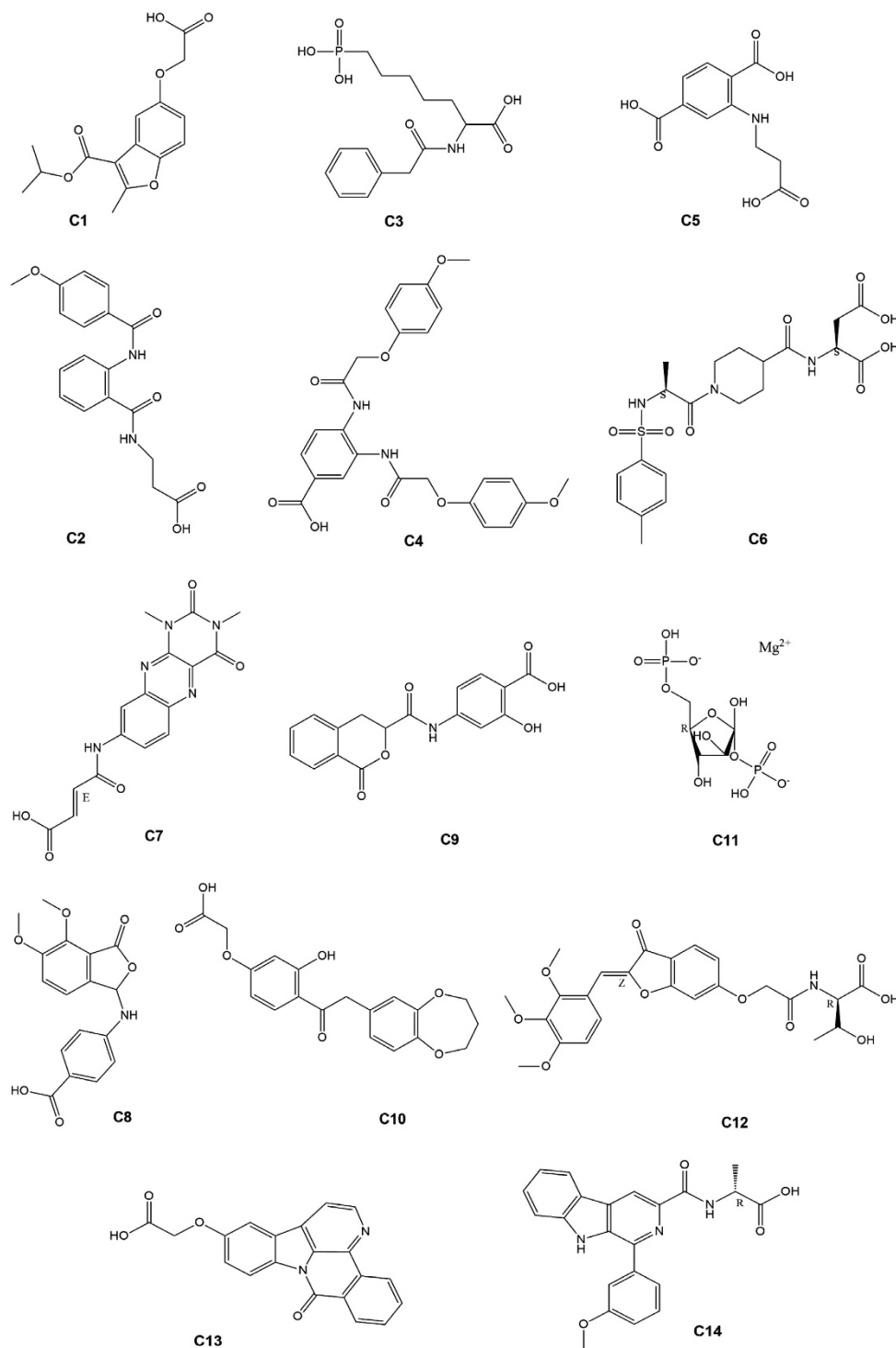


Figure 1 Structures of compounds 1–14 identified in the high-throughput virtual screening chosen for biological validation

nuclear extracts, and is selective for STAT3 over the closely related protein STAT1.

Molecular docking analysis. We performed molecular modeling of compound 1 with the STAT3 SH2 domain in order to further understand the mode of binding (Figure 4). Hydrogen bonds were predicted between the carboxylate

functionality of compound 1 with Ser611 (1.99 Å) and Glu612 (2.32 Å), respectively, whereas the ether moiety of compound 1 formed a hydrogen bond to Arg609 (2.06 Å). According to the model, the benzofuran and isopropyl ester moieties do not significantly interact with the protein, with the closest distance between the benzofuran and Lys591 being 3.11 Å and the closest distance between the isopropyl ester

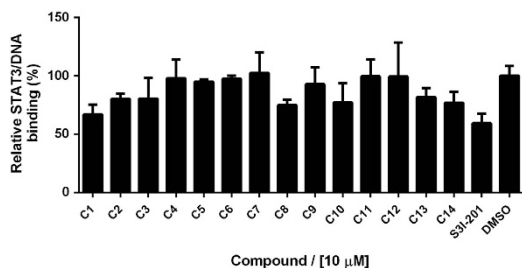


Figure 2 STAT3 DNA-binding inhibitory activity of compounds 1–14 as determined by ELISA. Microtitre plates coated with the STAT3 consensus sequence were incubated with nuclear extracts containing activated STAT3 and 10 μM of compounds. STAT3 binding was detected using anti-STAT3 primary antibody and horseradish peroxidase-conjugated secondary antibody. Results are representative of three independent experiments

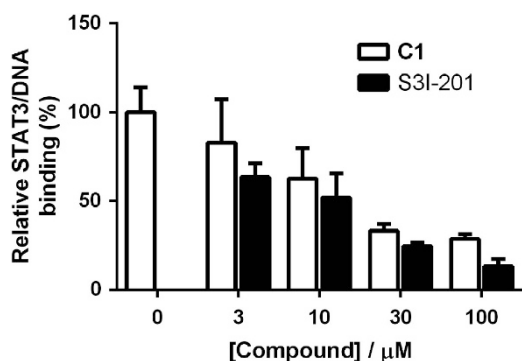


Figure 3 Compound 1 inhibits the DNA-binding activity of STAT3 in a dose-dependent manner as measured by ELISA. Microtitre plates coated with the STAT3 consensus sequence were incubated with HepG2 nuclear extracts containing EGF-activated STAT3 and 1 or S31-201 at the indicated concentrations. STAT3 binding was detected using anti-STAT3 primary antibody and horseradish peroxidase-conjugated secondary antibody. Results are representative of three independent experiments. Error bars represent the S.D. of triplicate results. Estimated IC_{50} values: compound 1: 15 μM , S31-201: 10 μM

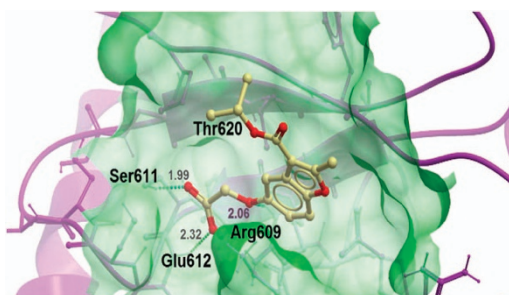


Figure 4 Low-energy-binding conformations of compound 1 bound to the STAT3 β homodimer generated by virtual ligand docking. Compound 1 is depicted as a ball-and-stick model showing carbon (yellow) and oxygen (red) atoms. Hydrogen bonds are depicted as dotted lines. The binding pocket of the STAT3 β is represented as a translucent green surface

and Thr620 being 5.83 Å. The binding score of -31.3 for compound 1 reflects the strong binding interaction between compound 1 and the STAT3 SH2 domain.

For further validation of the selectivity of compound 1, we performed molecular docking of compound 1 (Supplementary Figure S4) and S31-201 (Supplementary Figure S5) with STAT1 and STAT5 proteins. Surprisingly, the

lowest-energy-binding poses of compound 1 in the SH2 domain of STAT1 and STAT5 were very different to that with STAT3 (Supplementary Figure S4). Compound 1 was predicted to form hydrogen-bonding interactions with Glu612 and Lys591 of STAT1 via its carboxylate group, but not with any of the residues of the STAT5 SH2 domain. The unfavorable binding scores of compound 1 bound to STAT1 (-14.24) and STAT5 (-15.1) are consistent with the results of the STAT3 DNA-binding experiment described above. Similarly, the binding modes of S31-201 to STAT1, STAT3 and STAT5 were also different to each other (Supplementary Figure S5), which could possibly also account for the STAT3-specificity of this compound.

We also performed docking of a carboxylate derivative of compound 1 that could result from the cleavage of the isopropyl ester group by intracellular enzymes (Supplementary Figure S6). The lowest-energy-binding pose of the cleaved compound with STAT3 was almost identical to that exhibited by compound 1, though a less favorable binding score of -27.19 was obtained. Furthermore, several low-scoring molecules identified from the virtual screening campaign exhibited no STAT3 DNA-binding inhibitory activity, and were used as negative controls (Supplementary Figure S7). The docking scores of compounds 1–14 against STAT1, STAT3 and STAT5 are summarized in Supplementary Table S1. Additionally, the lowest-energy-binding poses of compounds 1–14 within the STAT3 SH2 domain are presented in Supplementary Table S2.

Inhibition of STAT3-driven transcriptional activity in cells. Given that compound 1 was able to block STAT3 DNA-binding activity in a cell-free system, we next investigated the ability of compound 1 to antagonize STAT3-driven transcriptional activity in living cells. Transfected HeLa cells were incubated with compound 1 or S31-201 and stimulated with EGF for 6 h. Compound 1 attenuated STAT3-directed transcription in a dose-dependent fashion ($\text{IC}_{50} = ca. 30 \mu\text{M}$), as revealed by a reduction in the ratio of firefly to *Renilla* luciferase activity (Figure 5a). A comparative assay was performed using IFN- α -stimulated HeLa cells transfected with plasmids bearing the STAT3 or STAT1 response element. Compound 1 selectively inhibited STAT3-directed transcriptional activity, while having negligible effect on STAT1-driven expression (Supplementary Figure S8). Notably, the potency of compound 1 was comparable to that of S31-201, which also showed selectivity for STAT3 over STAT1 under the same conditions. These results indicated that compound 1 could specifically inhibit STAT3-directed transcription in living cells, presumably due, at least in part, to the inhibition of STAT3 dimerization in the cytosol.

Inhibition of STAT3 dimerization. To further investigate the inhibition of STAT3 dimerization by compound 1 *in cellulo*, a co-immunoprecipitation assay was performed. HEK293T cells co-expressing STAT3-Flag and STAT3-GFP were treated with the indicated concentrations of compound 1 and S31-201 in a 6-well plate for 6 h. Protein lysates were purified by anti-Flag magnetic beads, eluted with low pH buffer and analyzed by western blotting with anti-Flag and anti-GFP antibodies. In the untreated cells, blotting

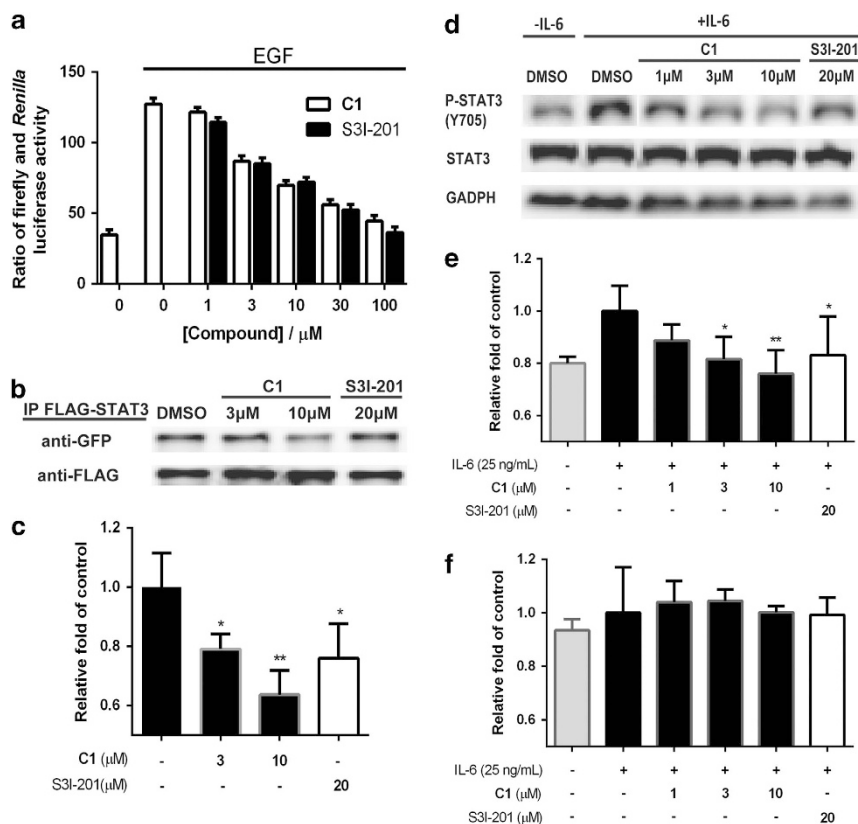


Figure 5 Effect of compound 1 on STAT3-driven transcription activity, STAT3 dimerization and STAT3 phosphorylation. (a) Compound 1 inhibits STAT3-driven transcription activity in EGF-stimulated HeLa cells as measured using a luciferase reporter assay. Estimated IC_{50} values: compound 1: 30 μM , S3I-201: 30 μM . (b) Compound 1 inhibits the dimerization of STAT3 in HEK293T cells. (c) Quantification of STAT3 dimerization inhibition by densitometry analysis. (d) Compound 1 inhibits STAT3 Y705 phosphorylation but not total STAT3 content in HepG2 cells. (e) Quantification of STAT3 Y705 phosphorylation inhibition by densitometry analysis. (f) Densitometry analysis for the cell-based western blot shows no inhibition of compound 1 on total STAT3. Error bars represent the S.D. of triplicate results. * $P < 0.05$, ** $P < 0.001$

of the immunoprecipitates with anti-GFP antibody revealed the presence of the STAT-GFP protein in the protein lysates, indicating that STAT3-GFP and STAT3-Flag proteins were pulled down together (Figures 5b and c). However, the level of STAT3-GFP in the anti-Flag immunoprecipitates decreased as the concentration of compound 1 was increased. This result suggests that compound 1 disrupted STAT3-STAT3 dimerization in a dose-dependent manner *in cellulo*. To investigate the selectivity of compound 1 for STAT3 over STAT1 dimerization, HeLa cells were co-transfected with pRc/CMV-STAT3-Flag and pCMV6-AV-STAT3-GFP or pRc/CMV-STAT1-Flag and pcDNA-GFP-STAT1. The results showed that compound 1 selectively interfered with STAT3 dimerization but had no significant effect on STAT1 dimerization (Supplementary Figure S9).

Inhibition of phosphotyrosine-STAT3 levels. Phosphorylation of STAT3 at critical tyrosine residues is an essential requirement for STAT3 dimerization. Therefore, we investigated the effect of compound 1 on STAT3 tyrosine phosphorylation in cells. HepG2 cells were treated with the indicated concentrations of compound 1 for 6 h and stimulated with IL-6 for 30 min. Protein lysates were subjected to western blotting with anti-pY705-STAT3 antibody. The results revealed a dose-dependent decrease in IL-6-induced STAT3 phosphorylation as the concentration of compound 1 was increased

(Figures 5d–f). However, compound 1 had no significant effect on total STAT3 expression. In a comparative assay, HeLa cells were treated with compound 1 and S3I-201 for 5 h and then stimulated with IFN- α for another 1 h. Compound 1 reduced STAT3, but not STAT1, phosphorylation in a dose-dependent manner (Figure 6). Moreover, the expression of STAT3 and STAT1 was not significantly reduced by treatment with compound 1. These results indicate that compound 1 could selectively decrease STAT3 phosphorylation over STAT1 phosphorylation *in cellulo*.

In vitro cytotoxicity. The cytotoxicity of compound 1 was investigated in LO2, HepG2, RAW264.7, and Caco2 cell lines. Cells were exposed to compound 1 (1–100 μM) for 72 h, and cellular proliferation was assessed using the MTT assay. Compound 1 was relatively nontoxic towards the normal liver LO2 cell line ($\text{IC}_{50} > 100 \mu\text{M}$), but showed dose-dependent inhibition of cellular proliferation of HepG2, RAW264.7 and Caco2 cell lines ($\text{IC}_{50} = \text{ca. } 30 \mu\text{M}$) (Figure 7). Taken together, the data suggest that compound 1 displays marked selectivity for cancer cells over normal cells.

Discussion

Natural products offer a rich source of bioactive structural motifs for the development of new pharmaceuticals, and a

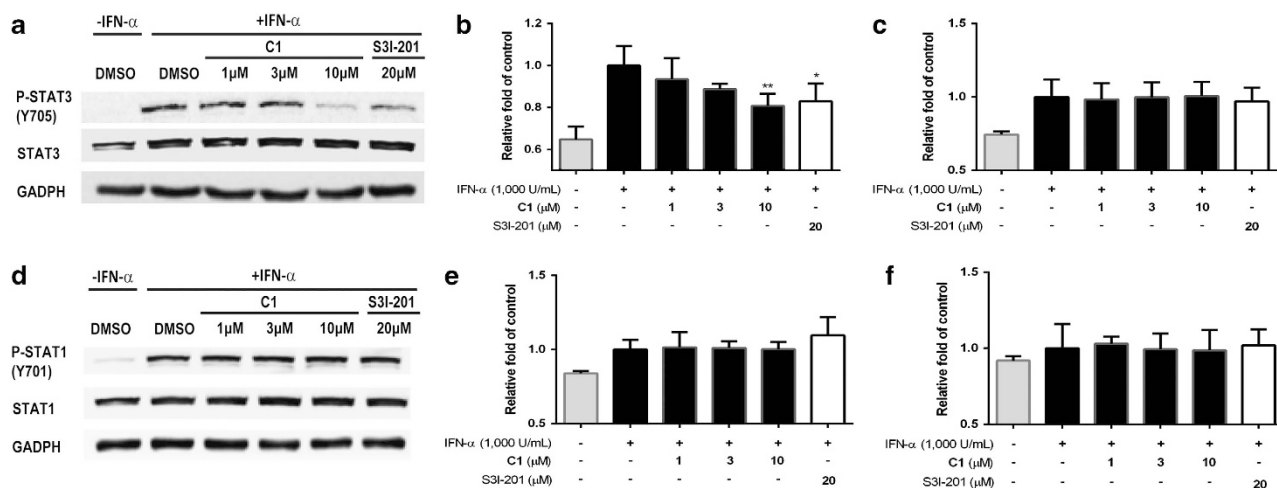


Figure 6 Effect of compound **1** on IFN- α -induced STAT3 Y705 phosphorylation and IFN- α -induced STAT1 Y701 phosphorylation in HeLa cells. (a) Compound **1** inhibits IFN- α -induced STAT3 Y705 phosphorylation but not IFN- α -induced total STAT3. (b) Quantification of STAT3 phosphorylation inhibition by densitometry analysis. (c) Quantification of total STAT3 expression by densitometry analysis. (d) Compound **1** has no effect on IFN- α -induced STAT1 Y701 phosphorylation and IFN- α -induced total STAT1. (e) Quantification of STAT1 phosphorylation inhibition by densitometry analysis. (f) Quantification of total STAT1 expression by densitometry analysis. Error bars represent the S.D. of triplicate results. * $P < 0.05$, ** $P < 0.001$

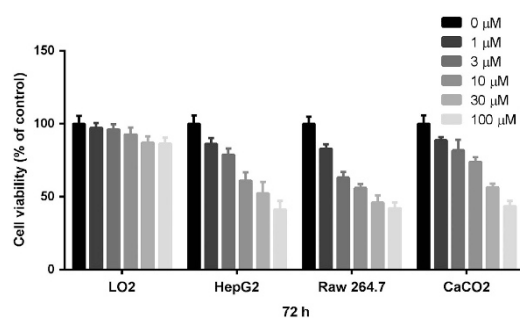


Figure 7 Cytotoxicity of compound **1** on cell viability as determined by the MTT assay. LO2, HepG2, RAW264.7, Caco2 were treated with the indicated concentrations of compound **1** for 72 h. The data are expressed as the percentage of living cells compared with the negative control. Results are representative of three independent experiments. Error bars represent the S.D. of triplicate results

number of natural products have been reported in the literature to inhibit STAT3 activity. Curcumin, a natural product from the herb *Curcuma long*, inhibited JAK-STAT signaling and suppressed cell cycle arrest and cell invasion *in vitro*.⁴² Derivatives of the alkaloid berbamine were cytotoxic against melanoma cells harboring aberrant STAT3 activity, and decreased pSTAT3, pJAK2, and pSRC levels in those cells.⁴³ Resveratrol, a widely studied polyphenolic compound found in variety of plants, inhibited constitutive and IL-6-induced STAT3 activity in multiple tumor cell types, leading to the decreased expression of anti-apoptotic proteins and the induction of apoptosis *in vitro*.⁴⁴ Resveratrol and its derivatives have also been reported to suppress breast cancer cell proliferation via inhibition of STAT3 mediated MMP9 expression⁴⁵ and to sensitize multiple myeloma, lung prostate, or pancreatic cancer cells to radiation or chemotherapeutic agents *in vitro*.⁴⁶ However, despite many of these natural products reaching clinical status, the precise mechanism of STAT3 inhibition by these candidates has yet to be fully elucidated.¹⁴

In this study, the benzofuran derivative **1** was identified as a potential inhibitor of STAT3 dimerization using *in silico* screening. Benzofurans are common structural motifs present in numerous bioactive natural products and pharmaceuticals⁴⁷ owing to their diverse biological activities, such as anti-angiogenic⁴⁸ and antitumor⁴⁹ properties. To the best of our knowledge, compound **1** has not been reported as a STAT3 inhibitor, and no biological activity of compound **1** has been presented in the literature.

In *in vitro* experiments, compound **1** inhibited STAT3 DNA-binding activity and reduced STAT3-driven luciferase activity in cells, but had no effect on IFN- α or IFN- γ -induced STAT1 DNA-binding activity or IFN- γ -induced STAT1-driven luciferase activity. Furthermore, compound **1** inhibited the interaction of STAT3-Flag and STAT3-GFP in protein lysates of treated cells, suggesting that the compound was able to antagonize STAT3 dimerization *in cellulo*, but had no effect on the interaction between STAT1-Flag and GFP-STAT1. Compound **1** also suppressed IL-6 or IFN- α -induced STAT3 Y705 phosphorylation in HepG2 cells and HeLa cells, respectively, but had no effect on IFN- α -induced STAT1 phosphorylation in HeLa cells. Additionally, compound **1** showed no significant effect on total STAT3 or STAT1 expression. Finally, in the cell viability assay, compound **1** exhibited pronounced cytotoxicity against the neoplastic HepG2, RAW264.7, and Caco2 cell lines, but was benign towards the normal liver cell line LO2. A possible mechanism for the selective cytotoxicity for compound **1** towards cancer cells over normal cells could be due to the enhanced requirement for STAT3 activity in cancer cells, leading to a greater reduction in cell proliferation upon inhibition of STAT3 signaling by compound **1**.

Taken together, these data suggest that compound **1** acts as an inhibitor of STAT3 transcriptional activity through the disruption of STAT3 dimerization, which could contribute, at least in part, to the observed cytotoxicity of compound **1**. Based on the molecular modeling analysis, we anticipate that

compound **1** may bind to the SH2 domain of STAT3 and block the STAT3 protein–protein interaction. Alternatively, the inhibition of STAT3 phosphorylation by compound **1** could also indirectly affect STAT3 dimerization. Further experiments will have to be performed to investigate the precise mechanism of STAT3 inhibitory activity of compound **1**.

The binding of compound **1** to STAT3 was examined by molecular modeling. In the low-energy conformation of compound **1** in the SH2 domain, the small molecule was predicted to form hydrogen-bonding interactions with Ser611, Glu612, and Arg609. In the native protein, residues Arg609 and Ser611 are involved in direct polar interactions with phospho-Tyr705, and are, therefore, important elements of the reciprocal phosphotyrosine-SH2 interactions.⁴¹ Notably, the ester group of compound **1** is located near residue Thr620 in the 'pY + 1' binding sub-site, which is a region that has been noted to be more divergent among STATs.³² On the other hand, docking of compound **1** to STAT1 or STAT3 resulted in less favorable binding interactions and inferior binding scores. The distinct binding modes of compound **1** to STAT3 *versus* STAT1 or STAT5, despite their similar SH2 domains, provides a possible basis for the observed STAT3-specificity of compound **1**. Other STAT3 small-molecule inhibitors in the literature that have been reported to inhibit STAT3 dimerization via binding to the SH2 domain include STA-21⁵⁰ and cryptotanshinone,⁵¹ which are in clinical trials for treatment of psoriasis⁵² and polycystic ovary syndrome,⁵³ respectively, as well as S3I-M2001,⁵⁴ LLL12,⁵⁵ FLLL32,⁵⁶ S3I-201.1066,⁵⁷ and BP-1-102.⁵⁸ Very recently, the groups of Luk and Lunku have identified the natural product garcinol as an antagonist of STAT3 dimerization, via binding to the SH2 domain, and demonstrated its ability to inhibit human hepatocellular carcinoma growth *in vitro* and *in vivo*.⁵⁹

As described in the introduction, the high degree of conservation in the SH2 domain of STAT proteins, including STAT3 and STAT1, makes the development of selective STAT inhibitors a difficult challenge.³² Owing to the different biological effects of STAT3 and STAT1, the requirement for STAT3-specificity has been thought to be important for the development of novel therapeutics agents. As a counterpoint, resveratrol³³ and curcumin,⁶⁰ which have been shown to inhibit both STAT3 and STAT1, have progressed to clinical trials for a variety of malignancies, suggesting that absolute specificity for STAT3 is not an essential requirement for beneficial activity. In this study, compound **1** showed selectivity for STAT3 over STAT1 against STAT DNA-binding activity *in vitro*, as well as in terms of STAT-directed transcription, STAT dimerization, and STAT phosphorylation *in cellulo*. The selectivity of compound **1** for STAT3 was supported by the molecular modeling analysis, which predicted a lower binding score and a distinct binding interaction between compound **1** with the SH2 domain of STAT3 *versus* STAT1 or STAT5, including contact with the less-conserved pY + 1 region. This study also confirmed the selectivity of the inhibitor S3I-201 for STAT3 over STAT1, which was consistent with findings in previous studies.^{22,61}

In conclusion, we have utilized structure-based virtual screening to discover a natural product-like inhibitor of STAT3. Compound **1** was able to inhibit STAT3 DNA-binding activity in a cell-free system and STAT3-driven luciferase

expression in living cells, with potencies comparable to the well-known STAT3 inhibitor S3I-201. Additionally, compound **1** antagonized STAT3 dimerization and STAT3 tyrosine phosphorylation *in cellulo*. Our molecular modeling analysis suggested that compound **1** might putatively function as an inhibitor of STAT3 dimerization. This study also validates the use of structure-based molecular docking to discover novel inhibitors of protein–protein interactions, which are typically considered difficult to target with small molecules. We envisage that compound **1** may be employed as a useful scaffold for the development of more potent STAT3 dimerization inhibitors.

Materials and Methods

Cells and reagents. TransAM Transcription Factor ELISA was obtained from Active Motif (Carlsbad, CA, USA). Dual-Luciferase Reporter Assay System and pRL-TK vector were purchased from Promega (Madison, WI, USA). pLuc-TK/STAT3 was purchased from Beyotime (Shanghai, Chian). Human epidermal growth factor (EGF) was purchased from BioVision Inc. (Mountain View, CA, USA). Interferon gamma protein (IFN- γ) was purchased from Abcam (Cambridge, MA, USA). Interleukin 6 (IL-6) was obtained from Sino Biological Inc. (Beijing, China). All antibodies were purchased from Cell Signaling Technology (Beverly, MA, USA). HeLa cells and HepG2 cells were cultured in DMEM containing 10% fetal bovine serum were incubated at 37 °C/5% CO₂ and maintained at a cell density of 0.2–1 × 10⁶ cells/ml. HeLa cells were generously provided by Wendy Yeung (The Hong Kong University of Science and Technology, Hong Kong). COS-7 cells and HEK293T cells were gifts from Ming-Yuen Lee (The University of Macau, Macao). All the compounds were dissolved in dimethyl sulfoxide (DMSO).

STAT3 and STAT1 transcription factor ELISA. The STAT3 and STAT1 DNA-binding assay was performed using TransAM Transcription Factor ELISA (Active Motif) according to the manufacturer's instructions. Briefly, HepG2 cells were stimulated with EGF (100 ng/ml) for activation of STAT3 for 15 min. COS-7 cells were stimulated with IFN- γ (25 ng/ml) for activation of STAT1 for 1 h.⁶² HeLa cells were stimulated with IFN- α (1000 U/ml) for activation of STAT3 and STAT1 for 1 h. Cells were harvested and nuclear extract (1 μ g) containing activated STAT3/STAT1 was added with compound (20 μ l) and complete binding buffer (30 μ l) to microtitre wells coated with the STAT3 consensus sequence. The mixture was incubated at room temperature for 1 h. The wells were washed three times with 1 × wash buffer, and incubated with STAT3/STAT1 α antibody for 1 h. The wells were washed as before and incubated with horseradish peroxidase-conjugated secondary antibody at room temperature for 1 h. The wells were washed as before, incubated with 100 μ l of developing solution, quenched with 100 μ l stop solution, and the absorbance was measured at $\lambda = 450$ nm.

Dual luciferase reporter assay. HeLa cells were co-transfected with pSTAT3-TA-luc (Beyotime) and pRL-TK (Promega) as a transfection efficiency control in a 6-well plate. After 12 h, cells were transferred to a 48-well plate with complete medium and cultured for 24 h. Cells were pre-treated with different concentration of compounds (in 0.05% DMSO) for 6 h in low FBS medium, and EGF (100 ng/ml) was subsequently added into the wells for 30 min to stimulate STAT3 expression and the cells were incubated for 30 min. In the IFN- α -induced comparative assay, HeLa cells were co-transfected with p-GAS-TA-luc (Biovector, Beijing, China) and pRL-TK in a 24-well plate for 6 h. Cell were cultured with complete medium for another 18 h. The indicated concentrations of compound **1** and S3I-201 were added to each well for 5 h, followed by the addition of 1000 U/ml IFN- α for 1 h to stimulate STAT3/STAT1 expression. Cell lysates were collected according to the dual luciferase assay protocol (Promega). Sample light output was analyzed using a luminometer and the resulting data were normalized relative to pRL-TK values.

Co-immunoprecipitation assay to evaluate STAT3 and STAT1 dimerization. HEK293T cells were co-transfected with pRc/CMV-STAT3-Flag (Addgene, Cambridge, MA, USA) and pCMV6-AC-STAT3-GFP (OriGene, Rockville, MD, USA) in a 6-well plate for 6 h. In the IFN- α -induced comparative assay, HeLa cells were co-transfected with pRc/CMV-STAT1-Flag (Addgene) and pcDNA-GFP-STAT1 (Addgene) in a 6-well plate for 6 h. Cells were cultured in

complete medium for another 12 h, and then treated with compounds (in 0.05% DMSO) or vehicle control in low FBS medium for an additional 6 h. Cell lysates were harvested and 30 μ g of each protein sample was pulled down with ANTI-FLAG M2 magnetic beads (Sigma-Aldrich, St. Louis, MO, USA) according to the manufacturer's protocol. The purified protein samples were denatured for 5 min and subsequently subjected to SDS-PAGE. The protein samples were then transferred to a PVDF membrane and blocked with 5% non-fat dry milk in TBST for 1 h at room temperature. The membranes were then blotted with anti-Flag (Sigma-Aldrich) and anti-GFP (OriGene) antibody at 4 °C for overnight at a dilution of 1 : 1000. The membranes were washed three times and blotted with secondary antibodies at a dilution of 1 : 10 000 at room temperature for 1 h. After washing for 30 min, membranes were analyzed with enhanced chemiluminescent Plus reagents (GE Healthcare, Waukesha, WI, USA).

Western blot analysis of STAT3 and STAT1 phosphorylation.

HepG2 cells were seeded in a 6-well plate at a density of 5×10^5 ml^{-1} in complete medium and cultured for 12 h. Cells were then pre-treated with compounds (in 0.05% DMSO) or vehicle control in low-FBS medium for 6 h. IL-6 (25 ng/ml) was added to the wells to induce the phosphorylation of STAT3.^{63,64} After 30 min, cells were harvested and protein samples were quantified by the Pierce BCA Protein Assay Kit. 30 μ g of each protein sample was separated by SDS-PAGE. In the IFN- α -induced comparative assay, HeLa cells were seeded in a 6-well plate at a density of 5×10^5 ml^{-1} in complete medium and cultured for 12 h. Cells were then pre-treated with compounds (in 0.05% DMSO) or vehicle control in low-FBS medium for 6 h. IFN- α (1000 U/ml) was added to the wells to induce the phosphorylation of STAT3 and STAT1. After 1 h, cells were harvested and proteins samples were quantified. 30 μ g of each protein sample was separated by SDS-PAGE. The proteins were transferred to a PVDF membrane, and blocked with 5% non-fat dry milk in TBST for 1 h at room temperature. The membranes were incubated with primary antibodies (pY705-STAT3, pY701-STAT1, STAT3, STAT1 and GAPDH antibodies, Cell Signaling Technology) overnight. After treatment with secondary antibodies, membranes were analyzed with enhanced chemiluminescent Plus reagents (GE Healthcare).

Cell viability assay. LO2, HepG2, RAW264.7, Caco2 cells were seeded at 4000 cells per well in a 96-well culture microplate and incubated overnight at 37 °C. Serial dilutions of compound 1 were added to each well and the microplate was incubated at 37 °C, 5% CO₂, 95% air in a humidified incubator for 72 h. Hundred microliter of MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-tetrazolium bromide) reagent (1 mg/ml) was added to each well. After 4 h, the medium was replaced with 100 μ l of DMSO and the wells were incubated at room temperature for 10 min with shaking. Color intensity was measured at 570 nm using a microplate reader. The IC₅₀ value of compound 1 was determined by the dose dependence of the surviving cells after exposure to 1 for 72 h.

Molecular docking and virtual screening

Model construction. The initial model of STAT3 was derived from the X-ray crystal structure of the STAT3 homodimer bound to DNA (PDB: 1BG1),²² using the molecular conversion procedure implemented in the ICM-pro 3.6-1d program (Molsoft, San Diego, CA, USA).⁶⁵ The molecular conversion procedure implemented in ICM-pro 3.6-1d program can read, build, convert, refine, analyze, and superimpose molecules and provide target evaluation to generate three dimensional models. DNA was removed and only one of the monomers was used. Hydrogen and missing heavy atoms were added to the receptor structure, and atom types and partial charges were assigned. The model was then subjected to local energy minimization to identify the optimal position by using the ICM biased probability Monte Carlo algorithm⁶⁶ and analytical derivatives in the internal coordinates. The optimization gradient was 0.05 kcal/M/radian.⁶⁶ For the selectivity analysis, X-ray crystal structures of STAT1 (PDB: 1BF5) and STAT5 (PDB: 1Y1U) were processed in a similar fashion.

High-throughput molecular docking. A chemical library containing over 90 000 natural product or natural product-like compounds (ZINC natural product database) was docked to the molecular model of STAT3 *in silico*. Molecular docking was performed using the virtual library screening module in the ICM-Pro 3.6-1d program (Molsoft). In the ICM fast docking and VLS procedure, the receptor all-atom model was converted into energy potential maps calculated on a fine 3D grid (0.5 Å cell). The grid potential maps account for van der Waals, hydrogen bonding, hydrophobic, and electrostatic interactions between ligand and receptor. The search

area for molecular docking was restricted to the SH2 domain of the STAT3 monomer. Each compound in the library was assigned the MMFF⁶⁷ force field atom types and charges and was then subjected to Cartesian minimization. During the docking analysis, the ligand was represented by an all-atom model and considered fully flexible in the potential field of the receptor, and the binding pose and internal torsions were sampled by the biased probability Monte Carlo (BPMC) minimization procedure, which involved local energy minimization after each random move. Each compound was docked to the protein complex binding pocket, and a score from the docking was assigned to each compound according to the weighed component of the ICM scoring function (see below). Each compound was docked three times to ensure the convergence of the Monte Carlo optimization, and the minimum score of each ligand from the three independent docking experiments was retained and used for ranking. The docking procedure takes about 30 s of time per compound on a Intel Xeon 2.8 GHz CPU using a 100 processor Linux cluster. A permissive cutoff score of -30.0 was chosen in order to weed out potential non-binding ligands and to reduce the size of compounds tested *in vitro*. The binding score distribution of all the compounds is depicted in Supplementary Figure S1. Fourteen compounds were purchased for *in vitro* biological testing.

ICM full-atom ligand-receptor complex refinement and scoring. Once the ligand-receptor complexes are generated by molecular docking, they have to be subjected to complex refinement and scoring. According to the ICM method,⁶⁸ the molecular system was described using internal coordinates as variables. Energy calculations were based on the ECEPP/3 force field with a distance-dependent dielectric constant. The BPMC minimization procedure was used for global energy optimization. This procedure consisted of four iterative steps. The BPMC global-energy-optimization method consists of (1) a random conformation change of the free variables according to a predefined continuous probability distribution; (2) local energy minimization of analytical differentiable terms; (3) calculation of the complete energy including non-differentiable terms such as entropy and solvation energy; (4) acceptance or rejection of the total energy based on the Metropolis criterion and return to step (1). The binding between the small molecules and STAT3 were evaluated with a full-atom ICM ligand binding score⁶⁹ from a multi-receptor screening benchmark as a compromise between approximated Gibbs free energy of binding and numerical errors. The scoring function should give a good approximation of the binding free energy between a ligand and a receptor and is usually a function of different energy terms based on a force field. The ICM scoring function is weighted according to the following parameters (i) internal force field energy of the ligand, (ii) entropy loss of the ligand between bound and unbound states, (iii) ligand-receptor hydrogen bond interactions, (iv) polar and nonpolar solvation energy differences between bound and unbound states, (v) electrostatic energy, (vi) hydrophobic energy, and (vii) hydrogen bond donor or acceptor desolvation. The lower the ICM score, the higher the chance the ligand is a binder. The score was calculated by:

$$S_{\text{bind}} = E_{\text{int}} + T\Delta S_{\text{for}} + E_{\text{vw}} + \alpha_1 E_{\text{el}} + \alpha_2 E_{\text{hb}} + \alpha_3 E_{\text{np}} + \alpha_4 E_{\text{sf}}$$

where E_{vw} , E_{el} , E_{hb} , E_{np} , and E_{sf} are van der Waals, electrostatic, hydrogen bonding, and nonpolar and polar atom solvation energy differences between bound and unbound states, respectively. E_{int} is the ligand internal strain, ΔS_{for} is its conformational entropy loss upon binding, and $T = 300$ K, and α_i are ligand- and receptor-independent constants.⁷⁰

Data analysis. All data were reported as the means of at least three separate experiments. Group comparisons between the control group and various drug treatment groups were done by a one-way ANOVA using GraphPad Prism software (Prism). Significance was considered if a P -value of < 0.05 was reached. Densitometry of western blots was performed by scanning of the exposed film and using Quantity One analysis software (Bio-Rad, Richmond, CA, USA).

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgements. This work is supported by Hong Kong Baptist University (FRG2/12-13/021 and FRG2/13-14/008), Centre for Cancer and Inflammation Research, School of Chinese Medicine (CCIR-SCM, HKBU), the Health and Medical Research Fund (HMRF/13121482), the Research Grants Council (HKBU/201811, HKBU/204612 and HKBU/201913), the French National Research

Agency/Research Grants Council Joint Research Scheme (A-HKBU201/12), the Science and Technology Development Fund, Macao SAR (103/2012/A3, 001/2012/A) and the University of Macau (MYRG091(Y3-L2)-ICMS12-LCH, MYRG121(Y3-L2)-ICMS12-LCH, MRG007/LCH/2014/ICMS and MRG023/LCH/2013/ICMS).

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