Review

www.nature.com/cddis

Defining the role of the Bcl-2 family proteins in Huntington's disease

J Sassone*,¹, A Maraschi^{1,3}, F Sassone^{1,3}, V Silani^{1,2} and A Ciammola*,¹

B-cell lymphoma 2 (Bcl-2) family proteins regulate survival, mitochondria morphology dynamics and metabolism in many cell types including neurons. Huntington's disease (HD) is a neurodegenerative disorder caused by an expanded CAG repeat tract in the *IT15* gene that encodes for the protein huntingtin (htt). *In vitro* and *in vivo* models of HD and HD patients' tissues show abnormal mitochondrial function and increased cell death rates associated with alterations in Bcl-2 family protein expression and localization. This review aims to draw together the information related to Bcl-2 family protein alterations in HD to decipher their potential role in mutated htt-related cell death and mitochondrial dysfunction.

Cell Death and Disease (2013) **4**, e772; doi:10.1038/cddis.2013.300; published online 15 August 2013 **Subject Category:** Neuroscience

Facts

- The B-cell lymphoma 2 (Bcl-2) family proteins regulate the outer mitochondrial membrane (OMM) permeability thus controlling the mitochondrial potential, mitochondrial morphology dynamics and the balance between survival and death in many cell types including neurons.
- HD is a neurodegenerative disease characterized by progressive neuronal dysfunction and death. In patients with HD and in *in vitro* and *in vivo* HD models, mutated htt expression leads to OMM permeabilization, loss of mitochondrial potential and mitochondrial fragmentation.
- HD cell culture models and HD mouse brain models show alterations in Bcl-2 family protein expression and localization. Autopsy studies in patients' brain have detected Bcl-2 family protein alterations in HD.

Open Questions

- We still need more precise evidence on the role of Bcl-2 family proteins in HD-related cell death.
- We do not know the role of Bcl-2 family proteins in HD-related mitochondrial dysfunction.
- Nor do we know through which molecular pathway Bcl-2 family proteins can cause mitochondrial dysfunctions and cell death in HD.

Introduction

Proteins belonging to the Bcl-2 family are key regulators of the apoptotic mitochondrial pathway. Growing evidence shows that Bcl-2 family proteins also modulate mitochondrial morphology¹⁻³ and cellular metabolism⁴ independently from cell death mechanisms. Given that HD typically leads to progressive neuronal death causing neuronal cells to display several mitochondrial dysfunctions (including decreased mitochondrial potential ($\Delta \psi$), abnormal calcium handling, decreased ATP synthesis capacity and increased mitochondrial fragmentation), research has for long investigated Bcl-2 family proteins in HD models and tissues from patients with HD. These studies highlighted evidence that mutated htt dysregulates Bcl-2 family protein levels and localization. Considering the complex Bcl-2 network, a literature review could help to elucidate these proteins' role in mutated htt-related mitochondrial dysfunction and cell death.

What Evidence Exists for Dysregulated Bcl-2 Family Proteins in HD?

Research has to date identified over 20 Bcl-2 family members classified, according to their function, in three Bcl-2 family subgroups: the pro-survival proteins, the pro-apoptotic BH3-only proteins and the pore-forming proteins. Here, we

³These authors equally contributed to this work.

Keywords: mutated huntingtin; Huntington's disease models; Bcl-2 family proteins

Abbreviations: HD, Huntington's disease; htt, huntingtin; OMM, outer mitochondrial membrane; ER, endoplasmic reticulum; $\Delta\psi$, mitochondrial potential; BH3, Bcl-2 Homology 3; CNS, central nervous system; OPA1, optic atrophy-1 protein; UPR, unfolded protein response; SGC, superior cervical ganglion; BDNF, brain-derived neurotrophic factor; Bcl-2, B-cell lymphoma 2; Bcl-xL, B-cell lymphoma-extra large; Bad, Bcl-2 antagonist of cell death; Bim, Bcl-2 interacting mediator of cell death; Bid, Bcl-2 interacting domain death agonist; Bnip3, Bcl-2/adenovirus E1B 19 kDa interacting protein 3; Puma/Bbc3, p53-upregulated modulator of apoptosis/Bcl-2 binding component 3; Bax, Bcl-2-associated X-protein; Bak, Bcl-2 antagonist/killer

Received 17.5.13; revised 12.7.13; accepted 15.7.13; Edited by A Verkhratsky

ıpg

¹IRCCS Istituto Auxologico Italiano, Department of Neurology and Laboratory of Neuroscience, Milan, Italy and ²Department of Pathophysiology and Transplantation, Dino Ferrari Center, Università degli Studi di Milano, Milan, Italy

^{*}Corresponding author: J Sassone, Department of Neurology and Laboratory of Neuroscience, IRCCS Istituto Auxologico Italiano, via Zucchi 18, Cusano Milanino 20095 Italy. Tel: + 39 02 619113045; Fax: + 39 02 619113033; E-mail: jennysassone@hotmail.com

or A Ciammola, Department of Neurology and Laboratory of Neuroscience, IRCCS Istituto Auxologico Italiano, piazzale Brescia 20, Milan 20149, Italy. Tel: + 39 02 619112937; Fax: + 39 02 619113033; E-mail: a.ciammola@auxologico.it

review the three subgroups, focusing on the Bcl-2 family members relevant for HD and gathering data on the cell culture model SThdH,⁵ cell culture models obtained by transient htt transfection, in genetic mouse models R6/1, R6/2,⁶ Tet/HD94,⁷ N171-82Q,⁸ YAC128⁹ and human cells from patients with HD.

The Pro-survival Bcl-2 Proteins: Bcl-2 and Bcl-xL

Bcl-2 and B-cell lymphoma-extra large (Bcl-xL) function to repress programmed cell death.^{10,11} They are stably inserted in intracellular membranes (OMM and the endoplasmic reticulum (ER) membrane) and inhibit cell death by sequestering the pro-apoptotic Bcl-2 family proteins into inactive complexes.¹² In the normal central nervous system (CNS), Bcl-2 is widespread expressed during the embryonic stage, whereas after birth it progressively declines with aging. Studies in adult monkey and adult human CNS disclosed, apart from rare Bcl-2 positive neurons, that most neurons and astrocytes express no Bcl-2 and that most Bcl-2 protein in adult brain results from microglial expression.¹³ Bcl-xL is encoded by the Bcl-x gene and, differently from Bcl-2, retains its expression in adult neurons at a relatively high level, predominantly in Purkinje cells in the cerebellum, in cerebral cortical neurons and in hippocampal neurons.14,15

Bcl-2 level alterations in HD remain controversial. Bcl-2 levels were first investigated in HD cell culture models: mutated htt expression significantly decreased Bcl-2 protein levels in the neuroblastoma cell line Neuro2A¹⁶ and significantly decreased Bcl-2 transcript and Bcl-2 protein levels in immortalized striatal cells.^{17,18} Although these results strongly suggest that mutated htt expression decreases Bcl-2 levels, they provide no definitive proof given that the protooncogene *Bcl-2* is highly dysregulated in many tumor cell lines including those of neural origin.^{19,20} Hence, data from immortalized cell cultures might not correspond to Bcl-2 levels in post-mitotic neurons.

Others have investigated Bcl-2 levels also in brains from HD mouse models. A first study detected no Bcl-2 protein level changes in the total brain or in mitochondrial fractions from R6/2 mice as compared with littermate controls.²¹ Accordingly, two independent studies reported no differences in Bcl-2 mRNA and protein levels between R6/1 and control mice.^{22,23} Conversely, more recent studies disclosed decreased Bcl-2 mRNA and protein levels in R6/2 mouse brains^{17,24} and N171-82Q mice.²⁵

After reappraising these seemingly contradictory findings, we find it difficult to provide a definitive answer about whether Bcl-2 levels are dysregulated in HD mouse models. To understand the potential protective role of Bcl-2 in HD, Zhang *et al.*²¹ crossed R6/2 mice with transgenic mice selectively overexpressing Bcl-2 in neurons: double transgenic R6/2-Bcl-2 mice survived longer than R6/2 littermates and their motor deficits had a significantly later onset. Notwithstanding contradictory data about Bcl-2 expression in HD mouse models, this result suggests that Bcl-2 overexpression can protect neurons from toxicity elicited by mutated htt.

Bcl-2 expression was analyzed by immunostaining in post-mortem controls and the HD patients' caudate nucleus.

Control neurons had low or negative Bcl-2 signals, in agreement with a previous report,¹³ whereas Bcl-2 labeling was stronger in HD neurons, and Bcl-2 expression reached maximum in the brains of HD patients with longer disease duration.²⁶ In accordance with a potential neuroprotective role of Bcl-2 in HD, the investigators suggested that HD patients' neurons may increase Bcl-2 expression in an attempt to survive. An alternative hypothesis is that Bcl-2 protein in HD brains is cleaved by caspase proteases activated by mutated htt expression.²⁷ If so, the caspase-dependent cleavage inactivates Bcl-2 antiapoptotic function and converts Bcl-2 into a Bcl-2 associated X-protein (Bax)-like protein that enhances cell death.²⁸ According to this hypothesis, rather than inhibiting cell death, high Bcl-2 levels in HD cells could worsen neurodegeneration. The Bcl-2 upregulation in HD patients seems to be brain-specific given that no differences were found between Bcl-2 protein levels in peripheral blood cells from patients with HD and from controls.²⁹

Few data are available about the other pro-survival protein, Bcl-xL in HD: Bcl-xL protein levels were investigated in the brain of HD mice, but no significant difference was found in total-brain lysates, striatum lysates or mitochondrial fractions in HD mice and controls.^{21,22,30} To our knowledge, only one report suggests a role of Bcl-xL in HD. The authors showed that Bcl-xL expression prevented the htt proteolysis induced by DNA damage and hypothesized that blocking Bcl-xL may activate the caspases that cause htt proteolysis.³¹

The Pro-apoptotic BH3-only Proteins

The pro-apoptotic BH3-only proteins, by responding to specific death and survival signals, function as sensors of cellular damage. Activating one or more among the BH3-only proteins culminates in allosteric activation of the pore-forming proteins Bax and Bcl-2 antagonist/killer (Bak).

BH3-only proteins could activate Bax and Bak through two mechanisms. The 'direct activation' model posits that certain BH3-only proteins termed 'activators' (eg, Bcl-2 interacting mediator of cell death (Bim), Bcl-2 interacting domain death agonist (Bid)/truncated Bid (tBid) and p53 upregulated modulator of apoptosis (Puma)) bind to Bax and Bak directly triggering their oligomerization in the OMM, whereas other BH3-only proteins, termed 'sensitizers' (eg, Bcl-2 antagonist of cell death (Bad), Bcl-2/adenovirus E1B 19 kDa interacting protein 3 (Bnip3) and Puma) would bind only to pro-survival Bcl-2 family members, thus liberating 'activator' proteins. Instead, the 'indirect activation' model posits that all the BH3-only proteins bind to pro-survival Bcl-2 family members, thereby preventing them from binding to and neutralizing Bax or Bak.³²

Bcl-2 Antagonist of Cell Death

Bad has a double function: it separately regulates apoptosis and glucose metabolism in multiple cell types including neurons.^{33,34} Investigating *in vitro* HD models, Rigamonti *et al.*³⁵ showed that Bad overexpression in immortalized striatal cells induces apoptosis only in clones co-expressing mutated htt, thus suggesting that mutated htt toxicity induces Bad-dependent cell death. For HD mouse models, no differences in striatal Bad protein levels were found between R6/1 and littermate controls.²² Also in the R6/2 mouse, no change was found in the brain Bad protein levels. Conversely, the investigators found a significant decrease in phosphorylated Bad (pBad) during the late-HD stages in R6/2 mice.²¹ Because pBad promotes mitochondrial respiration and ATP production, whereas non-pBad binds the anti-apoptotic partners Bcl-2 and Bcl-xL eliciting Bax and Bak activation,³⁶ the low pBad/Bad ratio found in R6/2 brain lysates suggests that Bad activity shifts from positive metabolic to negative apoptotic function in HD cells.

Bcl-2 Interacting Mediator of Cell Death

The BH3-only pro-apoptotic protein Bim is expressed in three major isoforms generated by alternative splicing: Bim short (BimS), Bim long (BimL) and Bim extra-long (BimEL). All these isoforms neutralize activity in pro-survival Bcl-2- like proteins, but they differ in pro-apoptotic potency, BimS being the most effective and BimEL the least effective killer.³⁷ In the mouse and human CNS, Bim is expressed primarily in neurons, and the most expressed isoform in the brain is BimEL.^{38,39}

Several observations show that mutated htt causes BimEL accumulation in various cell types. In particular, mutated htt expression induces BimEL accumulation in HEK293T cells, in Neuro-2a cells,⁴⁰ in sympathetic superior cervical ganglion neurons⁴¹ and in immortalized striatal cells.⁴² BimEL silencing in Neuro-2a cells significantly reduces cell death elicited by mutated htt.⁴⁰

In the mouse model R6/2, high BimEL levels were found in total-brain lysates and in mitochondrial fractions specifically at the late stages of disease.^{21,40} These data found further confirmation in two other HD mouse models showing increased striatal BimEL in R6/1 mice at the late stages of disease and in the conditional model Tet/HD94.²² Collectively, these results demonstrate that mutated htt causes BimEL accumulation and translocation to mitochondria.

Bcl-2 Interacting Domain Death Agonist

The BH3-only pro-apoptotic protein Bid is involved in neuronal cell death in many neurological disorders such as stroke,⁴³ ischemia⁴⁴ and Alzheimer's disease.⁴⁵ Full-length Bid has extremely weak pro-apoptotic activity but reaches its strongest pro-apoptotic activity after proteolytic cleavage by several proteases that produce truncated Bid (tBid).⁴⁶ Bid protein is widely expressed in embryonic and postnatal brain, and its expression in post-mitotic neurons in the limbic system, basal ganglia, mesencephalic tectum and cerebellum persists at a high level into adulthood.¹⁵

Several lines of evidence show that mutated htt causes Bid and tBid accumulation in neurons and in mitochondrial neuronal fractions. In particular, western blot analysis for Bid and tBid detection in HeLa cells and Neuro2A cells disclosed that mutated htt transfection causes Bid cleavage.¹⁶

Increased Bid cleavage was also detected in brain lysates from R6/2 mice at the middle stage of HD.²¹ Different results

come from the R6/1 model in which Garcìa-Martìnez *et al.*²² found no tBid accumulation, but instead found full-length Bid at high levels in the striatum and in striatal mitochondrial fractions. The Bid protein increase correlated with enhanced Bid mRNA expression. Increased full-length Bid protein levels were also found in the striatum in the conditional mouse model of HD, Tet/HD94: Bid accumulation specifically depended on mutated htt expression, given that transgene suppression completely reverted Bid protein to wild-type levels.²²

Bcl-2/adenovirus E1B 19kDa Interacting Protein 3

Bnip3 has had an emerging role in human health, as convincing evidence implicated its death, inducing activity in heart diseases, whereas Bnip3 loss of function was associated with tumor growth.^{47,48} Current knowledge shows that Bnip3 is involved in cell death, autophagy and programmed mitochondrial clearance.⁴⁹ Bnip3 induces cell death through at least two distinct mechanisms: Bnip3 can engage anti-apoptotic Bcl-2 family members to trigger Bax-Bak-dependent OMM permeability⁵⁰ or can induce a novel mitochondrial leak pathway by interacting with the optic atrophy-1 protein (OPA1).^{51,52}

Because Bnip3 is expressed in the brain and skeletal muscle.53,54 in recent years, we undertook a study aimed to investigate the potential role of Bnip3 in HD by assessing Bnip3 level and localization in htt-transfected neuronal cells. brain in HD mouse models and in muscle cells from HD patients.⁵⁵ We observed that mutated htt expression causes Bnip3 accumulation in whole-cell lysates and in the mitochondrial fraction of SHSY5Y and HEK293T cells. We also found higher Bnip3 protein levels in mitochondrial fractions from R6/2 mice and in striatum from YAC128 mice than in littermate controls. Finally, we observed that Bnip3 mainly co-localized with the mitochondria in HD patients' muscle cells, whereas in control cells it mainly localized in the cytosol and nucleus. We also observed that the expression of a dominant-negative Bnip3 named Bnip3 Δ TM^{56,57} rescued the mitochondrial membrane potential loss in HD muscle cells.⁵⁵ Overall, these data suggest that mutated htt enhances Bnip3 activity in neuronal and muscular cells.

p53-upregulated Modulator of Apoptosis/Bcl-2 Binding Component 3 (Puma/Bbc3)

Puma/Bbc3 is a BH3-only protein identified in 2001.^{58,59} Puma/Bbc3 appears not to be expressed in the normal adult brain, but its expression is strongly induced in some brain diseases such as status epilepticus⁶⁰ and cerebral ischemia.⁶¹ Originally identified as a transcriptional target of p53,^{58,59} Puma/Bbc3 is also transcriptionally induced in neuronal cells undergoing ER stress-induced unfolded protein responses (UPR).⁶² Few data are available about Puma/Bbc3 expression in HD models: in SCG neurons, mutated htt expression leaves Puma/Bbc3 mRNA unchanged,⁴¹ whereas in PC12 cells it increases Puma/Bbc3 protein sixfold. Although Puma/Bbc3 protein accumulation in HD may depend on mutated htt-induced ER stress, because this datum remains unconfirmed in other HD models, the possible pathogenetic role of Puma/Bbc3 in HD needs further clarification.

The Pore-forming Proteins Bax and Bcl-2 Antagonist/ Killer (Bak)

Bax and Bak act downstream of the pro-survival and BH3-only members and have a key role in the mitochondrial apoptotic pathway. Bak is normally inserted in the OMM; Bax is predominantly cytosolic, but, once activated, it translocates from the cytosol into the OMM where together with Bak it forms the apoptotic pore, the point of no return in mitochondrial apoptosis.⁶³ Besides inducing apoptosis, Bax and Bak influence mitochondrial fission–fusion dynamics in healthy cells.^{1,2} Both Bax and Bak are expressed in neurons: Bax is expressed in many regions in the human CNS^{64,65} including the human caudate nucleus;²⁶ and Bak is expressed in the human brain at the fetal, adult and elder stages.⁶⁶

Ample evidence underlining the link between Bax activation and HD pathogenesis comes from research assessing Bax protein levels and localization in neuronal cell culture models of HD and in the brains of HD mouse models. Mutated htt transfection in PC12 cells increased Bax protein levels fourfold as compared with wt htt transfection;⁶⁷ in Neuro2A cells, mutated htt expression induced Bax translocation from the cytosol to the mitochondria.⁴⁰ Only one *in vitro* study argues against a Bax role in mutated htt-induced cell death: King *et al.*²¹ showed that in sympathetic superior cervical ganglion (SCG) neurons mutated htt induced Bax-independent cell death.

Studies in HD mouse models showed increased Bax levels in the brain mitochondrial fractions from R6/1 and R6/2 mice^{21,22} and increased Bax mRNA in the cortex and cerebellum from R6/1 mice, brain areas in which Bax mRNA correlated with the number of apoptotic nuclei.²³ Bax expression was also analyzed in the caudate nucleus from HD patients. The cytoplasmic Bax signal was stronger in caudate neurons from HD patients than from controls. Bax expression was already maximal in HD brains at disease onset and the highest in the most severely compromised shrunken and dark neurons.²⁶ Hence, data from HD mouse models and human tissues converge to indicate that mutated htt expression causes Bax protein accumulation and Bax translocation to the OMM in neurons. Bax dysregulation in HD seems not to be brain specific insofar as Bax protein levels were also higher in lymphocytes and monocytes from HD patients than from controls.²⁹ This finding probably depends on the fact that htt is ubiquitously expressed in human tissues.68

Table 1 Evidence for Bcl-2 family protein dysregulation in cell culture models of HD (Significant differences between HD and control cells are highlighted in bold)

Cell culture model	Observations / Results	Reference
Neuro2A cells transfected with htt- exon1 (16Q vs 40Q)	↑tBid and ↓ Bid protein in total lysates ↓BcI-2 protein in total lysates	Majumder <i>et al</i> . ¹⁶
Neuro2A cells transfected with htt- exon1 (25Q vs 103Q)	↑Bax protein in mitochondrial fractions ↑BimEL protein in total lysates and in mitochondrial fractions ↓Bid protein in total lysates	Leon <i>et al.</i> ⁴⁰
STHdh cells expressing full-lenght htt (7Q vs 111Q)	∱BimEL protein in total lysates	Kong et al. ⁴²
STHdh cells expressing full-lenght htt (7Q vs 109Q)	↓BcI-2 mRNA and BcI-2 protein in total lysates	Ju <i>et al.</i> ¹⁷
STHdh cells expressing full-lenght htt (7Q vs 111Q)	\downarrow BcI-2 protein and \downarrow BcI-2/Bax protein ratio in total lysates	Ruiz <i>et al.</i> ¹⁸
HEK293 cells transfected with full- lenght htt (Q17 vs Q47)	个Bnip3 protein in total lysates and in mitochondrial fractions	Sassone <i>et al.</i> ⁵⁵
HEK293 cells transfected with htt- exon1 (25Q vs 72/103Q)	↑BimEL protein and ↑pSer69-BimEL in total lysates ↑BimEL mRNA (with htt 103Q)	Leon <i>et al.</i> ⁴⁰
PC12 cells transfected with htt N171 fragment (18Q vs 82Q)	∱Bax protein in total lysates ∱Puma/Bbc3 protein in total lysates	Bae <i>et al.</i> ⁶⁷
sympathetic superior cervical ganglion neurons infected with htt- exon1 (25Q vs 97Q)	↑BimEL protein in total lysates No change in Puma/Bbc3 mRNA in basal condition	King et al. ⁴¹
SHSY5Y cells trasfected with htt- exon1 (9Q vs 60Q)	↑Bnip3 protein in total lysates and in mitochondrial fractions	Sassone <i>et al.</i> ⁵⁵
HeLa cells transfected with htt- exon1 (16Q vs 40Q)	↓Bcl-2 mRNA ↑tBid and ↓Bid protein in total lysates	Majumder <i>et al.</i> ¹⁶



Through Which Molecular Pathway Could Bcl-2 Family Proteins Cause Mitochondrial Dysfunctions and Cell Death in HD?

By summarizing the data about Bcl-2 family proteins in cell culture models of HD (Table 1), mouse models of HD (Table 2) and cells and tissues from HD patients (Table 3), we formulated a hypothetical model for explaining Bcl-2 family protein activation in HD (Figure 1).

Because many results converge to demonstrate that mutated htt causes BimEL activation (Tables 1 and 2), we hypothesize that BimEL, a protein that in healthy cells associates with cellular microtubule complexes,69 in HD neurons localizes to mitochondria where, according to the 'direct activation' model (paragraph 2.2), it would trigger Bax activation. This hypothesis receives support from evidence that both BimEL and Bax accumulate in the mitochondrial fractions from R6/1 and R6/2 mouse models. Alternatively, according to the 'indirect activation' model, BimEL in HD cells may bind to Bcl-2, thereby preventing it from neutralizing Bax. In this context, Bcl-2 may have a neuroprotective role in HD. This hypothesis receives support from evidence that Bcl-2 overexpression in neurons slows down neurodegeneration in the R6/2 model.²¹ What remains unclear is how mutated htt activates BimEL. An interesting clue comes from evidence that BimEL expression is negatively regulated by the brainderived neurotrophic factor (BDNF).^{70,71} By wide consensus, the HD mutation results in lower BDNF levels in the brain⁷² because htt is indirectly involved in transcriptional control over the BDNF gene.⁷³ Mutated huntingtin might therefore elicit BimEL accumulation/activation by inhibiting BDNF expression. BimEL is also a well-known downstream target of ER stress.74 Insofar, as proteins with an abnormally long

Table 2 Evidence for Bcl-2 family protein dysregulation in mouse models of HD (Significant differences between HD and control cells are highlighted in bold)

Mouse model	Observations / Results	Reference
R6/1	At 18 weeks no change in Bcl-xL protein in the striatum.	Hansson <i>et al.</i> ³⁰
R6/1	At 8 and 12 weeks no change in Bid and BimEL protein in the striatum. At 16 weeks ↑Bid and ↑BimEL protein in the striatum. At 16 weeks no change in Bcl-2, Bcl-xL,Bax, Bak, and Bad protein in the striatum. At 16 weeks no change in Bid and BimEL protein in the cortex. At 30 weeks ↑Bid, ↑Bax and ↑BimEL protein in the striatal mitochondrial fractions. At 30 weeks no change in Bcl-2 and Bcl-xL protein in the striatal mitochondrial fractions. At 30 weeks ↑Bid and ↑BimEL protein in the striatal mitochondrial fractions. At 30 weeks ↑Bid and ↑BimEL protein in the striatal cytosolic fractions. At 30 weeks ↑Bid mantal the striatal cytosolic fractions. At 30 weeks ↑Bid mantal the striatal cytosolic fractions. At 30 weeks no change Bax, Bcl-2 and Bcl-xL protein in the striatal cytosolic fractions.	Garcia-Martinez <i>et al.</i> ²²
R6/1	At 12, 24 and 36 weeks no change in Bcl-2 mRNA in cerebellum and cortex. At 12, 24 and 36 months ∱Bax mRNA in cerebellum and cortex.	Teles <i>et al.</i> ²³
R6/2	At 9 weeks no change in Bad, pBad, BcI-2, BcI-xL, BimEL and Bax protein in total brain. At 9 weeks ↓Bid and ↑tBid protein in total brain. At 12 weeks no change in Bad, BcI-2, BcI-xL and Bax protein in total brain. At 12 weeks ↑BimEL and ↓pBad protein in total brain. At 9 and 12 weeks no change in BcI-2 and BcI-xL protein in mitochondrial fractions. At 9 and 12 weeks ↑Bax protein in mitochondrial fractions. At 12 weeks ↑BimEL protein in mitochondrial fractions. At 12 weeks ↑BimEL protein in mitochondrial fractions.	Zhang <i>et al.</i> ²¹
R6/2	At 13 weeks ∱BimEL protein in the striatum.	Leon <i>et al.</i> ⁴⁰
R6/2	At 10 weeks ↓BNip3 protein in total lysates and ↑in striatal mitochondrial fractions. At 10 weeks no change in BNip3 protein in the cortex.	Sassone <i>et al.</i> ⁵⁵
R6/2	At 10.5 and 12 weeks \downarrow Bcl-2 protein in the striatum.	Chiang et al. ²⁴
R6/2	At 12 weeks ↓Bcl-2 mRNA and Bcl-2 protein in total brain.	Ju et al. ¹⁷
Tet/HD94	At 22 months, transgene ON ↑Bid and ↑BimEL protein in the striatum. At 22 months, transgene OFF no change in Bid and ↑BimEL protein.	Garcia-Martinez <i>et al.</i> ²²
N171-82Q	At 12 weeks ↓Bcl-2 protein in the striatum.	Duan <i>et al.</i> ²⁵
YAC128	At 6 weeks ∱Bnip3 protein in striatum and striatal mitochondrial fractions.	Sassone <i>et al.</i> ⁵⁵

polyglutamine expansion cause ER stress,⁷⁵ BimEL activation in HD may also depend on mutated htt induced-UPR. A question awaiting further research is whether Puma/Bbc3, the other potent BH3-only protein activated in response to ER stress, has a role in HD.

Another BH3-only protein potentially involved in HD pathogenesis is Bid. In HD models, mutated htt elicits Bid cleavage or full-length Bid accumulation or both events (Tables 1 and 2). Bid activation was suggested as a key event in many neurodegenerative diseases because Bid is highly expressed in neurons.⁷⁶ In HD cells, full-length Bid and tBid, by migrating to mitochondria, could sustain Bax activation,⁷⁷ thus amplifying the mitochondrial damage.

The molecular mechanism by which mutated htt causes full-length Bid to accumulate is undefined. Because full-length

Bid pro-apoptotic activity is extremely weak, high Bid expression levels are nevertheless unlikely to cause cell death in HD. Conversely, because tBid exhibits strong proapoptotic activity, it may have a key role in HD cell death. This possibility receives support also from evidence that mutated htt enhances activity of caspase-8^{78, 79} and calpain⁸⁰ that both enzymes are able to cleave full-length Bid.⁴⁶

The most recently BH3-only protein potentially implicated in HD pathogenesis is Bnip3. Our results provide evidence that Bnip3, an apoptotic regulator that in healthy cells localizes to the cytosol or the nuclei, in neuronal and non-neuronal cells expressing mutated htt mainly localizes to the mitochondria.⁵⁵ Because Bnip3 activation causes $\Delta \psi$ loss, mitochondrial fragmentation and mitophagy,^{51,81} Bnip3 could be implicated in the mitochondrial dysfunction in HD. Precisely, how

Table 3 Evidence for Bcl-2 family protein dysregulation in HD patients' cells (Significant differences between HD and control cells are highlighted in bold)

Cells/Tissues	Observations / Results	Reference
Caudate nucleus from 18 HD patients	∱BcI-2 and ∱Bax protein in neurons	Vis <i>et al.</i> ²⁶
Peripheral blood mononuclear cells from 10 HD patients	No change in Bcl-2 protein ↑Bax protein in B/T lymphocytes and in monocytes	Almeida <i>et al.</i> ²⁹
Primary muscular cells from 4 HD patients	个Bnip3 protein in total lysates and in mitochondrial fractions	Sassone <i>et al.⁵⁵</i>



Figure 1 Hypothetical model explaining the pathway through which mutated htt (mhtt) causes OMM permeabilization. Bax activation in HD may depend on the 'activators' BH3-only proteins and the 'sensitizers' that bind only to pro-survival proteins Bcl-2 and Bcl-xL. Mutated htt may interact with the Bcl-2 protein network at multiple levels by modulating Bcl-2 expression, inducing Bid accumulation and Bid cleavage, promoting BNip3 activation and increasing non-pBad levels. BimEL accumulation/activation may depend on htt control over BDNF expression or ER stress-induced UPR or both mechanisms

mutated htt induces Bnip3 activation remains an unanswered question. Previous reports show that the Bnip3–mitochondria association is strongly stabilized by acidosis^{82,83} or by an increased cytosolic calcium concentration,⁸⁴ conditions that may both take place in HD cells owing to mitochondrial respiration inhibition. An alternative possibility is that htt binds directly or indirectly to Bnip3, but this issue awaits elucidation in future studies.

Summary and Perspective

Current evidence, derived from research efforts by many investigators over a lengthy time-span, increasingly implies that mutated htt adversely influences Bcl-2 family protein levels and localization. Studies conducted in vitro and in vivo provide convincing evidence that mutated htt expression activates at least four BH3-only proteins. Although mutated htt may upregulate each of these four proteins through different mechanisms, their activation culminates in an identical consequence, namely Bax activation. Bax activation, by promoting cytochrome c release, may underpin the progressive neuronal apoptosis in HD patients' brains. Although this hypothesis receives support from early studies that identified apoptotic-like cells in the HD striatum, 85,86 evidence for HD cell death arising through apoptosis alone is controversial.87 An alternative possibility is that Bcl-2 protein family dysrequlation alters HD mitochondrial dynamics in an apoptosisindependent manner. Especially interesting in this context is a report that Bax-induced mitochondrial fission and Baxinitiated cytochrome c release are separable events and that Bcl-2 family proteins can influence mitochondrial fissionfusion dynamics in HD cells independently of apoptosis.^{1,2} If true. Bcl-2 family proteins in HD may be responsible for the fragmented mitochondrial morphology, changes in mitochondrial ultrastructure and impaired mitochondrial trafficking demonstrated in in vitro and in vivo models of HD.88-90

Future studies should aim to go beyond analyzing Bcl-2 family protein levels because their expression level often poorly indicates protein activation, especially given that most BH3-only members undergo strong regulation by post-translational mechanisms. Future studies aimed to elucidate the molecular mechanisms underlying the Bcl-2 protein interactions documented in *in vitro* and *in vivo* HD models will exploit innovative fluorescence techniques⁹¹ that may more clearly illustrate the Bcl-2 role in HD. Equally important are the genetic approaches such as crossing HD mouse models onto mice knockout for *Bcl-2* genes that will provide definitive proof that Bcl-2 family members are pathogenetically involved in HD.

Finally, future studies involving innovative human cellular models such as iPS from patients with HD, will clarify whether Bcl-2 family proteins, already molecular targets in cancer therapy,⁹² may also be a therapeutic target for HD.

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgements. JS, AM and AC are supported by Cariplo Foundation (grant number 2012-0593). JS is also supported by Italian Health Minister (grant number GR-2010-2318394). FS is supported by Associazione Mauro

Emolo O.N.L.U.S and AICH Milano O.N.L.U.S. We thank Huntington's patients and their families for their precious support. We also thank the anonymous reviewer for helpful comments on the manuscript.

- Karbowski M, Norris KL, Cleland MM, Jeong SY, Youle RJ. Role of Bax and Bak in mitochondrial morphogenesis. *Nature* 2006; 443: 658–662.
- Sheridan C, Delivani P, Cullen SP, Martin SJ. Bax- or Bak-induced mitochondrial fission can be uncoupled from cytochrome C release. *Mol Cell* 2008; 31: 570–585.
- Berman SB, Chen YB, Qi B, McCaffery JM, Rucker EB, Goebbels S et al. Bcl-x L increases mitochondrial fission, fusion, and biomass in neurons. J Cell Biol 2009; 184: 707–719.
- Danial NN, Gramm CF, Scorrano L, Zhang CY, Krauss S, Ranger AM et al. BAD and glucokinase reside in a mitochondrial complex that integrates glycolysis and apoptosis. *Nature* 2003; 424: 952–956.
- Trettel F, Rigamonti D, Hilditch-Maguire P, Wheeler VC, Sharp AH, Persichetti F et al. Dominant phenotypes produced by the HD mutation in STHdh(Q111) striatal cells. *Hum Mol Genet* 2000; 9: 2799–2809.
- Mangiarini L, Sathasivam K, Seller M, Cozens B, Harper A, Hetherington C et al. Exon 1 of the HD gene with an expanded CAG repeat is sufficient to cause a progressive neurological phenotype in transgenic mice. Cell 1996; 87: 493–506.
- Yamamoto A, Lucas JJ, Hen R. Reversal of neuropathology and motor dysfunction in a conditional model of Huntington's disease. *Cell* 2000; 101: 57–66.
- Schilling G, Becher MW, Sharp AH, Jinnah HA, Duan K, Kotzuk JA et al. Intranuclear inclusions and neuritic aggregates in transgenic mice expressing a mutant N-terminal fragment of huntingtin. Hum Mol Genet 1999; 8: 397–407.
- Slow EJ, van Raamsdonk J, Rogers D, Coleman SH, Graham RK, Deng Y et al. Selective striatal neuronal loss in a YAC128 mouse model of Huntington disease. *Hum Mol Genet* 2003; 12: 1555–1567.
- Allsopp TE, Wyatt S, Paterson HF, Davies AM. The proto-oncogene bcl-2 can selectively rescue neurotrophic factor-dependent neurons from apoptosis. *Cell* 1993; 73: 295–307.
- Boise LH, González-García M, Postema CE, Ding L, Lindsten T, Turka LA *et al.* bcl-x, a bcl-2-related gene that functions as a dominant regulator of apoptotic cell death. *Cell* 1993; 74: 597–608.
- Kroemer G, Galluzzi L, Brenner C. Mitochondrial membrane permeabilization in cell death. *Physiol Rev* 2007; 87: 99–163.
- Merry DE, Veis DJ, Hickey WF, Korsmeyer SJ. bcl-2 protein expression is widespread in the developing nervous system and retained in the adult PNS. *Development* 1994; 120: 301–311.
- González-García M, García I, Ding L, O'Shea S, Boise LH, Thompson CB *et al.* Bcl-x is expressed in embryonic and postnatal neural tissues and functions to prevent neuronal cell death. *Proc Natl Acad Sci USA* 1995; 92: 4304–4308.
- Krajewska M, Mai JK, Zapata JM, Ashwell KW, Schendel SL, Reed JC *et al.* Dynamics of expression of apoptosis-regulatory proteins Bid, Bcl-2, Bcl-X, Bax and Bak during development of murine nervous system. *Cell Death Differ* 2002; 9: 145–157.
- Majumder P, Raychaudhuri S, Chattopadhyay B, Bhattacharyya NP. Increased caspase-2, calpain activations and decreased mitochondrial complex II activity in cells expressing exogenous huntingtin exon 1 containing CAG repeat in the pathogenic range. *Cell Mol Neurobiol* 2007; 27: 1127–1145.
- Ju TC, Chen HM, Lin JT, Chang CP, Chang WC, Kang JJ et al. Nuclear translocation of AMPK-alpha1 potentiates striatal neurodegeneration in Huntington's disease. J Cell Biol 2011; 194: 209–227.
- Ruiz C, Casarejos MJ, Rubio I, Gines S, Puigdellivol M, Alberch J et al. The dopaminergic stabilizer, (-)-OSU6162, rescues striatal neurons with normal and expanded polyglutamine chains in huntingtin protein from exposure to free radicals and mitochondrial toxins. Brain Res 2012; 1459: 100–112.
- Reed JC, Meister L, Tanaka S, Cuddy M, Yum S, Geyer C *et al.* Differential expression of bcl2 protooncogene in neuroblastoma and other human tumor cell lines of neural origin. *Cancer Res* 1991; **51**: 6529–6538.
- Placzek WJ, Wei J, Kitada S, Zhai D, Reed JC, Pellecchia M. A survey of the anti-apoptotic Bcl-2 subfamily expression in cancer types provides a platform to predict the efficacy of Bcl-2 antagonists in cancer therapy. *Cell Death Dis* 2010; 1: e40.
- Zhang Y, Ona VO, Li M, Drozda M, Dubois-Dauphin M, Przedborski S et al. Sequential activation of individual caspases, and of alterations in Bcl-2 proapoptotic signals in a mouse model of Huntington's disease. J Neurochem 2003; 87: 1184–1192.
- García-Martínez JM, Pérez-Navarro E, Xifró X, Canals JM, Díaz-Hernández M, Trioulier Y *et al.* BH3-only proteins Bid and Bim(EL) are differentially involved in neuronal dysfunction in mouse models of Huntington's disease. *J Neurosci Res* 2007; 85: 2756–2769.
- Teles AV, Rosenstock TR, Okuno CS, Lopes GS, Bertoncini CR, Smaili SS. Increase in bax expression and apoptosis are associated in Huntington's disease progression. *Neurosci Lett* 2008; 438: 59–63.
- Chiang MC, Chen CM, Lee MR, Chen HW, Chen HM, Wu YS et al. Modulation of energy deficiency in Huntington's disease via activation of the peroxisome proliferator-activated receptor gamma. Hum Mol Genet 2010; 19: 4043–4058.

- 25. Duan W, Peng Q, Masuda N, Ford E, Tryggestad E, Ladenheim B et al. Sertraline slows disease progression and increases neurogenesis in N171-82Q mouse model of Huntington's disease Neuropiol Dis 2008: 30: 312-322
- 26. Vis JC, Schipper E, de Boer-van Huizen RT, Verbeek MM, de Waal RM, Wesseling P et al. Expression pattern of apoptosis-related markers in Huntington's disease. Acta Neuropathol 2005 109 321-328
- 27. Sanchez Mejia RO, Friedlander RM. Caspases in Huntington's disease. Neuroscientist 2001: 7: 480-489
- 28. Cheng EH, Kirsch DG, Clem RJ, Ravi R, Kastan MB, Bedi A et al. Conversion of Bcl-2 to a Bax-like death effector by caspases. Science 1997; 278: 1966-1968.
- 29. Almeida S, Sarmento-Ribeiro AB, Januário C, Rego AC, Oliveira CR. Evidence of apoptosis and mitochondrial abnormalities in peripheral blood cells of Huntington's disease patients. Biochem Biophys Res Commun 2008: 374: 599-603.
- 30. Hansson O, Castilho RF, Korhonen L, Lindholm D, Bates GP, Brundin P. Partial resistance to malonate-induced striatal cell death in transgenic mouse models of Huntington's disease is dependent on age and CAG repeat length. J Neurochem 2001; 78: 694-703.
- 31. Khoshnan A, Ko J, Tescu S, Brundin P, Patterson PH. IKKalpha and IKKbeta regulation of DNA damage-induced cleavage of huntingtin. PLoS One 2009; 4: e5768.
- 32. Happo L, Strasser A, Cory S. BH3-only proteins in apoptosis at a glance. J Cell Sci 2012; 125: 1081-1087.
- 33. Danial NN, Walensky LD, Zhang CY, Choi CS, Fisher JK, Molina AJ et al. Dual role of proapoptotic BAD in insulin secretion and beta cell survival. Nat Med 2008; 14: 144-153
- 34. Giménez-Cassina A, Martínez-François JR, Fisher JK, Szlyk B, Polak K, Wiwczar J et al. BAD-dependent regulation of fuel metabolism and K(ATP) channel activity confers resistance to epileptic seizures. Neuron 2012; 74: 719-730.
- 35. Rigamonti D, Bauer JH, De-Fraja C, Conti L, Sipione S, Sciorati C et al. Wild-type huntingtin protects from apoptosis upstream of caspase-3. J Neurosci 2000; 20: 3705-3713
- 36. Danial NN. BAD: undertaker by night, candyman by day. Oncogene 2008; 27(Suppl 1): S53-S70
- 37. Ley R, Hadfield K, Howes E, Cook SJ. Identification of a DEF-type docking domain for extracellular signal-regulated kinases 1/2 that directs phosphorylation and turnover of the BH3-only protein BimEL. J Biol Chem 2005: 280: 17657-17663.
- 38 O'Reilly LA, Cullen L, Visvader J, Lindeman GJ, Print C, Bath ML et al. The proapoptotic BH3-only protein bim is expressed in hematopoietic, epithelial, neuronal, and germ cells. Am J Pathol 2000; 157: 449-461.
- 39. Shibata M, Hattori H, Sasaki T, Gotoh J, Hamada J, Fukuuchi Y. Temporal profiles of the subcellular localization of Bim, a BH3-only protein, during middle cerebral artery occlusion in mice. J Cereb Blood Flow Metab 2002: 22: 810-820.
- 40. Leon R, Bhagavatula N, Ulukpo O, McCollum M, Wei J. BimEL as a possible molecular link between proteasome dysfunction and cell death induced by mutant huntingtin. Eur J Neurosci 2010; 31: 1915-1925.
- 41. King MA, Goemans CG, Hafiz F, Prehn JH, Wyttenbach A, Tolkovsky AM. Cytoplasmic inclusions of Htt exon1 containing an expanded polyglutamine tract suppress execution of apoptosis in sympathetic neurons. J Neurosci 2008; 28: 14401-14415.
- 42. Kong PJ, Kil MO, Lee H, Kim SS, Johnson GV, Chun W. Increased expression of Bim contributes to the potentiation of serum deprivation-induced apoptotic cell death in Huntington's disease knock-in striatal cell line. Neurol Res 2009: 31: 77-83.
- 43. Culmsee C, Zhu C, Landshamer S, Becattini B, Wagner E, Pellecchia M et al. Apoptosisinducing factor triggered by poly(ADP-ribose) polymerase and Bid mediates neuronal cell death after oxygen-glucose deprivation and focal cerebral ischemia. J Neurosci 2005; 25: 10262-10272
- 44. Engel T, Plesnila N, Prehn JH, Henshall DC. In vivo contributions of BH3-only proteins to neuronal death following seizures, ischemia, and traumatic brain injury. J Cereb Blood Flow Metab 2011: 31: 1196-1210.
- 45. Jhamandas JH, Mactavish DJ. β-Amyloid protein (Aβ) and human amylin regulation of apoptotic genes occurs through the amylin receptor. Apoptosis 2012; 17: 37-47
- 46. Yin XM. Bid, a BH3-only multi-functional molecule, is at the cross road of life and death. Gene 2006: 369: 7-19.
- 47. Chinnadurai G, Vijayalingam S, Gibson SB. BNIP3 subfamily BH3-only proteins: mitochondrial stress sensors in normal and pathological functions. Oncogene 2008; 27(Suppl 1): S114-S127.
- 48. Burton TR, Gibson SB. The role of Bcl-2 family member BNIP3 in cell death and disease: NIPping at the heels of cell death. Cell Death Differ 2009; 16: 515-523.
- 49. Zhang J, Ney PA. Mechanisms and biology of B-cell leukemia/lymphoma 2/adenovirus E1B interacting protein 3 and Nip-like protein X. Antioxid Redox Signal 2011; 14: 1959-1969
- 50. Kubli DA, Ycaza JE, Gustafsson AB. Bnip3 mediates mitochondrial dysfunction and cell death through Bax and Bak. Biochem J 2007; 405: 407-415.
- 51. Landes T, Emorine LJ, Courilleau D, Rojo M, Belenguer P, Arnauné-Pelloquin L. The BH3-only Bnip3 binds to the dynamin Opa1 to promote mitochondrial fragmentation and apoptosis by distinct mechanisms. EMBO Rep 2010; 11: 459-465.
- 52. Quinsay MN, Lee Y, Rikka S, Sayen MR, Molkentin JD, Gottlieb RA et al. Bnip3 mediates permeabilization of mitochondria and release of cytochrome c via a novel mechanism. J Mol Cell Cardiol 2010: 48: 1146-1156.

- 53. Vande Velde C, Cizeau J, Dubik D, Alimonti J, Brown T, Israels S et al. BNIP3 and genetic control of necrosis-like cell death through the mitochondrial permeability transition pore. Mol Cell Biol 2000: 20: 5454-5468
- 54. Cho B, Choi SY, Park OH, Sun W, Geum D. Differential expression of BNIP family members of BH3-only proteins during the development and after axotomy in the rat. Mol Cells 2012: 33: 605-610
- 55. Sassone J, Colciago C, Marchi P, Ascardi C, Alberti L, Di Pardo A et al. Mutant Huntingtin induces activation of the Bcl-2/adenovirus E1B 19-kDa interacting protein (BNip3). Cell Death Dis 2010: 1: e7.
- 56. Kubasiak LA, Hernandez OM, Bishopric NH, Webster KA. Hypoxia and acidosis activate cardiac myocyte death through the Bcl-2 family protein BNIP3. Proc Natl Acad Sci USA 2002 99 12825-12830
- 57. Hamacher-Brady A, Brady NR, Logue SE, Sayen MR, Jinno M, Kirshenbaum LA et al. Response to myocardial ischemia/reperfusion injury involves Bnip3 and autophagy. Cell Death Differ 2007; 14: 146-157.
- 58. Yu J, Zhang L, Hwang PM, Kinzler KW, Vogelstein B. PUMA induces the rapid apoptosis of colorectal cancer cells. Mol Cell 2001: 7: 673-682.
- 59. Nakano K, Vousden KH. PUMA, a novel proapoptotic gene, is induced by p53. Mol Cell 2001; 7: 683-694.
- 60. Engel T, Murphy BM, Hatazaki S, Jimenez-Mateos EM, Concannon CG, Woods I et al. Reduced hippocampal damage and epileptic seizures after status epilepticus in mice lacking proapoptotic Puma. FASEB J 2010; 24: 853-861.
- 61. Niizuma K, Endo H, Nito C, Myer DJ, Chan PH. Potential role of PUMA in delayed death of hippocampal CA1 neurons after transient global cerebral ischemia. Stroke 2009; 40: 618-625
- 62. Reimertz C. Kögel D. Rami A. Chittenden T. Prehn JH. Gene expression during ER stressinduced apoptosis in neurons: induction of the BH3-only protein Bbc3/PUMA and activation of the mitochondrial apoptosis pathway. J Cell Biol 2003; 162: 587-597.
- 63. Wei MC, Zong WX, Cheng EH, Lindsten T, Panoutsakopoulou V, Ross AJ et al. Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. Science 2001; 292: 727-730.
- 64. Hara A, Hirose Y, Wang A, Yoshimi N, Tanaka T, Mori H. Localization of Bax and Bcl-2 proteins, regulators of programmed cell death, in the human central nervous system. Virchows Arch 1996; 429: 249-253.
- 65. MacGibbon GA, Lawlor PA, Sirimanne ES, Walton MR, Connor B, Young D et al. Bax expression in mammalian neurons undergoing apoptosis, and in Alzheimer's disease hippocampus. Brain Res 1997; 750: 223-234.
- 66. Obonai T, Mizuguchi M, Takashima S. Developmental and aging changes of Bak expression in the human brain. Brain Res 1998; 783: 167-170.
- 67. Bae BI, Xu H, Igarashi S, Fujimuro M, Agrawal N, Taya Y et al. p53 mediates cellular dysfunction and behavioral abnormalities in Huntington's disease. Neuron 2005: 47: 29-41.
- 68. Sassone J, Colciago C, Cislaghi G, Silani V, Ciammola A. Huntington's disease: the current state of research with peripheral tissues. Exp Neurol 2009; 219: 385-397.
- 69. Puthalakath H, Huang DC, O'Reilly LA, King SM, Strasser A. The proapoptotic activity of the Bcl-2 family member Bim is regulated by interaction with the dynein motor complex. Mol Cell 1999; 3: 287-296.
- 70. Li Z, Zhang J, Liu Z, Woo CW, Thiele CJ. Downregulation of Bim by brain-derived neurotrophic factor activation of TrkB protects neuroblastoma cells from paclitaxel but not etoposide or cisplatin-induced cell death. Cell Death Differ 2007; 14: 318-326.
- 71. Almeida S, Laço M, Cunha-Oliveira T, Oliveira CR, Rego AC. BDNF regulates BIM expression levels in 3-nitropropionic acid-treated cortical neurons. Neurobiol Dis 2009; 35: 448-456.
- 72. Zuccato C. Ciammola A. Bigamonti D. Leavitt BR. Goffredo D. Conti L et al. Loss of huntingtin-mediated BDNF gene transcription in Huntington's disease. Science 2001: 293: 493-498
- 73. Zuccato C, Tartari M, Crotti A, Goffredo D, Valenza M, Conti L et al. Huntingtin interacts with REST/NRSF to modulate the transcription of NRSE-controlled neuronal genes. Nat Genet 2003: 35: 76-83
- 74. Puthalakath H, O'Reilly LA, Gunn P, Lee L, Kelly PN, Huntington ND et al. ER stress triggers apoptosis by activating BH3-only protein Bim. Cell 2007; 129: 1337-1349.
- 75. Duennwald ML, Lindquist S. Impaired ERAD and ER stress are early and specific events in polyglutamine toxicity. Genes Dev 2008: 22: 3308-3319.
- 76. Culmsee C, Plesnila N. Targeting Bid to prevent programmed cell death in neurons. Biochem Soc Trans 2006; 34: 1334-1340.
- 77. Eskes R, Desagher S, Antonsson B, Martinou JC. Bid induces the oligomerization and insertion of Bax into the outer mitochondrial membrane. Mol Cell Biol 2000; 20: 929-935
- 78. Sánchez I, Xu CJ, Juo P, Kakizaka A, Blenis J, Yuan J. Caspase-8 is required for cell death induced by expanded polyglutamine repeats. Neuron 1999; 22: 623-633.
- 79. Gervais FG, Singaraja R, Xanthoudakis S, Gutekunst CA, Leavitt BR, Metzler M et al. Recruitment and activation of caspase-8 by the Huntingtin-interacting protein Hip-1 and a novel partner Hippi. Nat Cell Biol 2002: 4: 95-105.
- 80. Gafni J, Ellerby LM. Calpain activation in Huntington's disease. J Neurosci 2002; 22: 4842-4849
- 81. Zhang J, Ney PA. Role of BNIP3 and NIX in cell death, autophagy, and mitophagy. Cell Death Differ 2009: 16: 939-946.

- Frazier DP, Wilson A, Graham RM, Thompson JW, Bishopric NH, Webster KA. Acidosis regulates the stability, hydrophobicity, and activity of the BH3-only protein Bnip3. *Antioxid Redox Signal* 2006; 8: 1625–1634.
- Graham RM, Frazier DP, Thompson JW, Haliko S, Li H, Wasserlauf BJ et al. A unique pathway of cardiac myocyte death caused by hypoxia-acidosis. J Exp Biol 2004; 207: 3189–3200.
- Graham RM, Thompson JW, Wei J, Bishopric NH, Webster KA. Regulation of Bnip3 death pathways by calcium, phosphorylation, and hypoxia-reoxygenation. *Antioxid Redox Signal* 2007; 9: 1309–1315.
- Dragunow M, Faull RL, Lawlor P, Beilharz EJ, Singleton K, Walker EB *et al.* In situ evidence for DNA fragmentation in Huntington's disease striatum and Alzheimer's disease temporal lobes. *Neuro Report* 1995; 6: 1053–1057.
- Thomas LB, Gates DJ, Richfield EK, O'Brien TF, Schweitzer JB, Steindler DA. DNA end labeling (TUNEL) in Huntington's disease and other neuropathological conditions. *Exp Neurol* 1995; 133: 265–272.
- Portera-Cailliau C, Hedreen JC, Price DL, Koliatsos VE. Evidence for apoptotic cell death in Huntington disease and excitotoxic animal models. J Neurosci 1995; 15: 3775–3787.
- Wang H, Lim PJ, Karbowski M, Monteiro MJ. Effects of overexpression of huntingtin proteins on mitochondrial integrity. *Hum Mol Genet* 2009; 18: 737–752.

- Costa V, Giacomello M, Hudec R, Lopreiato R, Ermak G, Lim D et al. Mitochondrial fission and cristae disruption increase the response of cell models of Huntington's disease to apoptotic stimuli. EMBO Mol Med 2010; 2: 490–503.
- Song W, Chen J, Petrilli A, Liot G, Klinglmayr E, Zhou Y et al. Mutant huntingtin binds the mitochondrial fission GTPase dynamin-related protein-1 and increases its enzymatic activity. Nat Med 2011; 17: 377–382.
- Kale J, Liu Q, Leber B, Andrews DW. Shedding light on apoptosis at subcellular membranes. *Cell* 2012; 151: 1179–1184.
- Davids MS, Letai A. Targeting the B-cell lymphoma/leukemia 2 family in cancer. J Clin Oncol 2012; 30: 3127–3135.

Cell Death and Disease is an open-access journal published by Nature Publishing Group. This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-sa/3.0/