

Ion channel inhibitors block caspase activation by mechanisms other than restoring intracellular potassium concentration

E Benítez-Rangel¹, L García¹, MC Namorado², JL Reyes² and A Guerrero-Hernández^{*1}

Ion fluxes at the plasma membrane have an important role in early stages of apoptosis. Accordingly, plasma membrane depolarization and gain of Na⁺ and loss of K⁺ are initial events in apoptosis. We have studied the effect of staurosporine (STS), a well-established apoptosis inducer, on the membrane potential of HeLa cells to determine the nature of STS-activated ion conductances and their role in the activation of different caspases. We observed that STS can activate tetraethylammonium (TEA⁺) and 4-aminopyridine-sensitive K⁺ channels and flufenamic-sensitive cation channels as an early response. The combination of these ion channel inhibitors significantly reduced cytochrome *c* (cyt *c*) release and activation of caspase-9, -3 and -8. STS also induced a large reduction in the intracellular [K⁺]_i that was not blocked by the ion channel inhibitors. Our data suggest that reduction in the [K⁺]_i is necessary but not sufficient and that ion channel inhibitors block activation of caspase-3 by two different mechanisms: the inhibitors of K⁺ channels by reducing cyt *c* release while flufenamic acid by a different, unrelated mechanism that does not involve cation channels at the plasma membrane. Our data also imply that these ion channels activated by STS are not responsible for the reduction in the [K⁺]_i associated with apoptosis.

Cell Death and Disease (2011) 2, e113; doi:10.1038/cddis.2010.93; published online 13 January 2011

Subject Category: Cancer

Cell transformations associated with apoptosis result from the biochemical action of an execution program, whose main characteristic is activation of caspases.¹ Different inducers of apoptosis trigger plasma membrane potential (PMP) depolarization² while the inhibition of apoptosis by Bcl-2 and Mcl-1 is associated with PMP hyperpolarization.^{3,4} It has been shown that ion fluxes, particularly K⁺ efflux, have a key role in apoptosis. The activation of both K⁺^{5,6} and Cl⁻ channels is necessary for apoptotic volume decrease (AVD) or cell shrinkage and also for activation of caspases.^{7,8} It has been shown that, before AVD, there is an initial movement of monovalent ions. Although the inhibition of Cl⁻ channels while inhibiting AVD, does not always reduce activation of caspases.⁹ Different inducers of apoptosis trigger both accumulation of intracellular Na⁺ and loss of intracellular K⁺^{2,7,10–13} and these events are associated with PMP depolarization.² It has been also shown that the reduction in the intracellular [K⁺]_i and PMP depolarization are a late event since involve inhibition of Na⁺/K⁺ pump by caspase-mediated degradation of its β-subunit.^{2,14}

Nevertheless, the activation of caspases requires the reduction of the [K⁺]_i regardless the inducer of apoptosis used or cell type studied.^{13,15–17} High external [K⁺]_o inhibits both the extrinsic (Fas-mediated) and the intrinsic (etoposide-induced) cell death programs apparently by inhibiting the

release of cytochrome *c* (cyt *c*) from mitochondria.¹⁷ However, it has been shown that staurosporine (STS)-induced cyt *c* release in both HeLa and neuroblastoma cells (SK-N-BE(2)) is not inhibited by avoiding reduction of [K⁺]_i.¹⁶ Actually, it appears that high intracellular K⁺ protects against apoptosis by inhibiting the apoptosome assembly.^{13,16,18} Apparently, the procaspase-3 activity is inhibited by high [K⁺]_i because its activity decrease to ~50% in [K⁺]_i above 25 mM K, in contrast mature caspase-3 activity is unaltered by reducing [K⁺]_i.¹⁸ Recently, it has been suggested that the apoptosome assembly is regulated by ion strength more than a direct effect of K⁺ *per se*.^{15,16}

STS, a non-selective kinase inhibitor that has a strong apoptotic action, activates K⁺ currents⁵ and also induces PMP depolarization, which has been reported to depend on both depolarization of mitochondrial membrane potential (MMP) and activation of caspase-3 (Bortner *et al.*²; Dussmann *et al.*¹⁴). We have studied the effect of STS on PMP and ion channels inhibitors on the activity of caspases in HeLa cells. Our data suggest that STS activates both K⁺ and cation channels and that [K⁺]_i reduction is required for caspase activation, particularly caspase-3. Ion channel inhibitors while blocking activation of caspase-3 do not block STS-induced reduction in [K⁺]_i, implying that STS-induced K⁺ loss is not occurring by these ion channels activated by STS.

¹Department of Biochemistry, CINVESTAV-IPN, DF, México and ²Department of Physiology, Biophysics and Neuroscience, CINVESTAV-IPN, DF, México

*Corresponding author: A Guerrero-Hernández, Departamento de Bioquímica, CINVESTAV-IPN, Apdo. Postal, 14-740, DF, México. Tel: + 52 55 50613950; Fax: + 52 55 50613391; E-mail: aguerrero@cinvestav.mx

Keywords: plasma membrane potential; potassium channels; caspases; flufenamic; cytochrome *c* release; apoptosis

Abbreviations: AVD, apoptotic volume decrease; COX, cyclo oxygenase; cyt *c*, cytochrome *c*; FA or F, flufenamic acid; Gram, gramicidin; TEA⁺ or T, tetraethylammonium; 4AP or 4, 4-aminopyridine; PMP, plasma membrane potential; STS, staurosporine

Received 29.9.10; revised 22.11.10; accepted 29.11.10; Edited by V De Laurenzi

Results

High external K^+ inhibits STS-induced activation of caspases. Caspases are the main effectors in apoptosis, particularly caspase-3. Incubation of cells with $1 \mu\text{M}$ STS for a period of 4 h induced a significant activation of caspases-9, -3 and -8 in HeLa cells (Figure 1). This time period of 4 h was chosen because STS induced the maximal activation of caspase-3 and cyt *c* release (Supplementary Figure 3). STS-induced caspase-3 activation was much larger when compared with other apoptosis inducers, such as H_2O_2 and thapsigargin (not shown). Under our assay conditions (cells were in serum-free culture medium for 24 h) both caspase-9 and caspase-8 displayed a larger basal activity than caspase-3 when compared with the corresponding maximal response obtained with STS. Interestingly, STS induced a significant activation of caspase-8, the main effector of the extrinsic pathway in apoptosis. Caspase-8 can be activated by caspase-3 (Tang *et al.*¹⁹) but also the reverse has been described.²⁰ Incubation of cells for 4.5 h in 5, 70 or 140 K saline solutions (see Materials and Methods section) did not change the basal activity of these three caspases (not shown). However, STS-induced activation of all three caspases was significantly inhibited by 70 K solution in approximately 60% (Figure 1). Interestingly, 140 K solution totally abolished the effect of STS on all three caspases (Figure 1). These data confirm the importance of K^+ reduction in the activation of these three caspases. We next studied whether STS could activate ion channels that would function as the exit pathway for K^+ ions.

STS induces an early activation of ion channels at the plasma membrane. To study the ion conductance activated by STS, we incubated HeLa cells with bisoxonol. For all and each experiment, we determined first the resting PMP by

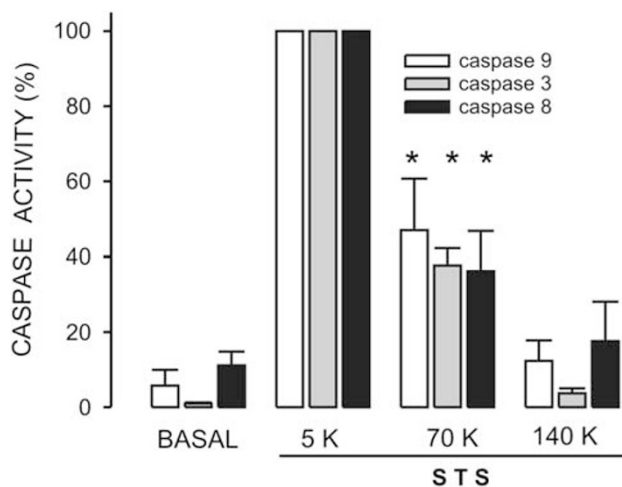


Figure 1 High extracellular $[K^+]$ inhibits activation of caspases by STS in HeLa cells. Caspase-9, -3 and -8 activities were assessed as indicated in Materials and Methods section. The caspase inhibitor-corrected fluorescence signal induced by STS was taken as 100% for each caspase. STS activated caspase-9, -8 and -3 about 20-, 10- and 100-fold, respectively. This activation was significantly inhibited by preincubating cells for 30 min in 70 or 140 K solutions. $n = 5$. $*P < 0.05$

increasing the activity of K^+ channels with histamine (Supplementary Figure 2). This agonist via H_1 receptors induces the increase in inositol trisphosphate levels, Ca^{2+} release and the activation of Ca^{2+} -dependent intermediate conductance K^+ channels.²¹ This maneuver sets the PMP at the reversal potential for K^+ channels and we used this gradient to determine resting PMP. As histamine-induced K^+ conductance becomes the predominant one, we can use Nernst equation to calculate resting PMP. HeLa cells had an average resting PMP of $-60 \pm 1.1 \text{ mV}$ ($n = 76$), these data were close to the resting PMP ($-51 \pm 2.4 \text{ mV}$) observed using patch clamp technique.^{22,23} The similar values for PMPs obtained with these two different approaches support the idea that bisoxonol is a reliable indicator to estimate PMP changes in HeLa cells during apoptosis.

It has been reported that STS induces an elevation of K^+ channel activity in PASM cells⁵ and in other types of cells.²⁴ To determine whether STS activates ion channels in HeLa cells, we have studied the effect of this apoptosis inducer on PMP within the first 10 min of STS addition (Figure 2). This time window was selected because it allowed the stabilization of STS-induced PMP changes. Although, STS always changed PMP in few minutes, the type and magnitude of the responses were variable. Most of the experiments (60%) displayed STS-induced depolarization of PMP (Figure 2a, dashed line) while the rest showed plasma membrane hyperpolarization that was either sustained (Figure 2a, dotted line) or transient (Figure 2a, solid line). This hyperpolarization increased PMP by approximately 20 mV ($-79.7 \pm 2.9 \text{ mV}$, $n = 28$, Figure 2c, fourth open bar) and the depolarization response to STS involved a 20 mV decreased in PMP ($-39.4 \pm 1.7 \text{ mV}$, $n = 47$, Figure 2c, fourth closed bar). The application of DMSO alone did not modify PMP ($-53.5 \pm 3 \text{ mV}$, $n = 13$, not shown). These data show two different types of PMP responses to STS that we called D or H when at the end of the 10-min recording period; the PMP was either above or below the resting membrane potential, respectively. However, the resting PMP was practically the same between cells having the H or D responses (Figure 2c, first bars). PMP for cells having D response was $-59.2 \pm 1.3 \text{ mV}$ ($n = 47$, closed bar) and for cells showing H response was $-61 \pm 2 \text{ mV}$ ($n = 28$, open bar). To assess whether the effect of STS was due to the increase in ion channel activity, we determined the effect of STS on the capability of gramicidin (Gram) to depolarize HeLa cells (Figures 2a and b). STS reduced the depolarization effect of Gram ($2.1 \pm 0.4 \text{ mV}$, $n = 28$) for either D ($-5.4 \pm 1.5 \text{ mV}$, $n = 29$) or H ($-24 \pm 9.2 \text{ mV}$, $n = 11$) type of responses, implying that STS is activating ion channels. Collectively, these data suggest that STS activates different types of ion channels as an early response in HeLa cells.

It has been reported that PMP depolarization is associated with apoptosis.^{2,14} However, it has been suggested that this PMP depolarization occurs after activation of caspases¹⁴ and is partly due to Na^+/K^+ ATPase degradation.^{2,14} To determine whether caspases were responsible for this effect on PMP, HeLa cells were incubated with $50 \mu\text{M}$ of Z-VAD-fmk. This pan-caspase inhibitor did not reduce STS-induced depolarization response ($-24.9 \pm 2.8 \text{ mV}$, $n = 15$). Additionally, we studied whether inhibition of Na^+/K^+ ATPase could

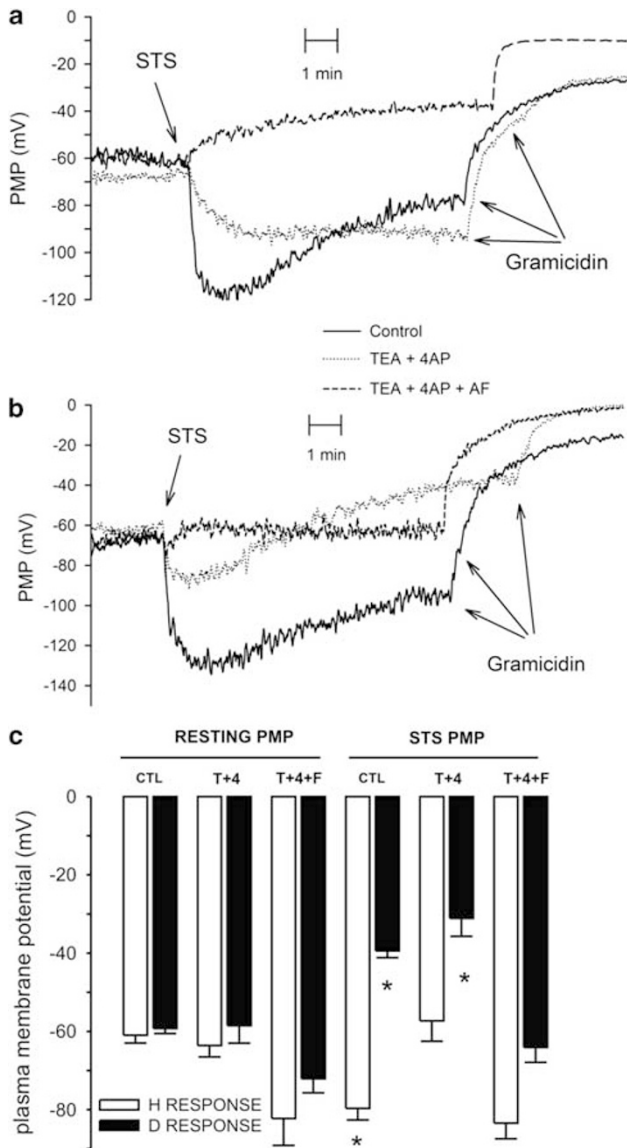


Figure 2 STS alters PMP as an early response in HeLa cells. **(a)** Cells loaded with bisoxonol to follow PMP were exposed to 1 μ M STS at the time indicated inducing membrane depolarization (dashed line, $n=48$), hyperpolarization (dotted line) or a mixed response (solid line), $n=28$ for both conditions. Once the fluorescence signal has stabilized, Gram was added as depolarization control. Depending whether the PMP finished above or below resting PMP (i.e., PMP before application of STS) were defined either depolarization (D) or hyperpolarization (H), respectively. These records show PMP responses of cells from three different passages, we do not know why the ion channels responding to STS vary among different passages. **(b)** Cells incubated in either TEA⁺ and 4AP or the combination of TEA⁺, 4AP and FA were exposed to STS were indicated. A mixed response (solid line, $n=26$) became depolarizing response by the addition of TEA⁺ and 4AP to inhibit K⁺ channels (dotted line, $n=12$). The combination of all three ion channels inhibitors blocked completely the STS-induced changes in PMP (dashed line, $n=14$). **(c)** Average resting PMP and the effect of ion channels inhibitors for cells that respond to STS with either hyperpolarization (white bars) or depolarization (dark bars) depicted as CTL. TEA⁺ and 4AP did not affect resting PMP but inhibited the STS-induced hyperpolarization, particularly for H responses and favored D responses. Notice that the combination of TEA⁺, 4AP and FA-induced hyperpolarization of the resting PMP and totally abolished the STS-induced PMP changes. In this figure, * $P<0.05$ with respect to the corresponding resting PMP. From left to right $n=26, 47, 8, 12, 3, 9, 28, 48, 8, 12, 3$ and 14 respectively

explain the STS-induced depolarization. Incubating cells with 100 μ M ouabain, a Na⁺/K⁺ ATPase inhibitor, produced PMP depolarization in few minutes (-28.8 ± 3.1 mV, $n=10$), in this case Gram-induced membrane depolarization (1.9 ± 0.4 mV, $n=8$), which was similar to the one obtained in cells without STS (2.1 ± 0.4 mV, $n=28$), implying that ouabain-induced PMP depolarization was not because of the opening of ion channels but the inhibition of Na⁺/K⁺ pump. The application of STS after ouabain-induced membrane depolarization still produced either depolarization or hyperpolarization of PMP because it was -20.5 ± 3.1 mV ($n=8$) or -59.1 ± 8.1 mV ($n=12$), respectively. All these data indicate that the effect of STS on PMP cannot be explained by inhibition of Na⁺/K⁺ ATPase at this time window.

It has been shown that STS induces an increase in the activity of plasma membrane K⁺ channels, which are sensitive to tetraethylammonium (TEA⁺) and 4-aminopyridine (4AP).²⁴ To test whether this was the case in HeLa cells, we studied the effect of these inhibitors on the STS-induced changes in PMP (Figure 2b, dotted line). These K⁺ channel inhibitors (T+4) did not modify resting PMP (Figure 2c, second bars). However, The STS-induced hyperpolarization was strongly inhibited by the combination of TEA⁺ and 4AP because PMP in the presence of STS (-57.3 ± 5.2 mV, $n=8$) was practically the same as the resting PMP in the presence of these inhibitors (-63.6 ± 2.9 mV, $n=8$; Figure 2b, dotted line and Figure 2c, sixth open bar). In the case of cells showing D responses, these ion channels inhibitors made STS-induced membrane depolarization slightly larger -31.1 ± 4.6 mV ($n=12$, Figure 2c, sixth closed bar) from a resting PMP of -58.5 ± 4.5 mV ($n=12$). These data imply that STS activates K⁺ channels, mainly in cells displaying the H type of response, while cells with D type of response appear to have a poor activity of STS-induced K⁺ channels. These data suggest that STS activates cation channels in those cells having depolarization responses. To study this situation, we used low concentration (50 μ M) of flufenamic acid (FA or F) to inhibit cation channels. FA increased the resting PMP, reaching the value of -72.1 ± 3.6 mV, ($n=7$, Figure 2c, third closed bar). The last result implies that FA had a hyperpolarization effect on PMP and this effect could be explained by inhibition of cation channels, which are open under resting conditions. The combination of FA and K⁺ channel inhibitors strongly reduced the STS-induced depolarization of PMP (Figure 2b dashed line and Figure 2c, sixth solid bar). All these data suggest that STS activates, as an early response, both K⁺ and cation channels in HeLa cells. We next studied whether these channels have any role in the activation of caspases in these cells.

Role of ion channels in the intrinsic pathway of apoptosis. It has been proposed that STS induces apoptosis by activation of the intrinsic pathway.^{14,25,26} HeLa cells were incubated with z-VAD-fmk (10 or 50 μ M) for 30 min before induction of apoptosis by 4-h incubation with 1 μ M STS. Cyt *c* was released to the cytoplasm in response to STS by a mechanism that does not involve the activation of caspases (Figure 3a). We also studied the role of external [K⁺] on STS-induced cyt *c* release by incubating

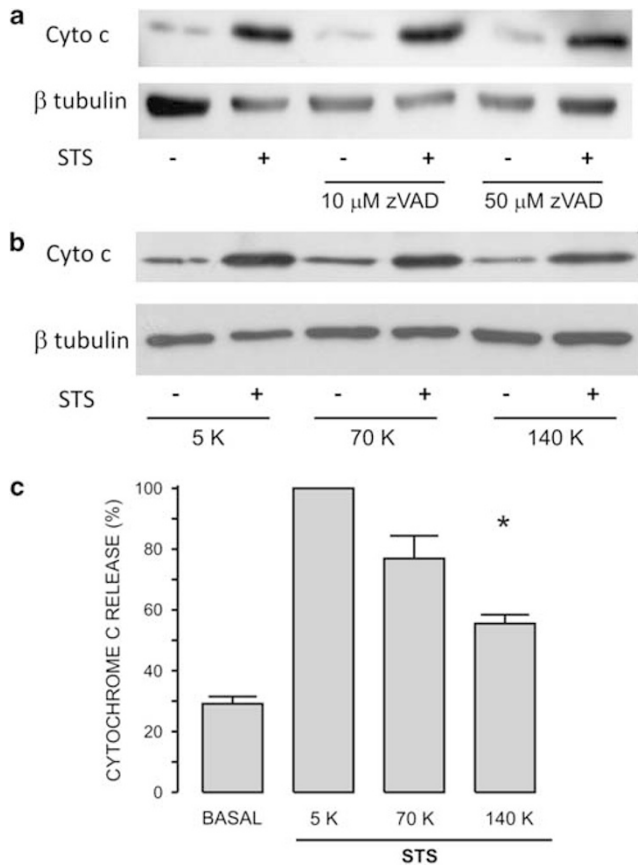


Figure 3 High external [K⁺] reduces STS-induced cyto c release. (a) Incubation of cells with either 10 or 50 μ M z-VAD-fmk, a pan-caspase inhibitor, did not block the STS-induced cyto c release ($n=4$). (b) HeLa cells preincubated in either culture medium (5 K) or 70 or 140 K solutions were challenged with STS for 4 h to assess cyto c release by western blot assay and using β -tubulin as loading control. Notice that the different external [K⁺] did not change the basal release of cyto c. (c) The basal level of cyto c was high because of the absence of serum for 24 h (see Supplementary Figure 1). However, the STS-induced cyto c release was significantly reduced only by 140 K ($n=4$). * $P<0.05$

cells in either 70 or 140 K solutions (Figure 3b). The addition of STS to cells in 70 K solution did not inhibit cyto c release (Figure 3c). However, STS-induced cyto c release was significantly reduced when cells were incubated in 140 K solution (Figures 3b and c). Preincubation of HeLa cells with the combination of ion channel inhibitors for 30 min reduced STS-induced cyto c release (Figure 4a). This effect was only significant for K⁺ channels inhibitors alone or in combination with FA (Figure 4b). FA alone did not have any effect on STS-induced cyto c release. These data suggest that only K⁺ channels have a role, still a limited one, in the STS-induced cyto c release.

Role of ion channels in the activation of caspases. Caspase-9 is activated as a consequence of cyto c-mediated apoptosome assembly.²⁷ Interestingly, caspase-9 activity was reduced ~40% by the combination of K⁺ channel inhibitors and FA (Figure 5, open bars). These data are similar to the effect of these inhibitors on the release of cyto c, this is in agreement with the importance of cyto c for the

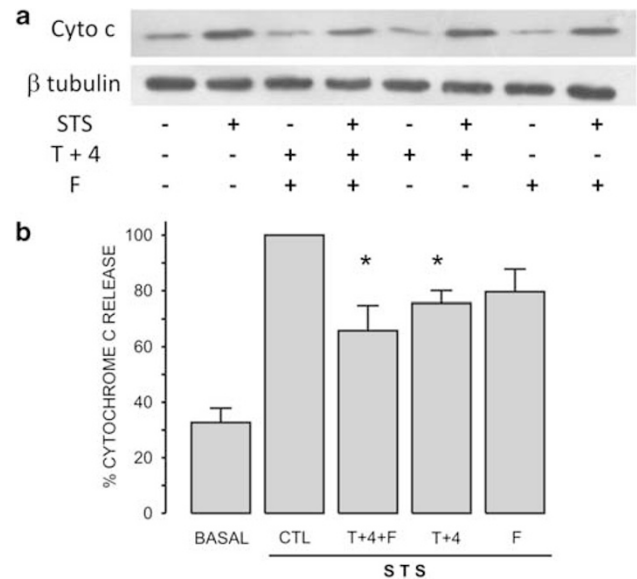


Figure 4 Ion channel inhibitors partially reduced cyto c release. (a) The presence of cyto c in the cytosol was detected by western blot assay. The optical density ratio (cyto c/ β -tubulin) was normalized considering the effect of STS as 100%. (b) FA did not have any effect on STS-induced cyto c release, but the combination of K⁺ channel inhibitors (T + 4) reduced significantly the STS-induced cyto c release, while the addition of FA did not increase any further the inhibitory effect of the combination of K⁺ channel inhibitors ($n=15$). * $P<0.05$

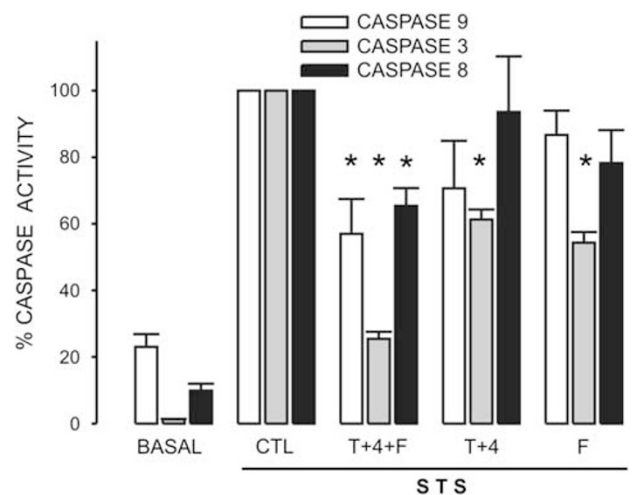


Figure 5 Ion channel inhibitors block caspase activation by different mechanisms. Activities of caspase-9 ($n=15$), -3 ($n=18$) and -8 ($n=7$) induced by STS were assessed in the presence of different ion channel inhibitors. Neither caspase-9 nor caspase-8 were affected by either the combination of TEA⁺ and 4AP (T + 4) or FA. However, caspase-3 was significantly inhibited by both types of ion channel inhibitors separately or in combination. This combination had an additive effect suggesting that TEA⁺ and 4AP had a different mechanism to inhibit caspase-3 than FA. Interestingly, only this combination of ion channel inhibitors reduced significantly the activation of caspase-9 and -8. * $P<0.05$

assembly of the apoptosome, which in turn activates caspase-9.

Caspase-3 is the main executioner downstream caspase-9. We have studied the role of ion channels in STS-induced activation of caspase-3 by two different approaches; one was

protease activity while the other was PARP degradation. In the case of caspase-3 activity, the ion channels inhibitors separately reduced STS-triggered caspase-3 activity by ~40% each one (Figure 5, gray bars). This is completely different to the effect of ion channel inhibitors on caspase-9 activity. Moreover, the combination of these ion channels inhibitors significantly reduced by ~80% STS-induced caspase-3 activity. Importantly, the effect of the combination of ion channel inhibitors is additive, suggesting that these two types of ion channel inhibitors work independently of each other to avoid reduction in the intracellular $[K^+]$. These data suggest that cation channels might be also a pathway for K^+ exit from the cell in STS-induced apoptosis, as relevant as K^+ channels themselves (Figure 5, gray bars).

Additionally, we also studied the participation of ion channels in STS-induced activation of caspase-8. In this case ion channels inhibitors did not have any consistent effect, the variability was so high that there is no significant difference with respect to the basal activity. The combination of these ion channel inhibitors produced a partial but significant reduction (~40%) in the STS-induced caspase-8 activity (Figure 5, closed bars).

Role of ion channels in degradation of PARP a caspase-3 substrate. The enzyme PARP is one of the proteins cleaved by caspase-3 (Soldani and Scovassi²⁸). Accordingly, 4-h incubation with STS displayed complete degradation of cytoplasmic PARP (Figure 6a), which was significantly reduced only by the combination of ion channel inhibitors

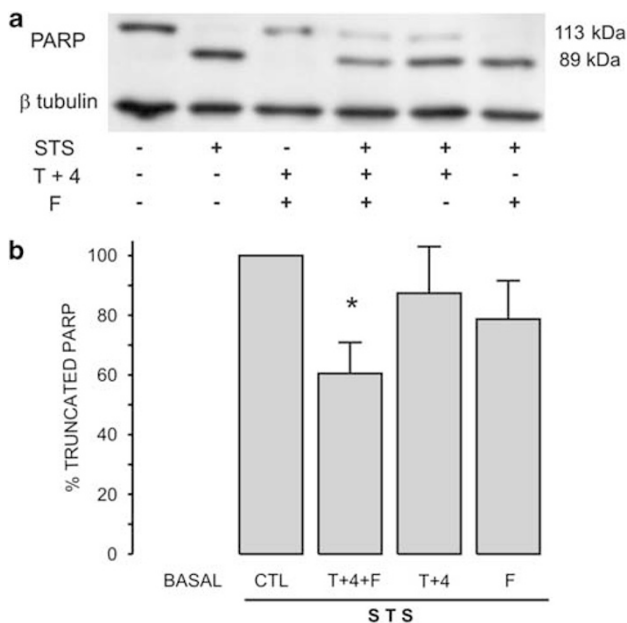


Figure 6 The combination of ion channel inhibitors reduced PARP degradation. (a) Supernatant cell extracts were used to assess PARP degradation by western blot and the optical density ratio between the fragment detected at 89 kDa and β -tubulin resulting of incubating cells with STS was considered as 100% response. Notice that in control cells truncated PARP is absent while STS-induced PARP degradation completely. (b) None of the ion channels inhibitors alone decreased the normalized level of truncated PARP, but this level was significantly decreased by the combination of TEA⁺, 4AP and FA (T + 4 + F, $n = 9$). * $P < 0.05$

(Figure 6b). Both approaches, that is, caspase-3 activity and PARP degradation showed that the combination of the two types of ion channel inhibitors had a larger effect than when they were applied separately. Data shown in Figures 5 and 6 argue for PARP being highly sensitive to caspase-3 activity because only 25% of caspase-3 activity (Figure 5, gray bars) induces practically 60% degradation of PARP (Figure 6b).

Intracellular $[K^+]$ and $[Na^+]$ changes induced by STS as detected by atomic absorption spectrometry.

Collectively, these data suggest that these ion channel inhibitors reduce K^+ loss associated with apoptosis. To corroborate this hypothesis, we determined the effect of STS and the combination of these ion channel inhibitors on the intracellular $[K^+]$ and $[Na^+]$ after 4-h incubation using atomic absorption spectrometry. STS-induced a significant decrease in the $[K^+]_i$ from 140 mM to 62.6 ± 4.3 mM ($n = 4$, Figure 7), while the increase in the $[Na^+]_i$ went from 30 mM to 42.1 ± 7.5 mM ($n = 4$), which was not large enough to be significant. Unexpectedly, neither the combination of TEA⁺ and 4AP nor flufenamic alone reduced the K^+ loss triggered by STS. The combination of these ion channel inhibitors showed a slight recovery of $[K^+]_i$, which was too small to reach significance (Figure 7). These data corroborate that apoptosis is associated with a reduced $[K^+]_i$; however, suggest that ion channels inhibitors block caspase-3 activation by other mechanisms that do not involve the reduction in K^+ loss.

Discussion

Apoptosis is clearly associated with PMP depolarization,^{2,14,29} while inhibition of apoptosis by Bcl-2 and Mcl-1 is associated with PMP hyperpolarization.^{3,4} The role of K^+ channels in apoptosis has been studied extensively because it

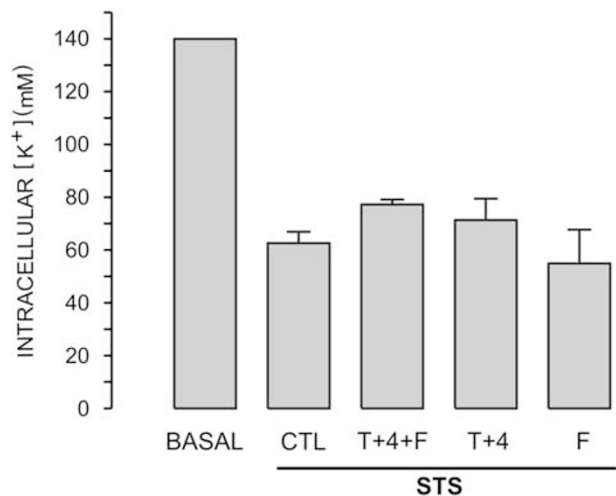


Figure 7 Ion channel inhibitors did not decrease STS-induced intracellular $[K^+]$ reduction. $[K^+]$ detected with atomic absorption spectrometry and normalized by protein was taken to represent 140 mM in the basal condition. STS in a period of 4 h produced a large and reproducible decrease in the intracellular $[K^+]_i$ (CTL response), which was not reversed by the preincubation with either the ion channel inhibitors alone or their combination ($n = 4$)

is thought to be a pathway for K^+ exit from the cytoplasm,^{5,6,24,29,30} which is a necessary step for the activation of caspases.^{5,13,18} We have studied the PMP changes induced by STS, a strong activator of apoptosis in HeLa cells. To this end we have used bisoxonol, a fluorescent lipophilic anion, which reports mainly PMP with little interference from the mitochondrial membrane potential and because it has a large ratio of $\% \Delta F/mV$.² This approach, particularly at low concentrations, does not modify plasma membrane resistance nor induces cell dialysis, conditions associated with single-cell patch clamp electrophysiology. One caveat of this approach is that PMP values represent the average response of 7.5×10^5 cells in the cuvette. Nevertheless, under our recording conditions the calculated resting PMP was similar to the one reported for HeLa cells using the patch clamp technique.^{22,23} We think that the use of sub- μM bisoxonol has allowed us to detect small changes in the activity of ion channels as an initial response to the application of STS. Importantly, plasma membrane depolarization can be originated by activation of ion channels (either cation or anion) or by inhibition of either K^+ channel or Na^+/K^+ pump, to determine which of these two conditions were occurring we used Gram, as a depolarization control. Our data indicate that STS activates both K^+ and cation channels in few minutes. The former were sensitive to the combination of TEA^+ -4AP while the latter to FA, based on the effect of these ion channel inhibitors on the STS-induced changes in PMP.

We have corroborated that reducing the driving force for K^+ efflux by increasing external $[K^+]_o$ inhibits activation by STS of caspase-9, -3 and -8 in HeLa cells. These data highlight the importance of K^+ efflux mechanisms in apoptosis, particularly for caspase activation because the ionic strength was kept constant for 5, 70 and 140 K solutions.^{5,13,18} Actually, it has been shown in different cell types that STS activates delayed rectifier K^+ channels of K_{v1} and K_{v3} subfamilies, which are sensitive to 4AP and TEA^+ , respectively.³⁰ In this regard, our data show that the combination of TEA^+ and 4AP inhibited the STS-induced hyperpolarization response. Nevertheless, our data suggest that the combination of K^+ channel inhibitors block caspase activation by reducing the STS-induced cyt *c* release more than inhibiting the loss of $[K^+]_i$. Accordingly, the 140 K solution inhibited to a similar extent than K^+ channel inhibitors the STS-induced cyt *c* release. Importantly, we did not find any direct effect of these ion channels inhibitors when assessed on previously activated caspase-3 (data not shown).

Flufenamic acid-induced plasma membrane hyperpolarization and totally abolished the activation by STS of the depolarization conductance. FA did not reduce the STS-induced cyt *c* release and affected neither caspase-9 nor -8 activities. Nevertheless, FA at low concentrations significantly reduced the STS-induced caspase-3 activity by a mechanism that is independent of the one blocked by the combination of TEA^+ and 4AP. FA has different targets within cells, it is an inhibitor of cyclo oxygenase (COX) 1 and 2 (Lovering *et al.*³¹); of AKR1C3 (Lovering *et al.*³¹) and also of chloride³² and cation³³ channels. None of these targets appears to explain the effect of FA on caspase-3. High doses of nonsteroidal antiinflammatory drugs induce apoptosis by inhibiting COX enzymes and triggering ceramide production.³⁴ Moreover, FA

would increase the level of prostaglandin 15d-PGJ₂, an apoptosis inducer, by inhibiting the enzyme AKR1C3 that transform PGD₂ into PGF_{2 α} , the former being the precursor of 15d-PGJ₂ (Lovering *et al.*³¹). Ca^{2+} -dependent Cl^- channels are sensitive to this low concentration of FA.³² However, removing external $[Ca^{2+}]_o$ or chelating $[Ca^{2+}]_i$ did not affect the STS-induced caspase-3 activation (not shown), suggesting that Ca^{2+} -dependent Cl^- channels are not involved. Moreover, the inhibition of Cl^- channels is not additive with respect to the inhibition of K^+ channels.⁸ Additionally, blockers of Cl^- channels that inhibit AVD, does not always inhibit the activation of caspase-3 (Wei *et al.*⁹). We think that FA is blocking cation channels because it hyperpolarized PMP and induced a transient reduction in the $[Ca^{2+}]_i$ (not shown) that reflects the inhibition of Ca^{2+} -permeable cation channels that are active at resting PMP. Nevertheless, these cation channels activated by STS do not represent a substantial K^+ exit pathway because FA did not inhibit the STS-induced loss of $[K^+]_i$. Moreover, our atomic absorption spectrometry data did not show a consistent and substantial increase in the $[Na^+]_i$ in response to STS, most likely because the Na^+ pump was not yet compromised at 4 h with STS. Our data also argue that STS-induced K^+ reduction occurs via either K^+ channels insensitive to TEA^+ and 4AP or most likely via non-electrogenic mechanism that do not involve K^+ channels.

In conclusion, K^+ channel inhibitors blocked caspase-3 activation by inhibiting cyt *c* release while FA uses a different mechanism. These data highlight the presence of different and multiple mechanisms to control activation of caspases.

Materials and Methods

Materials. Cell culture reagents were obtained from Gibco-Invitrogen (Grand Island, NY, USA). STS, histamine, TEA^+ or T, 4AP or 4, FA or F, ouabain, Gram, Ac-DEVD-AMC (caspase-3 substrate), Ac-DEVD-CHO (caspase-3 inhibitor), Ac-LEDH-AFC (caspase-9 substrate), Ac-LEDH-CHO (caspase-9 inhibitors), Ac-IETD-CHO (caspase-8 inhibitor) and protease inhibitor cocktail were purchased from Sigma-Aldrich (St. Louis, MO, USA). Ac-IETD-AMC (caspase-8 substrate) was acquired from Enzo Life Sciences (Lausen, Switzerland). DiBaC₄(3) (bisoxonol) was obtained from Invitrogen (Molecular Probes, Eugene, OR, USA). z-VAD-fmk was bought from Promega (Madison, WI, USA). Mouse anti-cyt *c* monoclonal antibody (clone 7H8.2C12) and mouse anti-human PARP (clone 4C10-5) were purchased from BD-Pharmingen (Chicago, IL, USA). Mouse anti- β -tubulin was acquired from ZYMED (Carlsbad, CA, USA).

Cell culture. HeLa cells were cultured in Dulbecco's modified Eagle's medium (with high D-glucose (4500 mg/l), L-glutamine and sodium pyruvate (110 mg/l)) supplemented with 5% fetal bovine serum, 3% newborn calf serum and penicillin-streptomycin. Cells were cultured for 24 h without serum before inducing apoptosis by the addition of 1 μM STS for a period of 4 h. The absence of serum increased for $\sim 30\%$ the release of cyt *c* to the cytoplasm with respect to the effect of STS (Supplementary Figure 1). Nevertheless, cell viability was always $\geq 95\%$ (0.08% Trypan blue exclusion).

PMP determination with bisoxonol. HeLa cells that have been cultured in the absence of serum for 24 h were harvested, washed and the cell pellet resuspended in saline solution containing: 121 mM NaCl, 5.4 mM KCl, 0.8 mM MgCl₂, 1.8 mM CaCl₂, 6 mM NaHCO₃ and 25 mM Hepes, at pH 7.3 (NaOH)³⁵ at a density of 3×10^6 cells/ml. To determine PMP, 125 nM bisoxonol was added to the saline solution followed by 3×10^5 cells/ml and fluorescence signal was excited at 490 nm and collected at 530 nm using a QM-8 spectrofluorometer (PTI). Once the fluorescence signal has leveled off, K^+ conductance was increased with 100 μM histamine, which induces plasma membrane hyperpolarization (Supplementary Figure 2) and the K^+ equilibrium potential was changed by applying five consecutive additions of 25 mM KCl (Supplementary Figure 2). Finally, 1 μM Gram

was added as depolarization control. As histamine increased the K^+ conductance, we can use Nernst equation to calculate PMP as the equilibrium potential at different external $[K^+]$. These data were fitted by the following equation $\%F = k \log_{10} (E_r / (E_m - h))$ where $\%F$ is $\%(\Delta F / F_0)$, F_0 is fluorescence signal at rest, $k = \text{slope}$, $h = \text{constant}$, $E_r = \text{PMP when } \%F \text{ is zero}$, $E_m = \text{PMP for different } [K^+]$. From the fitting of these data to the equation, we derived the values for k , E_r and h for each experiment (passage of cells) and used these values to calculate PMP from the value of E_m using the following relationship $E_m = [E_r / 10^{(\%F/k)}] + h$. To determine the effect of STS on the PMP, bisoxonol was added to saline solution alone or with ion channel inhibitors (either the combination of 10 mM TEA⁺ and 2 mM 4AP or 50 μ M FA or all three together) followed by cell addition. In the case of the pan-caspase inhibitor (z-VAD-fmk), this chemical was incubated with cells for 30 min before initiating PMP recording. Once the fluorescence signal leveled off after cell addition, 1 μ M STS was added and the bisoxonol fluorescence signal was recorded for 5–10 min before the application of 1 μ M Gram as depolarization control.

Induction of apoptosis for cells in culture. Confluent HeLa cells that were cultured without serum for 24 h were preincubated for 30 min with or without the combination of K^+ channel inhibitors, 50 μ M FA, all three ion channel inhibitors, 10 or 50 μ M z-VAD-fmk before STS. For experiments with different external $[K^+]$ cells were left for 30 min in culture medium (5 K), in 70 K saline solution containing in mM: 25 Hepes, 70 KCl, 51.5 NaCl, 0.8 MgCl₂, 1.8 CaCl₂, 6 NaHCO₃, 5.5 glucose at pH 7.3 and in 140 K saline solution containing in mM: 25 Hepes, 126.5 KCl, 0.8 MgCl₂, 1.8 CaCl₂, 6 NaHCO₃, 5.5 glucose at pH 7.3 before inducing apoptosis with STS. After 4-h incubation with STS, cell were lysed and lysates were used for caspase assays, western blot assay and determination of intracellular $[K^+]$ and $[Na^+]$ with atomic absorption spectrometry.

Detection of caspase activity. In all caspase assays, cells cultured in six-well plates were lysed with 500 μ l of caspase lysis buffer per well containing in mM: 50 Hepes, 5 DTT and 1% Triton X-100 and well plate were stirred on ice for 30 min before collecting the samples.

Caspase-3 activity. Assay was carried out according to the specifications in the kit with slight variations. Caspase-3 assay buffer contained in mM: 20 Hepes, 5 DTT, 2 EDTA and 0.1% Triton X-100 at pH 7.4. Assay buffer was supplemented with 1 μ M Ac-DEVD-AMC (caspase-3 substrate) with or without 2 μ M Ac-DEVD-CHO (caspase-3 inhibitor) and 50 μ l of cell lysate. This mix was incubated at room temperature for 90 min. Caspase-3 activity was determined subtracting to the fluorescence signal to the one obtained in the presence of caspase inhibitor. The fluorescence value obtained from cells incubated with STS was considered as 100% response.

Caspase-9 activity. Assay was carried out as previously described⁹⁶ with slight differences. Caspase-9 assay buffer contained in mM: 100 MOPS, 10 DTT, 0.5 EDTA, 0.1% Triton X-100 at pH 6.5 and 10% glycerol. Assay buffer with 100 ml of cell lysate was preincubated for 30 min at 32 °C before supplementing with 1 μ M Ac-LEDH-AFC (caspase-9 substrate) with or without 2 μ M Ac-LEDH-CHO (caspase-9 inhibitor) and incubated at 32 °C for 24 h. Fluorescence signal was treated similarly as for caspase-3 assay.

Caspase-8 activity. Assay was carried accordingly to the kit manufacturer with slight differences. Assay buffer contained in mM: 20 Hepes, 5 DTT, 2 EDTA, 5% sucrose and 0.1% Triton X-100 at pH 7.4. Assay buffer was supplemented with 1 μ M Ac-IETD-AMC (caspase-8 substrate) and with or without 2 μ M Ac-IETD-CHO and 50 μ l of cell lysate. This mix was incubated at 32 °C for 24 h. The fluorescence signal was treated similarly as for caspase-3 assay.

Cyt c release and PARP fragmentation. Cells in PBS with protease inhibitor cocktail were harvested with a Cell Lifter (Costar, Corning, Corning, NY, USA) and microcentrifuged at $10^4 \times g$ for 30 min to separate supernatant (cytosolic fraction) from pellet (particulate fraction). In all, 20 μ g of protein per lane were separated by SDS-PAGE in discontinuous gradient gels, blotted in PVDF membranes and revealed with anti-cyt c antibody (1 : 2000), anti-PARP antibody (1 : 1000) and anti- β -tubulin antibody (1 : 5000) as loading control. Optical density for each band was divided by the corresponding optical density of β -tubulin and this ratio in the presence of STS was taken as 100%.

STS-induced changes in the $[K^+]$, and $[Na^+]$, using atomic absorption spectrometry. Cells were washed with cold D-mannitol (6%) twice followed by the addition of 100 μ l of HNO₃ 0.1 N and harvested with Cell Lifter (Costar, Corning) and protein concentration determined. Sample $[K^+]$ and $[Na^+]$

were determined based on standard K^+ and Na^+ curves in 0.2% CsCl using a PerkinElmer (Waltham, MA, USA) AAnalyst 200 spectrometer. The internal ion concentration detected was normalized by corresponding protein concentration and assumed to represent 140 mM $[K^+]$ or 30 mM $[Na^+]$.

Statistical analyses. All data are presented as mean \pm S.E.M., where n represent different experiment (different cell passage). Statistical significance was evaluated by ANOVA and Tukey's tests. In all cases the STS-induced response was significantly different to basal level, so this analysis is not shown. * means that $P < 0.05$ when the tested condition was different with respect to the STS-induced response, except for Figure 2c.

Conflict of interest

The authors declare no conflict of interest.

Acknowledgements. We thank Conacyt for fellowship 162555 to E Benítez-Rangel and grant 44699 to A Guerrero-Hernández.

- Wyllie AH. Where o death, is thy sting? A brief review of apoptosis biology. *Mol Neurobiol* 2010; **42**: 4–9.
- Bortner CD, Gomez-Angelats M, Cidlowski JA. Plasma membrane depolarization without repolarization is an early molecular event in anti-Fas-induced apoptosis. *J Biol Chem* 2001; **276**: 4304–4314.
- Gilbert MS, Saad AH, Rupnow BA, Knox SJ. Association of BCL-2 with membrane hyperpolarization and radioresistance. *J Cell Physiol* 1996; **168**: 114–122.
- Wang L, Zhou P, Craig RW, Lu L. Protection from cell death by mcl-1 is mediated by membrane hyperpolarization induced by K(+) channel activation. *J Membr Biol* 1999; **172**: 113–120.
- Platoshyn O, Zhang S, McDaniel SS, Yuan JX. Cytochrome c activates K⁺ channels before inducing apoptosis. *Am J Physiol Cell Physiol* 2002; **283**: C1298–C1305.
- Yu SP, Yeh CH, Sensi SL, Gwag BJ, Canzoniero LM, Farhangrazi ZS *et al*. Mediation of neuronal apoptosis by enhancement of outward potassium current. *Science* 1997; **278**: 114–117.
- Bortner CD, Hughes Jr FM, Cidlowski JA. A primary role for K⁺ and Na⁺ efflux in the activation of apoptosis. *J Biol Chem* 1997; **272**: 32436–32442.
- Maeno E, Ishizaki Y, Kanaseki T, Hazama A, Okada Y. Normotonic cell shrinkage because of disordered volume regulation is an early prerequisite to apoptosis. *Proc Natl Acad Sci USA* 2000; **97**: 9487–9492.
- Wei L, Xiao AY, Jin C, Yang A, Lu ZY, Yu SP. Effects of chloride and potassium channel blockers on apoptotic cell shrinkage and apoptosis in cortical neurons. *PLoS Arch* 2004; **448**: 325–334.
- Arrebola F, Canizares J, Cubero MA, Crespo PV, Warley A, Fernandez-Segura E. Biphasic behavior of changes in elemental composition during staurosporine-induced apoptosis. *Apoptosis* 2005; **10**: 1317–1331.
- Bortner CD, Cidlowski JA. Caspase independent/dependent regulation of K(+), cell shrinkage, and mitochondrial membrane potential during lymphocyte apoptosis. *J Biol Chem* 1999; **274**: 21953–21962.
- Bortner CD, Sifre MI, Cidlowski JA. Cationic gradient reversal and cytoskeleton-independent volume regulatory pathways define an early stage of apoptosis. *J Biol Chem* 2008; **283**: 7219–7229.
- Hughes Jr FM, Bortner CD, Purdy GD, Cidlowski JA. Intracellular K⁺ suppresses the activation of apoptosis in lymphocytes. *J Biol Chem* 1997; **272**: 30567–30576.
- Dussmann H, Rehm M, Kogel D, Prehn JH. Outer mitochondrial membrane permeabilization during apoptosis triggers caspase-independent mitochondrial and caspase-dependent plasma membrane potential depolarization: a single-cell analysis. *J Cell Sci* 2003; **116**: 525–536.
- Cain K, Langlais C, Sun XM, Brown DG, Cohen GM. Physiological concentrations of K⁺ inhibit cytochrome c-dependent formation of the apoptosome. *J Biol Chem* 2001; **276**: 41985–41990.
- Karki P, Seong C, Kim JE, Hur K, Shin SY, Lee JS *et al*. Intracellular K(+) inhibits apoptosis by suppressing the Apaf-1 apoptosome formation and subsequent downstream pathways but not cytochrome c release. *Cell Death Differ* 2007; **14**: 2068–2075.
- Thompson GJ, Langlais C, Cain K, Conley EC, Cohen GM. Elevated extracellular [K⁺] inhibits death-receptor- and chemical-mediated apoptosis prior to caspase activation and cytochrome c release. *Biochem J* 2001; **357**: 137–145.
- Feeney B, Clark AC. Reassembly of active caspase-3 is facilitated by the propeptide. *J Biol Chem* 2005; **280**: 39772–39785.
- Tang D, Lahti JM, Kidd VJ. Caspase-8 activation and bid cleavage contribute to MCF7 cellular execution in a caspase-3-dependent manner during staurosporine-mediated apoptosis. *J Biol Chem* 2000; **275**: 9303–9307.

20. Wurstle ML, Laussmann MA, Rehm M. The caspase-8 dimerisation/dissociation balance is a highly potent regulator of caspase-8, -3, -6 signalling. *J Biol Chem* 2010; **285**: 33209–33218.
21. Sauve R, Simoneau C, Parent L, Monette R, Roy G. Oscillatory activation of calcium-dependent potassium channels in HeLa cells induced by histamine H1 receptor stimulation: a single-channel study. *J Membr Biol* 1987; **96**: 199–208.
22. Stein MA, Mathers DA, Yan H, Baimbridge KG, Finlay BB. Enteropathogenic *Escherichia coli* markedly decreases the resting membrane potential of Caco-2 and HeLa human epithelial cells. *Infect Immun* 1996; **64**: 4820–4825.
23. Szabo I, Brutsche S, Tombola F, Moschioni M, Satin B, Telford JL *et al*. Formation of anion-selective channels in the cell plasma membrane by the toxin VacA of *Helicobacter pylori* is required for its biological activity. *EMBO J* 1999; **18**: 5517–5527.
24. Remillard CV, Yuan JX. Activation of K⁺ channels: an essential pathway in programmed cell death. *Am J Physiol Lung Cell Mol Physiol* 2004; **286**: L49–L67.
25. Nicolier M, Decrion-Barthod AZ, Launay S, Pretet JL, Mougín C. Spatiotemporal activation of caspase-dependent and -independent pathways in staurosporine-induced apoptosis of p53wt and p53mt human cervical carcinoma cells. *Biol Cell* 2009; **101**: 455–467.
26. Tafani M, Minchenko DA, Serroni A, Farber JL. Induction of the mitochondrial permeability transition mediates the killing of HeLa cells by staurosporine. *Cancer Res* 2001; **61**: 2459–2466.
27. Johnson CR, Jarvis WD. Caspase-9 regulation: an update. *Apoptosis* 2004; **9**: 423–427.
28. Soldani C, Scovassi AI. Poly (ADP-ribose) polymerase-1 cleavage during apoptosis: an update. *Apoptosis* 2002; **7**: 321–328.
29. Franco R, Bortner CD, Cidlowski JA. Potential roles of electrogenic ion transport and plasma membrane depolarization in apoptosis. *J Membr Biol* 2006; **209**: 43–58.
30. Storey NM, Gomez-Angelats M, Bortner CD, Armstrong DL, Cidlowski JA. Stimulation of Kv1.3 potassium channels by death receptors during apoptosis in Jurkat T lymphocytes. *J Biol Chem* 2003; **278**: 33319–33326.
31. Lovering AL, Ride JP, Bunce CM, Desmond JC, Cummings SM, White SA. Crystal structures of prostaglandin D(2) 11-ketoreductase (AKR1C3) in complex with the nonsteroidal anti-inflammatory drugs flufenamic acid and indomethacin. *Cancer Res* 2004; **64**: 1802–1810.
32. Greenwood IA, Leblanc N. Overlapping pharmacology of Ca²⁺-activated Cl⁻ and K⁺ channels. *Trends Pharmacol Sci* 2007; **28**: 1–5.
33. Gogelein H, Dahlem D, Englert HC, Lang HJ. Flufenamic acid, mefenamic acid and niflumic acid inhibit single nonselective cation channels in the rat exocrine pancreas. *FEBS Lett* 1990; **268**: 79–82.
34. Chan TA, Morin PJ, Vogelstein B, Kinzler KW. Mechanisms underlying nonsteroidal antiinflammatory drug-mediated apoptosis. *Proc Natl Acad Sci USA* 1998; **95**: 681–686.
35. Bootman MD, Cheek TR, Moreton RB, Bennett DL, Berridge MJ. Smoothly graded Ca²⁺ release from inositol 1,4,5-trisphosphate-sensitive Ca²⁺ stores. *J Biol Chem* 1994; **269**: 24783–24791.
36. Garcia-Calvo M, Peterson EP, Rasper DM, Vaillancourt JP, Zamboni R, Nicholson DW *et al*. Purification and catalytic properties of human caspase family members. *Cell Death Differ* 1999; **6**: 362–369.



Cell Death and Disease is an open-access journal published by Nature Publishing Group. This work is licensed under the Creative Commons Attribution-NonCommercial-No Derivative Works 3.0 Unported License. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/3.0/>

Supplementary Information accompanies the paper on Cell Death and Disease website (<http://www.nature.com/cddis>)