

Editorial

Autophagy in stress and disease

M Chiara Maiuri^{1,2,3,4} and G Kroemer^{*,1,2,3,4,5}

Cell Death and Differentiation (2015) 22, 365–366; doi:10.1038/cdd.2014.236

Macroautophagy, here referred to as ‘autophagy,’ constitutes one of the most spectacular phenomena in cell biology beyond cell fusion, cell division, differentiation and demise. Metaphorically spoken, it constitutes a process in which the cell sequesters portions of itself in its stomach (the autophagosomes) and then assures their complete degradation in its digestive tract (the lysosomes).

Autophagy constitutes one of the most elementary reactions that a cell may have to adapt itself to a changing microenvironment. For instance, in conditions of dwindling external resources, be it nutrients, growth factors or oxygen, the cell may mobilize its stock of potentially energy-rich macromolecules by autophagy, thereby converting proteins and lipids into life-preserving fuel for bioenergetic reactions. In addition, the cell can take advantage of the autophagic machinery to remove damaged, dysfunctional and potentially harmful organelles such as uncoupled mitochondria from its cytoplasm or to destroy useless and even dangerous protein aggregates. Hence, autophagy constitutes an essential mechanism for the recycling of cytoplasmic material and in fine cleaning and rejuvenating extranuclear compartments, especially in non-dividing cells such as neurons or cardiomyocytes. Beyond its homeostatic function, autophagy also has a major role in hormetic reactions. Hormesis can be defined as a process in which the exposure of cells, organs or organisms to a mild stress allows them to mount an adaptive response that allow them to tolerate a later, stronger and normally lethal stress. One well-known example of hormesis is ischemic preconditioning in which a short episode of ischemia reduces the death of heart muscle cells to an otherwise fatal infarction. In this context, autophagy induction has a major role in increasing the robustness of the system, protecting it from deadly stress. Nonetheless, there are also specific situations in which an excess of autophagy may ultimately cause the death of cells by excessive self-digestion. This potentially lethal role of autophagy has received the name of ‘autosis’.

The present Special Issue of *Cell Death & Differentiation* deals with the physiological and pathological functions of autophagy in cell stress and disease. Liu and Levine¹ provide an overview over the potential roles of autosis and autophagic cell death in health and disease. Filomeni *et al.*² demonstrate the importance of autophagy regulation by reactive oxygen species and reactive nitrogen species in the context of

cytoplasmic processes and DNA damage signaling. Orhon *et al.*³ insist on the important role of primary cilia, which are microtubule-based structures located at the cell surface of many cell types, as potential sensors of autophagy-inducing stimuli that are in turn affected in their biogenesis and function by autophagic responses. Nikolettou *et al.*⁴ demonstrate the essential role of autophagy for the normal function of neurons in model organisms such as *Caenorhabditis elegans*, as well as the potential role of autophagy in the the unwarranted demise of neurons induced by pathological stimuli including overexcitation.

Two original articles deal with the complex regulation of mitophagy, a version of autophagy that assures the specific turnover of damaged mitochondria. Rossin *et al.*⁵ demonstrate that transglutaminase-2 has an unexpected essential role in mitophagy. Moreover, Strapazzon *et al.*⁶ show that AMBRA-1 can stimulate mitophagy through a novel pathway that does not require parkin nor STQM1/LC3. Menzies *et al.*⁷ reveal that inhibition of calpains with calpastatin increases autophagic turnover, thereby reducing neuronal damage in fly and mouse models of transgene-enforced expression of mutant huntingtin protein. In another differentiated cell type, Rožman *et al.*⁸ show that autophagy is not essential for neutrophil granulopoiesis, having instead a negative impact on the generation of neutrophils. Anding and Baehrecke⁹ reveal the broad implication of a specific protein, Vps15, which is part of the autophagy-regulatory Beclin 1 complex, in different cellular contexts and find that it is necessary for both stress-induced and developmentally programmed autophagy in various tissues in *Drosophila melanogaster*.⁹ Ber *et al.*¹⁰ present data on DAPK2 (death-associated protein kinase 2), a Ca²⁺-regulated serine/threonine kinase, that directly interacts with and phosphorylates the protein raptor within the mTORC1 (mammalian target of rapamycin complex 1), which is one of the major autophagy regulators. Rodríguez-Muela *et al.*¹¹ make a convincing case in favor of the cytoprotective role of autophagy in a mouse model of retinitis pigmentosa, an inherited, degenerative eye disease that causes severe vision impairment and often blindness. Similarly, Zhou *et al.*¹² reveal the essential role of autophagy for maintaining the survival and normal function of rod photoreceptors. Finally, two papers deal with the mode of action of two autophagy-inducing longevity-extending compounds. Pietrocola *et al.*¹³ show that spermidine, which has universal

¹Equipe 11 Labellisée Ligue Contre le Cancer, Centre de Recherche des Cordeliers, INSERM U 1138, Paris, France; ²Université Paris Descartes, Sorbonne Paris Cité, Paris, France; ³Université Pierre et Marie Curie, Paris, France; ⁴Metabolomics and Cell Biology Platforms, Gustave Roussy, Villejuif, France and ⁵Pôle de Biologie, Hôpital Européen Georges Pompidou, AP-HP, Paris, France

*Corresponding author: G Kroemer, Equipe 11 Labellisée Ligue Contre le Cancer, Centre de Recherche des Cordeliers, INSERM U1138, 15 rue de l'école de médecine, F-75006 Paris, France. Tel: +33 1 44 27 76 67; Fax: +33 1 44 27 76 74; E-mail: kroemer@orange.fr

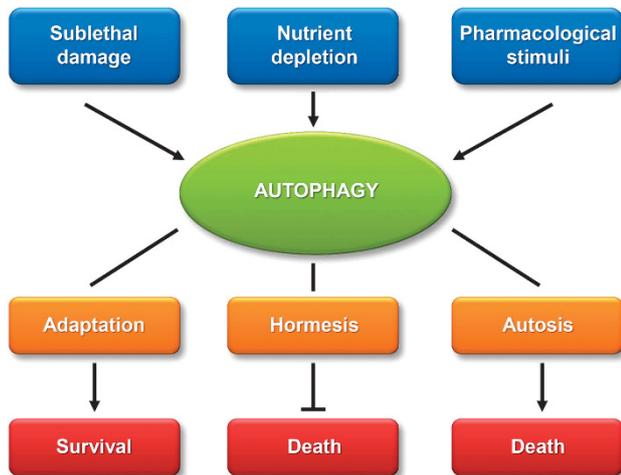


Figure 1 Physiological roles of autophagy. Autophagy can be induced by damage, starvation or pharmacological cues. Autophagy then favors the adaptation of cells, hormetic reactions (protection against otherwise lethal stimuli) or autosis (cell death mediated by autophagy)

anti-aging effects across multiple model organisms including yeast, nematodes, flies and mice, induces autophagy owing to its capacity to inhibit the enzymatic activity of the EP300 acetyltransferase. Rockenfeller *et al.*¹⁴ describe the capacity of phosphatidylethanolamine to induce autophagy in yeast and mammalian cells, an effect that is echoed by a dramatic anti-aging effect in yeast.

Altogether, this special issue of *Cell Death & Differentiation* reflects the stunning role of autophagy in multiple cell types, its induction by a myriad of different stressful stimuli, as well as its essential role in maintaining homeostatic and hormetic

functions. Beyond these life-preserving and anti-aging effects of autophagy, recent evidence suggest that this phenomenon also has a dark side, causing pathological cell death in specific circumstances (Figure 1). Hence, the present series of articles will incite cell death researchers all over the world to study autophagy in its multiple positive and negative aspects with respect to cellular and organismal survival. No reader can remain indifferent.

1. Liu Y, Levine B. *Cell Death Differ* 2015; **22**: 367–376.
2. Filomeni G, De Zio D, Cecconi F. Oxidative stress and autophagy: the clash between damage and metabolic needs. *Cell Death Differ* 2015; **22**: 377–388.
3. Orhon I, Dupont N, Pampliega O, Cuervo AM, Codogno P. Autophagy and regulation of cilia function and assembly. *Cell Death Differ* 2015; **22**: 389–397.
4. Nikolettou V, Papandreou M-E, Tavernarakis N. Autophagy in the physiology and pathology of the central nervous system. *Cell Death Differ* 2015; **22**: 398–407.
5. Rossin F *et al.* *Cell Death Differ* 2015; **22**: 408–418.
6. Strappazzon F, Nazio F, Corrado M, Cianfanelli V, Romagnoli A, Fimia GM *et al.* AMBRA1 is able to induce mitophagy via LC3 binding, regardless of PARKIN and p62/SQSTM1. *Cell Death Differ* 2015; **22**: 419–432.
7. Menzies FM, Garcia-Arencibia M, Imarisio S, O'Sullivan NC, Ricketts T, Kent BA. Calpain inhibition mediates autophagy-dependent protection against polyglutamine toxicity. *Cell Death Differ* 2015; **22**: 433–444.
8. Rožman S, Yousefi S, Oberson K, Kaufmann T, Benarafa C, Simon HU. The generation of neutrophils in the bone marrow is controlled by autophagy. *Cell Death Differ* 2015; **22**: 445–456.
9. Anding AL, Baehrecke EH. *Vps15* is required for stress induced and developmentally triggered autophagy and salivary gland protein secretion in *Drosophila*. *Cell Death Differ* 2015; **22**: 457–464.
10. Ber Y *et al.* *Cell Death Differ* 2015; **22**: 465–475.
11. Rodríguez-Muela N *et al.* *Cell Death Differ* 2015; **22**: 476–487.
12. Zhou Z, Doggett TA, Sene A, Apte RS, Ferguson TA. *Cell Death Differ* 2015; **22**: 506–516.
13. Pietrocola F, Lachkar S, Enot DP, Niso-Santano M, Bravo-San Pedro JM, Sica V *et al.* Spermidine induces autophagy by inhibiting the acetyltransferase EP300. *Cell Death Differ* 2015; **22**: 488–495.
14. Rockenfeller P, Koska M, Pietrocola F, Minois N, Knittelfelder O, Sica V *et al.* Phosphatidylethanolamine positively regulates autophagy and longevity. *Cell Death Differ* 2015; **22**: 496–505.