Very low oxygen concentration (0.1%) reveals two FDCP-Mix cell subpopulations that differ by their cell cycling, differentiation and p27^{KIP1} expression

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Oxygen (O₂) concentrations in bone marrow vary from 4% in capillaries to <0.1% in subendosteum, in which hematopoietic stem cells reside in specific niches. Culture at low O₂ concentrations (3, 1 and 0.1%) influences hematopoietic stem and progenitor cells survival, proliferation and differentiation, depending on their level of differentiation. Culture of human CD34⁺ cells at low O₂ concentrations (O₂ ≤ 3%) maintains stem cell engraftment potential better than at 20% O₂ (NOD/Scid xenograft model). In contrast, progenitors disappear from cultures at/or <1% O₂ concentrations. A very low O₂ concentration (0.1%) induces CD34⁺ quiescence in G₀. The exploration of molecules and mechanisms involved in hematopoietic stem and progenitor cells' quiescence and differentiation related to low O₂ concentrations is unfeasible with primary CD34⁺ cells. Therefore, we performed it using murine hematopoietic nonleukemic factor-dependent cell Paterson (FDCP)-Mix progenitor cell line. The culture of the FDCP-Mix line at 0.1% O₂ induced in parallel G₀ quiescence and granulo-monocytic differentiation of most cells, whereas a minority of undifferentiated self-renewing cells remained in active cell cycle. Hypoxia also induced hypophosphorylation of pRb and increased the expression of p27^{KIP1}, the two proteins that have a major role in the control of G₀ and G₁ to S-phase transition.

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Physiological oxygen (O₂) concentrations in tissues are much lower than in atmospheric air (20%), ranging from 14% in pulmonary alveoli to <1% in several organs.¹ Numerous publications show the major role of variations of O₂ concentration in the responses of normal cells, transformed cells and leukemic cells from primoculture or from continuous cell lines.^{2–8} Low O₂ concentration induces apoptosis or cell survival, proliferation or quiescence, self-renewal or differentiation, depending on the degree of hypoxia, the cell type and the level of differentiation.

Response to hypoxia is largely dependent on hypoxia-inducible factor 1 (HIF-1), a heterodimeric transcription factor. At O₂ concentrations <5%, the HIF-1 α subunit is no longer degraded by the proteasome^{9,10} and dimerizes with HIF-1 β . The active complex then induces the expression of numerous genes including those of growth factors (erythropoietin (Epo) and vascular endothelial growth factor (VEGF)), transcription factors and cell cycle proteins.^{11–14}

In bone marrow, hematopoietic cells are permanently exposed to O₂ concentrations ranging from 4% in capillaries to <0.1% in endosteal areas,^{15–17} in which the most primitive

stem cells reside.^{18,19} Hematopoietic stem cells (HSCs) are better preserved in cultures at low O₂ (1–3%) than at atmospheric O₂ concentration (20%).^{5,20,21} We previously showed that culture at 0.1% O₂ prevents cord blood G₀ CD34⁺ cells to enter in active cell cycle and induces those in active proliferation to re-enter in G₀.²² The effects of low O₂ concentration on the survival, proliferation and differentiation of normal and leukemic hematopoietic progenitors are more controversial. Indeed, in some articles a pro-differentiative effect^{6,23–25} was pointed out, whereas in others it was suggested that oxygen levels <1% block cell proliferation and induce their rapid disappearance from cultures, probably by inducing apoptosis.^{4,21,26,27}

Culture at low O₂ concentration (0.5%) causes cell cycle arrest in G₁ of mouse embryonic fibroblasts and splenic B lymphocytes by inducing Rb hypophosphorylation and p27^{KIP1} upregulation.^{28,29} All these results led us to explore the effects of low O₂ concentration on the proliferation and differentiation of hematopoietic progenitors. Extensive protein studies using normal human or murine primoculture progenitor cells were quite unfeasible for two reasons: heterogeneity

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Abbreviations: O₂, oxygen; HIF-1, hypoxia-inducible factor-1; Epo, erythropoietin; VEGF, vascular endothelial growth factor; HSC, hematopoietic stem cell; PI, propidium iodide; CFSE, carboxyfluorescein diacetate succinimidyl diester; MFI, mean fluorescence intensity; FDCP, factor-dependent cell Paterson; FCM, flow cytometry; CRA, culture-repopulating ability; CFC, colony-forming cell; LC, liquid culture; KIP, kinase inhibitor protein; CDK, cyclin-dependent kinase; IMDM, Iscove's modified Dulbecco's medium; CM, conditioned medium; HS, horse serum; RT, room temperature; CFA, colony-forming ability

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of the CD34⁺ cell population and difficulty of obtaining them in large numbers. As the murine cell line, factor-dependent cell Paterson (FDCP)-Mix, which is less heterogeneous, has major features of normal hematopoietic progenitors, we tested its response to hypoxia. Indeed, FDCP-Mix cells have a normal karyotype, are non-leukemic, strictly depend on IL-3 for their survival and proliferation, and differentiate toward different myeloid lineages when cultured with other cytokines.³⁰ Varying the concentrations of O₂ from 20 to 0.1% in cultures of FDCP-Mix cells had a time-dependent effect on their proliferation and cell cycle, with cells reaching quiescence (G₀) after 72 h of culture at 0.1% O₂. We then used these cells to analyze the self-renewal/proliferation/differentiation balance and the variations of p27^{KIP1} levels and pRb phosphorylation, the two proteins potentially involved in G₀ cell cycle arrest.

Results

Very low O₂ concentration inhibits FDCP-Mix cell growth. Low O₂ concentration induced a time-dependent FDCP-Mix growth inhibition (Figure 1a). The number of living cells at 24 h was not significantly different in 20% O₂ versus 0.1% O₂ concentration cultures, nor was their doubling time (approximately 30 h in both conditions; Figure 1b). At later time points, the growth of FDCP-Mix cells cultured at 0.1% O₂ was dramatically reduced, with a doubling time that reached 104 h after >48 h of culture. As the percentage of apoptotic cells in two conditions was similar (Figure 1c) after 24, 48 and 72 h of culture, we concluded that the growth arrest observed at 0.1% O₂ was because of the inhibition of proliferation of FDCP-Mix cells after 24 h.

Very low O₂ concentration (0.1%) induces FDCP-Mix cell arrest in G₀. We then analyzed the FDCP-Mix cell cycle status after 24, 48 and 72 h of culture at 20 and 0.1% O₂, using double labeling with propidium iodide (PI, a quantitative DNA fluorescent dye) and anti-Ki-67 antibodies (Ki-67 antigen is expressed in the nucleus of cycling cells and absent from G₀ cells) to discriminate cells in G₀ (DNA = 2N; Ki-67⁻) from cells in G₁ (DNA = 2N; Ki-67⁺), S (2N < DNA < 4N; Ki-67⁺) and G₂/M (DNA = 4N; Ki-67⁺). As shown in Figure 2a, the percentage of FDCP-Mix cells in G₀ remained stable (13.6 ± 2.3%) during 72 h of culture at 20% O₂. In contrast, at 0.1% O₂ the percentage of G₀ cells significantly increased to 48.8 ± 5.4% after 72 h of culture, whereas the percentage of cells in G₁, S and G₂/M decreased in parallel (Figure 2b–d). To determine whether this G₀ phase accumulation was due to cells previously in G₀ and G₁ or because of cells entering in G₀ after one or several divisions, FDCP-Mix cells labeled with carboxyfluorescein diacetate succinimidyl diester (CFSE; a stable vital cellular fluorescent dye whose decay allows to analyze cell divisions) were cultured for 72 h at 20% O₂ and at 0.1% O₂. At this time point, division rates (CFSE labeling) and cell cycle status (Ki-67⁺ or Ki-67⁻) of cultured FDCP-Mix cells were co-analyzed by flow cytometry (FCM). As shown by the comparison of panels b and c in Figure 3, the division rate of FDCP-Mix cells was slower at 0.1% than at 20% O₂. This is because of the

Figure 1 Influence of O₂ concentration on the proliferation and apoptosis of FDCP-Mix cells. Cells were seeded at 1×10^5 cells/ml and cultured for 24, 48 and 72 h at 20, 3, 1 or 0.1% O₂. (a) Living cell counts. (b) The cell doubling time was calculated as follows: $t \ln 2 / \ln (cf/ci)$ where t is time of culture in hours; $\ln$ is natural logarithm; and cf and ci are living cells concentrations at final and initial culture time points, respectively. (c) Percentage of apoptotic cells (annexin V-positive cells). Values are the mean ± S.D. of five independent experiments ([†] $P < 0.05$; Kruskal–Wallis test)

association of two cell cycle events: (1) the extension of the cell cycle duration as concluded from the respective percentages of Ki-67⁺ cells in each division class and (2) the progressive accumulation of G₀ cells after each division (revealed by comparing the percentages of Ki-67⁻ cells in each division class at 20 and 0.1% O₂).

Very low O₂ concentration (0.1%) induces the differentiation of a majority of FDCP-Mix cells. May-Grünwald/Giemsa staining of FDCP-Mix cytopins showed that after 72 h of culture at 0.1% O₂, the majority of cells

Figure 2 Influence of O₂ concentration on the cell cycle of FDCP-Mix cells. Cells were seeded at 1×10^5 cells/ml and cultured at 20% or 0.1% O₂ for 24, 48 and 72 h. Double labeling with PI and Ki-67 antibody identifies cells (a) in G₀ (DNA = 2N; Ki-67⁻), (b) in G₁ (DNA = 2N; Ki-67⁺), (c) in S (2N < DNA < 4N; Ki-67⁺) and (d) in G₂/M (DNA = 4N; Ki-67⁺). Values are the mean $\pm$ S.D. of seven independent experiments ($^{\dagger}P < 0.05$, $^*P < 0.01$; Friedman test)

acquired a granulo-monocytic differentiation (Figure 4a) that was confirmed by FCM analysis of the expression of Gr-1 (granulocytic differentiation marker), Mac-1 (monocytic differentiation marker), as well as CD34 and CD117 (markers of primitive hematopoietic progenitors) cell surface markers (Figure 4b–d). Indeed, after 3 days of culture at 0.1% O₂, the percentage of cells positive for Gr-1 and CD117, respectively, increased from 26 to 63% and decreased from 27 to 14% (Figure 4b and c). In parallel, as expected, we observed a decrease in the mean fluorescence intensity (MFI) of the CD34 primitive marker together with an increase in the MFI of the granulo-monocytic marker Mac-1 and Gr-1 (Figure 4d). These results confirmed that culture at 0.1% O₂ induces the granulo-monocytic differentiation of the FDCP-Mix cell line.

Very low O₂ concentration (0.1%) also maintains a minor population of FDCP-Mix cells with culture-repopulating ability (CRA). FDCP-Mix cells issued from 72 h of liquid culture (LC1) at 0.1% O₂ were sequentially replated every 72 h at 1.10^5 cells/ml at 20% O₂ to test their ability to repopulate later LCs (LC2–LC5), as determined by counting the total number of cells in LC2 to LC5 (CRA_{cell}). The growth rate of cells incubated in LC1 at atmospheric O₂ concentration remained stable throughout the experiment (Figure 5a, left part of the graph). The growth rate of cells issued from LC1 incubated at 0.1% O₂ (Figure 5a, right part of the graph) in nonselective, normoxic LC2–LC5 (Figure 5a) remained stable during LC2 and LC3, increased during LC4 and reached the growth rate showed by cells incubated in normoxic LC1 during LC5. In the meantime, the percentage of cells in G₀ decreased and the percentage of cells in G₁, S and G₂/M started to increase during LC2 to reach control levels at LC4 (data not shown). To confirm that this progressive repopulation of LC was because of the maintenance in hypoxia of a limited cell subpopulation,

cells rescued from nonselective (normoxic) LC2–LC5 were cultured in semisolid medium to identify and count, after 11 days, the number of colonies issued from colony-forming cell (CFC). The percentage of these CRA_{CFC} cells at the end of 0.1% O₂ LC1 was 14 times lower than after 20% LC1 and progressively returned to similar values in LC4 (Figure 5b).

Relationships between G₀ quiescence, Gr-1 expression and CRA_{CFC} cells. As the differentiation after 3 days of culture at 0.1% O₂ was accompanied by the G₀ entry of a large percentage of FDCP-Mix cells, we explored the relationship between these two phenomena. As a first step, we sorted Gr-1⁺ and Gr-1⁻ cells after 3 days of LC1 at 20 or 0.1% O₂. The microscopic observation on May-Grünwald/Giemsa cytopins (Figure 6a) confirmed that Gr-1⁺ cells had a typical morphology of terminally differentiated granulocytes including donut-shaped nuclei (arrows ►), whereas Gr-1⁻ cells kept round nuclei with more basophilic cytoplasm as observed in control culture conditions (Figure 4a). At both O₂ concentrations, all CRA_{cell}⁻ and CRA_{CFC}-type cells were contained within the Gr-1⁻ population (Figure 6b and c). Analysis of the cell cycle status of Gr-1⁺ and Gr-1⁻ cells showed that CRA-type undifferentiated Gr-1⁻ cells were mostly cycling, whereas differentiated Gr-1⁺ cells without CRA were in G₀ (Figure 6d). In addition, a 24-h treatment of FDCP-Mix cells with 5FU (10 μ g/ml) at the end of LC1, followed by medium renewal and replating in LC2, dramatically decreased CRA_{cell} and CRA_{CFC} similarly in both O₂ conditions (data not shown). Altogether, these data show that at very low 0.1% O₂ concentration, most FDCP-Mix cells differentiate and enter in the quiescent G₀ status, whereas a minority of undifferentiated cells, those that maintain the self-renewal of the cell line, remain in active cell cycle.

Figure 3 Hypoxia (0.1% O₂) reduces the number of cell divisions of FDCP-Mix cells. Dot plots of FDCP-Mix cells labeled with CFSE and anti Ki-67 antibody. Cells before culture (a) and after 72 h of culture at 20% (b) and 0.1% (c) O₂. The Ki-67 antibody labeling discriminates quiescent cells (Ki-67⁻) from cycling cells (Ki-67⁺). Decreasing levels of CFSE fluorescence (from right to left) reflect the increasing number of cell divisions. One representative experiment out of three is shown

FDCP-Mix cells in G₀ overexpress hypophosphorylated pRb and p27^{KIP1} proteins. Because of their role in the regulation of cell cycling, we analyzed the total amount of p27^{KIP1} and pRb proteins and the phosphorylation status of pRb after 24, 48 and 72 h of culture at 20% O₂ and 0.1% O₂. As expected, the total quantity of pRb protein remained stable and similar during 72 h culture at 20 and 0.1% O₂ (Figure 7a). The S-phase transition is dependent on pRb phosphorylation on several specific sites. Therefore, we studied this protein phosphorylation status. Indeed, unphosphorylated pRb progressively increased whereas phosphorylated pRb disappeared during 72 h culture of FDCP-Mix cells at 0.1% O₂ (data not shown). We then analyzed several pRb phosphorylation sites involved in the G₁ to S-phase transition. The Ser 608, Ser 807/811 and

Thr 821 sites (stably phosphorylated at 20% O₂) became very faint at 0.1% O₂ (Figure 7b). In the meantime, the p27^{KIP1} level in FDCP-Mix cells strongly increased during cultures at 0.1% O₂, whereas it remained stable at 20% O₂ (Figure 8a). Upregulation of the p27^{KIP1} protein by hypoxia in FDCP-Mix cells was partly related to transcriptional mechanisms as p27^{KIP1} mRNA level significantly increased during cultures at 0.1% O₂ (2.76-, 3- and 2.4-fold at 24, 48 and 72 h, respectively; Figure 8b). To know whether the regulation of the p27^{KIP1} protein was linked to the G₀ quiescence and differentiation of the FDCP-Mix cell line, we explored its expression in the Gr-1⁻ and Gr-1⁺ populations by flow cytometry. Both the level of p27^{KIP1} fluorescence intensity (Figure 8c and d) and the percentages of cells expressing high levels of p27^{KIP1} were different in Gr-1⁻ and Gr-1⁺ cells.

Discussion

In the past 10 years, an increasing number of publications were devoted to the physiological low bone marrow O₂ concentrations (from <0.1% O₂ in sub-endosteal bone marrow niches to 4–5% in vascular areas compared with 21% in atmospheric air) and to the exploration of their role in the regulation of hematopoiesis.^{1,15,16} We and others showed that varying O₂ concentrations (from 20 to 0.1%) in cultures of hematopoietic stem and progenitor cells modified their quiescence *versus* proliferation and self-renewal *versus* differentiation balances.^{2,4–7,19–23,31–33} To investigate the molecules and mechanisms involved in these phenomena, we used the FDCP-Mix hematopoietic cell line that is considered to share numerous phenotypic and functional properties with bone marrow hematopoietic progenitors. Indeed, FDCP-Mix cells have a normal karyotype, are non-leukemic, strictly depend on IL-3 for their survival and proliferation, and differentiate towards different myeloid lineages when cultured with other cytokines.³⁰ In addition, when cultured at 0.1% O₂, cycling FDCP-Mix cell line progressively accumulated in G₀ after one to few divisions.

This accumulation in G₀ was accompanied by a major granulo-monocytic differentiation, as shown by sorting Gr-1⁺ (differentiated) and Gr-1⁻ (undifferentiated) FDCP-Mix cells after 3 days of culture at 20 and 0.1% O₂. In both O₂ conditions, the Gr-1⁻ FDCP-Mix subset had a high CRA and a high percentage of cycling cells, whereas the Gr-1⁺ differentiated fraction was mostly in G₀ with a quite null CRA. This result suggests a link between cell cycle arrest and differentiation.

Some articles found a differentiating effect of low O₂ concentrations on leukemic cells, either from primoculture or from cell lines^{6,23–25} similar to the one observed with FDCP-Mix cells. In contrast, Giuntoli *et al.*⁸ showed that when the K562 cell line (issued from a blast-crisis chronic myeloid leukemia – an HSC malignancy – patient) was cultured at 0.1% O₂ for 7 days, the number of cells dramatically decreased (by 1 log) and most cells underwent apoptosis, whereas a minor hypoxia-resistant subpopulation repopulated LC2 incubated at 20% O₂, showing full CRA maintenance. Using the murine MEL cell line, they obtained similar results, and found, in addition, at 0.1% O₂ a quiescent subset of CRA cells that resisted to 5FU treatment.⁴ However, in these two

Figure 4 Hypoxia (0.1% O₂) increases granulo-monocytic differentiation of FDCP-mix cells. **(a)** After 72 h of culture at 20 and 0.1% O₂, cells were harvested, cytospun and stained with May-Grünwald/Giemsa. **(b)** After 72 h of culture at 20 and 0.1% O₂, cells were labeled with appropriate isotypic antibodies, anti-Gr-1, anti-Mac-1, anti-CD34 or anti-CD117-specific antibodies and analyzed by FCM. Histograms of one representative experiment are shown. **(c)** This table gives the mean ± S.D. of the percentage of cells positive for the Gr-1, Mac-1, CD34 and CD117 markers after 3 days of culture at 20 and 0.1% O₂. These percentages were calculated from three independent experiments. **(d)** Variation of the MFI of Gr-1-, Mac-1-, CD34- and CD117-positive cells after 72 h of culture at 20 or 0.1% O₂. For each marker, the value represents the mean ratio ± S.D. of the MFI measured at 0.1% O₂/MFI measured at 20% O₂ in three independent experiments

articles, cell differentiation was not studied. When the FDCP-Mix cell line was cultured under similar conditions, apoptosis did not increase significantly, and most cells underwent proliferation arrest and differentiated, whereas a small population of slowly cycling CRA cells (as killed by 24 h of 5FU) persisted after 3 and even 7 days of culture at 0.1% O₂. These differences between FDCP-Mix (and the leukemic cell lines that differentiate in hypoxia) on one hand and K562, MEL and CD34⁺ cells on the other could be due (1) to the fact that FDCP-Mix and the leukemic cell lines that differentiate in hypoxia^{6,23–25} derive from progenitors and not from HSC, (2) to their lower sensitivity to apoptosis than primary CD34⁺ cells in hypoxic cultures owing to their immortalized status (but this does not apply to the K562 and MEL cell lines) or (3) to an association of both phenomena.

Our previous results showing that primoculture progenitors disappear rapidly during cultures of murine bone marrow or human CD34⁺ cells at 1% O₂^{21,27} whereas leukemic and FDCP-Mix cell lines survive and differentiate strengthen the hypothesis that the maturation of FDCP-Mix cells in hypoxia is linked to their acquired immortalized status. This allows their prolonged survival as differentiated cells in hypoxia, whereas primoculture progenitors rapidly die in these conditions before reaching maturation.

The permanent cycling of CRA type and the absence of apoptosis in cultures of FDCP-Mix cells at very low O₂

concentration that contrasts with the apoptosis of a large percentage and the quiescence of a minority of primoculture CD34⁺ and MEL cells cannot be explained by the immortalized status of FDCP-Mix, as MEL is also a permanent (and leukemic) cell line. We suggest that this major functional difference in response to very low O₂ concentrations is because of the 'stem' or 'progenitor' cell status of these different cell types. Together, the above-mentioned results with primary CD34⁺ cells, K562 cells, other leukemic cell lines and our present work imply that regulation by very low O₂ concentrations (<1%) of cycling, differentiation and survival, depending on the differentiation level of cells (stem cells *versus* progenitors) whether normal or leukemic tissue, is in question.

These results led us to explore the molecules involved in the cell cycle modifications induced by hypoxia. The G₀ and/or G₁ (undefined as neither Ki-67 nor any other G₀ marker was used) quiescence of fibroblasts and B lymphocytes induced by serum starvation or hypoxia is linked to p27^{KIP1} increase that blocks the cyclin/cyclin-dependent kinase (CDK) activity, thus leading to pRb hypophosphorylation.^{11,28,29,34–37} Conversely, phosphorylations of pRb on multiple Ser/Thr sites by cyclin/CDK during the G₁ phase lead to the release of molecules required for the transition to S phase.³⁸ In brief, the phosphorylation of Thr 821 releases histone deacetylase; the phosphorylation of Ser 807/811 releases c-abl and the

Figure 5 Hypoxia (0.1%) maintains a minor population of FDCP-Mix cells with culture-repopulating ability (CRA). After 3 days of primary liquid culture (LC1) at 20 or 0.1% O₂, cells were serially replated at 1×10^5 cells/ml at 20% O₂ every 3 days (LC2–LC5). At the end of each LC (LC1–LC5), (a) the number of living cells (CRA_{cell}) in liquid culture was counted and (b) their CFC content were revealed by the CFA assay that allowed to count the number of colonies after 11 days of culture at 20% in semisolid medium (CRA_{CFC}). Values are the mean $\pm$ S.D. from four independent experiments. Statistical analysis of differences between similar LC at 20 and at 0.1% O₂ was performed using Mann–Whitney test ($^{\dagger}P < 0.05$, $^*P < 0.01$). Statistical analysis of differences between serial LCs in the 20 and 0.1% O₂ group used the Friedman test ($^{\dagger}P < 0.05$)

phosphorylation of Ser 608 participates in the release of the master transcription factor E2F. These three molecules have a major role in the G₁ to S transition. Hypoxia dramatically reduced the phosphorylation of these three pRb sites in FDCP-Mix cells, suggesting that dephosphorylated pRb participate to their quiescence. Hypoxia also induced a significant increase in the p27^{KIP1} protein and mRNA in FDCP-Mix cells. Finally, we showed that the quiescent differentiated Gr-1⁺ cells expressed higher p27^{KIP1} levels than the Gr-1 cycling cells, a fraction containing CRA cells. That is most probably responsible for the accumulation of unphosphorylated pRb. Our present data that extend personal preliminary results³⁹ are similar to those very recently published by Eliasson *et al.*⁴⁰ Altogether, they show that p27^{KIP1} is involved in the regulation of FDCP-Mix cell cycling in hypoxia.

The last important point of our work is that it revealed a minor stem cell-like subpopulation (that repopulates LCs after returning them to 20% O₂) of the FDCP-Mix cell line that kept its self-renewal potential after 3–7 days of culture at 0.1% O₂. Similar to both normal stem cells and cancer cells, the CRA-type FDCP-Mix cells are highly O₂ independent, as they survive and proliferate in nearly anoxic conditions (0.1% O₂). Further experiments will explore: (1) the metabolic peculiarities of the low-energy proliferation mechanism of Gr-1⁺ FDCP-Mix cells, (2) the relationship of this mechanism to

self-renewal divisions uncoupled to differentiation and (3) the roles of HIF-1, p27^{KIP1} and pRb in the regulation of balance between quiescence and differentiation of FDCP-Mix.

Materials and Methods

Maintenance of the FDCP-Mix cell line in culture. The FDCP-Mix cells (kindly provided by G Mouchiroud; Lyon, France) were routinely replated at 5×10^4 cells/ml every 3 days in Iscove's modified Dulbecco's medium (IMDM, Invitrogen, Carlsbad, CA, USA) complemented with penicillin (50 units/ml), streptomycin (50 μ g/ml), 20% horse serum (HS; Invitrogen), and 10% WEHI conditioned medium (CM) as a source of IL-3. All cultures were carried out at 37°C in a humidified 5% CO₂ incubator (Binder, Tuttlingen, Germany). For experiments, cells in exponential growth at day 3 of culture were centrifuged, washed and resuspended in complete medium containing WEHI CM as required.

Cell proliferation, viability and apoptosis in ambient and low O₂ concentration. FDCP-Mix cells were seeded at 1×10^5 cells/ml and cultured for 24, 48 and 72 h in the same culture medium, either at 20% O₂ in a humidified 5% CO₂ incubator or at 0.1% O₂ concentration in an O₂/CO₂ incubator (Xvivo, Biospherix, Redfield, NY, USA). After culture, viable cells were counted on Malassez Cell with Trypan Blue exclusion. Apoptosis was measured using the Annexin-V-FITC Kit (PN IM3546; Beckman Coulter, Fullerton, CA, USA) following the manufacturer's protocol. Samples were analyzed by FCM with Becton Dickinson FACSCanto II and FACSDiva software (San Jose, CA, USA).

Analysis of cell cycle and divisions by FCM. Cell cycle was analyzed at 24, 48 and 72 h of culture at 20% O₂ and at 0.1% O₂. To distinguish G₀, G₁, S and G₂/M cell cycle phases, cells were double labeled with PI (quantitative DNA labeling) and Ki-67 antibody (absent only in G₀ phase cells). In brief, cells washed in PBS were incubated for 30 min at room temperature (RT) in a PFA/saponin solution (0.4% PFA, 10 mM HEPES, 0.02% saponin and H₂O). Cells were washed two times in 'washing buffer' (5% FBS, 0.05% sodium azide, 55 mg/l EDTA and 0.02% saponin) and incubated for 30 min at RT with an anti-Ki-67 antibody or isotype (Alexa Fluor 647 conjugated anti-Ki-67 no. 558615, and Alexa Fluor 647 conjugated isotype no. 557732; BD Pharmingen, San Diego, CA, USA). Washed cells were resuspended in PBS supplemented with PI (20 μ g/ml) to quantitatively label DNA. Double-labeled cells were then analyzed by FCM using a Becton Dickinson FACSCanto II and FACSDiva software. To know whether G₀ cells were issued from cells previously in G₀ and G₁ phases or from cells in S, G₂ and M phases that divided before entering in G₀, cells were labeled with CFSE (a stable vital cellular fluorescent dye whose decay allows to analyze cell divisions; no. C34554, Invitrogen) according to the manufacturer's instructions. In brief, 1×10^6 FDCP-Mix cells were incubated in the dark for 15 min at 37°C in 1 ml of pre-warmed PBS + 1% BSA and 10 μ M CFSE. Cells were then washed in FDCP-Mix culture medium and incubated for 30 min at 37°C in the dark. After a last wash, cells were resuspended in complete culture medium (3×10^5 cells/ml) and incubated (37°C, 20% O₂, 5% CO₂) for 24 h to eliminate dead cells and use cell populations that are able to proliferate. Cell aliquots were then cultured either at 20 or 0.1% O₂ for 24, 48 or 72 h before being labeled with anti Ki-67 and PI as described and analyzed by FCM.

Cytospin and May-Grünwald/Giemsa staining. 1.5×10^5 cells resuspended in 50 μ l of HS were cytospun at 300 r.p.m. for 20 min using a cytocentrifuge (Shandon CytoSpin III; GMI, Ramsey, MN, USA), and dried and stained with May-Grünwald before microscopical examination at $\times 100$ and $\times 600$ magnification (AX70, Olympus, Tokyo, Japan).

Analysis of cell surface markers and cell sorting. Cells cultured at 20% O₂ and at 0.1% O₂ were washed with washing buffer, incubated for 30 min with fluorescein isothiocyanate- or phycoerythrin-conjugated isotype, anti Mac-1, Gr-1, CD34 and CD117 antibodies (respectively, no. 2225033, no. 22159113, no. 22155243, no. 22150344 and no. 22151173; Immunotools, Friesoythe, Germany). Washed cells were then analyzed by FCM using a Becton Dickinson FACSCanto II and FACSDiva software or sorted according to their Gr-1 expression using a Becton Dickinson FACSARIA.

Analysis of p27^{KIP1} expression in Gr-1⁺ and Gr-1⁺ cells. Cells were double labeled first with anti-Gr-1 (as described in the previous paragraph) and

Figure 6 Gr-1⁺ and Gr-1⁻ FDCP-Mix cells have different morphology, CRA_{cell} and CRA_{CFC} capacities and cell cycle status. After 3 days of LC1 at 20 or 0.1% O₂, cells were sorted according to their absence or presence of the Gr-1 antigen. (a) Gr-1⁻ and Gr-1⁺ cells issued from 20 and 0.1% O₂ LC1 were cytopun and stained with May-Grünwald/Giemsa and observed at $\times 600$ magnification. Arrows (►) indicate cells with donut-shaped nuclei characteristic of terminally differentiated murine granulocytes. (b) Gr-1⁻ and Gr-1⁺ cells issued from 20 and 0.1% O₂ LC1 were serially replated at 20% O₂ every 3 days (LC2 and LC3) and living cells were counted. (c) Gr-1⁻ and Gr-1⁺ cells issued from 20 and 0.1% O₂ LC1 were plated in semisolid culture to measure their CFA. (d) Distribution of the percentage of G₀ cells in the Gr-1⁻ and Gr-1⁺ cell subpopulations issued from 20 and 0.1% O₂ LC1. Values are the mean $\pm$ S.D. of three independent experiments (* $P < 0.001$; Mann-Whitney test)

Figure 7 Time-dependent modifications of pRb during culture in hypoxia. After 24, 48 or 72 h of culture at 20% O₂ or 0.1% O₂, FDCP-Mix cell lysates were analyzed by western blot. (a) Time- and O₂ concentration-dependent expression of pRb, and (b) time- and O₂ concentration-dependent expression of phospho-Ser 608 pRb, phospho-Ser 807/811 pRb and phospho-Thr 821 pRb. Detection of β -actin was used as loading control

then with anti-p27^{KIP1} antibody according mostly to the procedure described for cell cycle analysis. In brief, cells were incubated for 30 min with biotinylated anti-p27^{KIP1} antibody (diluted 1:300, no. MS-256-B; Thermo Scientific, Rockford, IL, USA) instead of anti-Ki-67. Cells were then washed twice with washing buffer before a final 20-min incubation with streptavidin-PE (diluted 1:1000, no. IM0557; Beckman Coulter).

CRA assay. After 72 h of LC at 20% or 0.1% O₂ (LC1), cells were centrifuged for 5 min at 300 g at RT and transferred at 1×10^5 cells/ml in fresh medium into

secondary cultures always incubated at 20% O₂, irrespective of whether LC1 had been incubated at 20% or 0.1% O₂ (nonselective LC2). Cells were then further replated every 3 days and always incubated at 20% O₂ (LC3–LC5). At the end of each LC, cell aliquots were harvested for counting after Trypan Blue exclusion (CRA_{cell}) and colony-forming ability (CFA) assay (CRA_{CFC}).

The CRA assay estimates the maintenance of the stem cell subset contained within a cell population that has undergone a treatment (e.g., incubation in hypoxia) in a selective LC1 by means of cell transfer into nonselective (e.g., normoxic) LC2 and a further incubation therein. LC2 repopulation is then measured by counting the total number of cells (CRA_{cell}) or the number of clonogenic (CFC) cells (CRA_{CFC}) using CFA assay (see below). In normal hematopoietic cell populations, the standard CRA assay has been shown to detect short-term repopulating hematopoietic stem cells.⁴¹

CFA assay. 1×10^3 FDCP-Mix cells were seeded in 250 μ l of methyl-cellulose (Methocult, no. H4230; Stemcell Technologies, Vancouver, Canada) with 10 ng/ml of mIL3 (no. 213–13; Peprotech, Rocky Hill, NJ, USA) in duplicate. Culture dishes were incubated for 11 days at 37°C in a humidified atmosphere with 20% O₂ and 5% CO₂ before counting the number of colonies (aggregates > 50 cells).

Western blot analysis. Cells cultured at 20% O₂ and at 0.1% O₂ were washed in PBS and lysed in loading buffer (2% SDS, 10% glycerol, 50 mM Tris pH 7.8). Protein concentration was determined using the BCA Pierce assay (Thermo Scientific). Then, 40 μ g of whole protein cell extract was resolved in SDS-PAGE and transferred onto a PVDF Membrane (no. RPN303F; GE Healthcare, Uppsala, Sweden) to probe for pRb and p27 proteins, respectively. The membrane was saturated with nonfat milk (5%) in TBS-T (10 mM Tris pH 7.8, 150 mM NaCl and Tween 0.1%) and then incubated overnight at 4°C with specific antibodies to phospho-Ser 608 pRb, phospho-Ser 807/811 pRb, p27^{KIP1} (respectively, no. 2181, no. 9308 and no. 2552; Cell Signaling Technology, Beverly, MA, USA), pRb (no. 554136, BD Pharmingen) or phospho-Thr 821 pRb (no. 44–582, Invitrogen) diluted as indicated by the manufacturer. After three washes in TBS-T, membranes were

Figure 8 Increase in $p27^{KIP1}$ during culture of FDCC-Mix cells at 0.1% O₂ is related to their terminal granulocytic differentiation. **(a)** Time- and O₂ concentration-dependent expression of $p27^{KIP1}$ was analyzed by western blot. Detection of β -actin was used as loading control. **(b)** $p27^{KIP1}$ mRNA level (arbitrary units) was quantified by RT-qPCR after 24, 48 and 72 h of culture at 20% O₂ or at 0.1% O₂. Results are the mean $\pm$ S.D. of four independent experiments performed in duplicates (** $P < 0.005$, * $P < 0.05$; Mann-Whitney test). **(c)** Flow cytometric analysis of the $p27^{KIP1}$ expression in Gr-1⁻ and Gr-1⁺ populations. FDCC-Mix cells were double labeled (anti-Gr-1 and anti- $p27^{KIP1}$ antibodies) after 3 days of culture at 20 and 0.1% O₂. High and low $p27^{KIP1}$ cells were defined as indicated on the flow cytometry histogram (left panel). **(d)** Percentages of Gr-1⁻ and Gr-1⁺ cells with high and low $p27^{KIP1}$ expression after 3 days of culture at 20 and 0.1% O₂. High and low $p27^{KIP1}$ cells were defined as indicated on the flow cytometry histogram (left panel). The right panel graph shows the mean percentage $\pm$ S.D. of four independent experiments

incubated for 1 h with anti-rabbit or anti-mouse horseradish peroxidase-conjugated antibodies. The bands were visualized using the ECL system (no. WBKLS; Millipore, Billerica, MA, USA). The membrane was washed again, saturated, stained with an anti- β -actin antibody (no. A5316; Sigma, St Louis, MO, USA) and finally revealed as described above.

Reverse transcription and quantitative PCR. Cells cultured at 20% O₂ and at 0.1% O₂ were washed in PBS and treated with 250 μ l Trizol (no. 15596-026; Invitrogen). Total RNA extraction, DNase treatment (no. M6101; Promega, Madison, WI, USA) and reverse transcription with M-MLV Reverse Transcriptase (no. M1705, Promega) were realized according to the manufacturer's instructions. Samples were run three times in duplicates using the brilliant SYBR Green QPCR (no. 600548; Stratagene, La Jolla, CA, USA) in the MX4000 apparatus (Stratagene) according to the manufacturer's instruction. The real-time PCR reactions were carried out in the presence of mouse $p27^{KIP1}$ gene-specific oligonucleotide primers (forward primer: 5'-AGGCGGTGCCTTTAATTGGG-3', reverse primer: 5'-TTA CGTCTGGCGTCGAAGGC-3') or 36B4 gene-specific oligonucleotide primers (forward primer: 5'-GCTTCATTGTGGGAGCAGAC-3' and reverse primer: 5'-CA TGGTGTCTTGCCCATCAG-3') as follows: 10 min activation at 95°C, followed by 40 cycles at 95°C for 30 s, 58°C for 1 min and 72°C for 1 min.

A relative quantification of $p27^{KIP1}$ transcript expression was obtained by standardizing to 36B4 levels that was shown to be stable in hypoxia.⁴² The final value was calculated by taking an average of relative transcript copy numbers amplified from the three sets of PCR reaction performed in duplicates.

Statistics. All data are presented as mean $\pm$ S.D. of 3–7 independent experiments. The Friedman or Kruskal–Wallis tests were, respectively, used to compare >2 experimental conditions paired or not. For statistical analysis of differences between two not paired experimental conditions, Mann–Whitney test was used.

Conflict of interest

The authors declare no conflict of interest.

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