

**Keywords:** DJ-1 (PARK7); breast cancer cells; invasion; epithelial–mesenchymal transition (EMT); KLF17; ID-1; Ras

# DJ-1 upregulates breast cancer cell invasion by repressing KLF17 expression

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**Background:** DJ-1 (PARK7) was reported as an oncogene in a Ras-dependent manner. Recent studies have shown that DJ-1 stimulates cell proliferation, cell invasion, and cancer metastasis. However, the molecular mechanism by which DJ-1 induces cancer cell invasion and metastasis remains unclear.

**Methods:** Breast cancer cells were transfected with DJ-1 siRNA or DJ-1 overexpression to investigate the effect of DJ-1 on KLF17 expression. ID-1 luciferase promoter assay was performed to evaluate DJ-1-dependent KLF17 expression changes. In addition, Epistasis analysis of DJ-1 and KLF17 was performed to evaluate their regulatory interactions. Ras inhibitors were pretreated to determine whether DJ-1 regulates cell invasion in a Ras-dependent manner.

**Results:** In the present study, we found increased DJ-1 expression in highly invasive breast cancer cells as compared with non-metastatic cells. Furthermore, DJ-1 promoted breast cancer cell invasion by downregulating E-cadherin and increasing Snail expression. Interestingly, exogenous DJ-1 overexpression markedly decreased mRNA and protein expression of KLF17, the EMT negative regulator. These data were confirmed by ID-1 promoter activity, which is directly regulated by DJ-1-dependent KLF17 transcription factor. Epistasis analysis showed that KLF17 overexpression overcomes increased cell invasion by DJ-1, suggesting that KLF17 might be one of the downstream signalling molecules of DJ-1. Acceleration of cell invasion by DJ-1 was alleviated by Ras inhibitors, suggesting that DJ-1 cooperates with Ras to increase cell invasion.

**Conclusion:** Altogether, these data suggest for the first time that DJ-1 acts as an EMT-positive regulator in breast cancer cells via regulation of the KLF17/ID-1 pathway.

DJ-1, originally identified as a novel oncogene product involved in a Ras-related signal transduction pathway (Nagakubo *et al*, 1997), belongs to the peptidase C56 family. The human DJ-1 gene maps to 1p36.2-36.3, a locus for many chromosome aberrations in cancers (Taira *et al*, 2001). It is generally known that loss-of-function *DJ-1* mutations can cause early-onset Parkinson's disease. Mitochondrial membrane perturbation, oxidative stress, and proteasome dysfunction are among the several hypotheses suggested to explain the molecular basis of neuronal damage (Dawson and Dawson, 2003). DJ-1 protects several kinds of cancer cells such as pancreatic (Inberg and Linial, 2010), neuronal (Yokota *et al*, 2003; Zhang *et al*, 2010), leukaemic (Liu *et al*, 2008), lung (MacKeigan *et al*, 2003), and breast (Ismail *et al*, 2012) against

oxidative stress-induced apoptosis. DJ-1 is reported to modulate the PTEN/Akt survival pathway inactivation (Kim *et al*, 2005). In addition, DJ-1 binds multiple RNA targets, implicating involvement in multiple cellular pathways (van der Brug *et al*, 2008).

The prognostic significance of DJ-1 has been studied in breast cancer, in which the serum level of DJ-1 is increased compared with healthy subject (Le Naour *et al*, 2001). In breast cancer, an increased expression of cytoplasmic DJ-1 was also observed (Kim *et al*, 2005). A previous study showed that DJ-1 is associated with endometriotic cell migration as well as invasion potential (Rai and Shivaji, 2011). Moreover, there was a positive association between higher DJ-1 levels and disease recurrence in stage I lung cancer (Kim *et al*, 2005). Bai *et al* (2012) also showed that DJ-1 expression

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is significantly correlated with lung cancer lymphatic metastasis. He *et al* (2012) represented that DJ-1 regulates the invasion and metastasis of pancreatic cells by activating the SRK/ERK/uPA cascade. They showed that knockdown of DJ-1 led to cytoskeleton disruption and diminished uPA activity and expression, all these effects being reversed by restoration of DJ-1 expression (He *et al*, 2012). Interestingly, DJ-1 was validated as a MMP substrate (Dean *et al*, 2007) and identified as being secreted (Butler *et al*, 2008), suggesting roles in both intracellular and extracellular microenvironments.

Cancer metastases, rather than primary tumours, are responsible for most of the cancer deaths (Steege, 2006; Eccles and Welch, 2007; Steeg, 2007). Epithelial-mesenchymal transition (EMT) is a cellular program in which polarised epithelial cells undergo complex biological changes such that the epithelial cells express a mesenchymal phenotype, which induces enhanced migratory capacity, invasiveness, metastatic potential, and drug resistance (Zhou *et al*, 2009). A previous study showed that DJ-1 is upregulated in renal fibrosis and mediates EMT by suppressing cytoplasmic PTEN expression (Yao *et al*, 2011). However, the molecular mechanism by which DJ-1 inhibits PTEN expression remains unclear.

KLF17 (Krüppel-like transcription factor 17; ZNF393) is a member of the Sp/KLF zinc finger protein family with diverse functions (Lomberk and Urrutia, 2005; van Vliet *et al*, 2006). Krüppel-like factor (KLF) family members are critical regulators of various cellular processes, including reprogramming of differentiated cells to stem cells (Takahashi *et al*, 2007), cell survival (Perkins *et al*, 1995), and tumour development (Foster *et al*, 2000; Ghaleb *et al*, 2005). Interestingly, KLF17 binds to the DNA consensus sequence (5'-CACCC-3') to inhibit ID-1 transcription (van Vliet *et al*, 2006), resulting in the inhibition of EMT and breast cancer cell invasion (Gumireddy *et al*, 2009). ID-1 is a novel negative regulator of PTEN that could activate the Akt pathway and its downstream effectors such as the Wnt/TCF pathway (Lee *et al*, 2009), suggesting that the oncogenic function of ID-1 may be partly attributed to its PTEN inhibition in human breast carcinogenesis. ID-1 is overexpressed in highly invasive cancer cells, including prostate (Zhang *et al*, 2007), breast (Fong *et al*, 2003), cervical (Darnel *et al*, 2010), and bladder cancer (Ding *et al*, 2006). It has been suggested that the combined expression patterns of KLF17 and ID-1 can serve as a potential biomarker for lymph node metastasis in breast cancer (Gumireddy *et al*, 2009).

In the present study, we tried to evaluate the molecular mechanism of DJ-1 protein underlying the EMT status and *in vitro* cell invasion of breast cancer cells. In addition, we also studied whether Ras is involved in DJ-1-induced cell invasion.

## MATERIALS AND METHODS

**Chemicals and reagents.** MTT (3-[4,5-dimethyl-2-thiazolyl]-2,5-diphenyl-2H-tetrazolium bromide) was purchased from Sigma (St. Louis, MO, USA). DMEM, RPMI, FBS and penicillin/streptomycin antibiotics were purchased from Gibco (Invitrogen, Carlsbad, CA, USA). Qiazol was purchased from Qiagen (Valencia, CA, USA), and 2 × SYBR Green PCR master mix was purchased from Takara Biotechnology (Dalian, Japan). The CytoSelect 96-well cell invasion assay kit was purchased from Cell Biolabs (San Diego, CA, USA). Rabbit polyclonal anti-DJ-1, anti-Snail, anti-KLF17, and rabbit monoclonal anti-E-cadherin antibodies were purchased from Abcam (Cambridge, UK). Mouse monoclonal anti-ID-1 antibody was purchased from Millipore (Upstate Chemicon, Temecula, CA, USA). Goat polyclonal anti-occludin, anti-fibronectin, and mouse monoclonal IgG HRP-conjugated anti-β-actin antibody was purchased from Santa Cruz Biotechnology (Santa Cruz, CA,

USA). HRP-conjugated goat anti-rabbit and anti-mouse secondary antibodies were purchased from Pierce (Rockford, IL, USA). ECL Western Blotting Detection Reagent was purchased from Neuro-nex (Daegu, South Korea). Ras inhibitors such as PD98059 and LY294002 were purchased from Sigma. All other reagents were obtained from standard commercial sources.

**Cell culture.** The human breast cancer cell lines MCF-7, T47D, MDA-MB-231, and MDA-MB-435 were obtained from the American Type Culture Collection and maintained in DMEM (MCF-7 and MDA-MB-231) or RPMI (T47D and MDA-MB-435) media containing 10% FBS and 1% penicillin/streptomycin solution. Cells were grown at 37 °C in a humidified atmosphere containing 5% CO<sub>2</sub>. Although the origin of the MDA-MB-435 cell line has been disputed, it has many properties similar to those of other aggressive breast cancer cell lines (Chambers, 2009).

**Overexpression or knockdown of genes.** MCF-7, MDA-MB-231, and MDA-MB-435 cells were transfected with mixed DJ-1 siRNA (Santa Cruz Biotechnology). Briefly, 1 × 10<sup>5</sup> cells were seeded in 60-mm plates, media being replaced on the following day with serum-free media just before transfection. Cells were then transfected with siRNA at a final concentration of 10 nM using Lipofectamine 2000 (Invitrogen) as a transfecting reagent in Opti-MEM (Invitrogen). For exogenous DJ-1 overexpression, pc3DNA-flag-DJ-1 vector (provided by Dr. Jong-In Yook at Yonsei University, Seoul, Korea) or pc3DNA control vector were transfected under the same experimental conditions. For exogenous KLF17 overexpression, pCMV6-AC-KLF17-GFP (Origene, Rockville, MD, USA) or pCMV6 control vector were transfected under the same experimental conditions. After 8 h, serum-free media containing the transfection mixture was replaced with fresh serum-containing media and incubated for the indicated periods. Cells were collected after 8–48 h of transfection and used for cell invasion assay, chromatin immunoprecipitation (ChIP) assay, real-time qPCR, and western blotting analyses.

**In vitro cell invasion assay.** The effect of DJ-1 expression on breast cancer cell invasion was determined using the CytoSelect 96-well cell invasion assay kit (Cell Biolabs) containing polycarbonate membrane inserts (8-μm pore size) as indicated in the instruction manual and previously described (Cabello *et al*, 2009). The basement membrane layer was rehydrated by adding 100 μl of warm, serum-free medium to the inner compartment and incubated for 1 h in a cell culture incubator. A cell suspension (1 × 10<sup>5</sup> per ml) in serum-free medium was cultured on the basement membrane after removing the rehydration media. In all, 150 μl of serum-containing medium was added to the feeder tray. After insertion of the basement membrane chamber, 100 μl of cell suspension was added. After 24 h of incubation, the medium in the membrane chamber was transferred to a new harvesting tray containing 150 μl of detachment solution for 30 min. The cells were dislodged completely from the underside of the membrane by gently tilting the membrane several times. Fifty microlitres of 4 × lysis buffer/CyQuant GR dye solution was added to all samples, which were then incubated for 20 min at room temperature. Fluorescence measurements were performed in a fluorescence plate reader at 480 nm/520 nm.

To evaluate the effect of Ras inhibitors on DJ-1-induced cell invasion, control vector- or DJ-1-Flag-transfected cells were seeded onto matrigel-coated transwells in matrigel invasion chambers (BD Biosciences, Franklin Lakes, NJ, USA). Then, a specific inhibitor of PI3K (LY294002, 10 μM) or BRAF/MEK inhibitor (PD98059, 25 μM) was treated to the lower well. After 48 h, non-invading cells were removed from the top well with a cotton swab while the bottom cells were fixed and stained with a 0.2% solution of crystal violet in 2% ethanol. After washing, the crystal violet stain was

solubilised with a 10% acetic acid solution, and cell invasion was quantitated by measuring the absorbance at 595 nm.

**Quantitative real-time PCR.** The effect of DJ-1 expression on a panel of genes involved in invasion and EMT regulation in breast cancer cells was investigated using real-time quantitative RT-PCR analysis. In brief,  $1 \times 10^5$  cells were plated in 60-mm dishes and grown for 24 h, followed by transfection with DJ-1 siRNA, DJ-1-Flag, or KLF17 overexpression vector for 48 h as indicated. Total RNA was extracted with Qiazol (Qiagen) according to the manufacturer's instructions. Extracted RNA (5  $\mu$ g) was reverse transcribed into cDNA using a first-strand cDNA synthesis kit (Applied Biosystems, Foster City, CA, USA), and the resulting cDNA was diluted 10-fold and kept at  $-20^\circ\text{C}$  until use. The real-time qPCR primers were designed using the Primer Express 1.5 software (Applied Biosystems) as follows: DJ-1 forward, 5'-GTCAT TTGTCCTGATGCCAGC-3', reverse, 5'-TCAGATAAATTCTGT GCGCCC-3'; E-cadherin forward, 5'-CGACCCAACCCAAGA ATCTA-3', reverse 5'-CTCCAAGAAATCCCCAGAATG-3'; Snail forward, 5'-AGCTCTCTGAGGCCAAGGATCT-3', reverse 5'-TG TGGCTTCGGATGTGCAT-3'; occludin forward 5'-CGGTCTAG GACGCAGCAGAT-3', reverse 5'-AAGAGGCCTGGATGACA TGG-3'; fibronectin forward 5'-CAGGATCACTTACGGAGAAA CAG-3', reverse 5'-GCCAGTGACAGCAACACAGTG-3'; KLF17 forward, 5'-CTGCCTGAGCGTGGTATGAG-3', reverse 5'-TCAT CCGGAAGGAGTGAGA-3'; ID-1 forward, 5'-CTCTACGACAT GAACGGCTG-3', reverse 5'-TGTCACCTTGCGGTTCTG-3'; PTEN forward 5'-TCAGTGGCGGAACCTTGCAA-3', reverse 5'-CA TGAACCTGTCTCCCGTCG-3'; and GAPDH forward, 5'-AGA TCATCAGCAATGCCTCCTG-3', reverse 5'-ATGGCATGGACT GTGGTCATG-3'. The expressions of these genes were normalised to GAPDH. Real-time PCR was carried out using an ABI Prism 7500 sequence detection system (Applied Biosystems). Each 20  $\mu$ l PCR reaction contained 10  $\mu$ l SYBR Green PCR master mix, 4  $\mu$ l diluted cDNA, and 200 nM primer set. All samples were amplified in triplicate in a 96-well plate using the following cycling conditions: 2 min at  $50^\circ\text{C}$ , 10 min at  $95^\circ\text{C}$ , and 40 cycles at  $95^\circ\text{C}$  for 15 s followed by 1 min at  $60^\circ\text{C}$ . Calculations were performed using the  $\Delta\text{Ct}$  method, normalising the average Ct value of each treatment compared with its endogenous control (GAPDH) and then calculating the  $2^{-\Delta\text{Ct}}$  for each treatment. These experiments were repeated three times.

**Western blotting analyses.** Cells were washed twice with cold PBS, after which 200  $\mu$ l of PRO-PREP protein extraction solution (Intron, Daejeon, South Korea) was added. The cell lysates were centrifuged and protein concentrations were estimated using the Coomassie protein assay reagent (Thermo Scientific, Rockford, IL, USA). Protein samples (30–40  $\mu$ g) were electrophoresed on 8%–15% SDS-PAGE gels. Proteins were transferred to nitrocellulose membranes blocked in 5% skim milk in TBS (25 mM Tris base and 150 mM NaCl) for 2 h at room temperature and then incubated with primary antibody overnight at  $4^\circ\text{C}$ . The membranes were then incubated with HRP-conjugated secondary antibodies at 1:5000 dilutions for 1 h at room temperature and then washed three times in TBST (TBS and 0.1% Tween 20).  $\beta$ -Actin was used as a reference protein for the normalisation of protein contents. The target proteins were detected with ECL detection reagents, and the relative intensities of the bands were analysed by the Image-J software (U.S. NIH, Bethesda, MD, USA).

**ChIP assay.** The effect of DJ-1 expression on ID-1 promoter activity regulation through KLF17 was investigated using ChIP assay as previously described (Gumireddy *et al.*, 2009). Briefly, Lipofectamine 2000 control and transiently-transfected MDA-MB-231 cells were fixed in 1% paraformaldehyde at  $37^\circ\text{C}$  for 10 min. After washing twice with ice-cold PBS containing protease inhibitor cocktail, cells were collected in lysis buffer (1% SDS,

10 mM EDTA, and 50 mM Tris, pH 8.1), incubated for 30 min on ice, and then sonicated to shear DNA. The lysates were then centrifuged, and the supernatants were diluted 10-fold with ChIP dilution buffer (0.01% SDS, 1% Triton X-100, 2 mM EDTA, 20 mM Tris-HCl at pH 8.0, 150 mM NaCl, and protease inhibitors). Rabbit anti-KLF17 or normal rabbit IgG were added to the supernatants and incubated overnight at  $4^\circ\text{C}$  with mild rotation. The immunocomplex was incubated with protein G-agarose, then washed with low-salt buffer, high-salt buffer, and lithium chloride wash buffer, and eluted with elution buffer (1% SDS, 0.1 M  $\text{NaHCO}_3$ , and 200 mM NaCl). Reversal of crosslinking was done by heating at  $65^\circ\text{C}$  overnight in the presence of NaCl. DNA was purified using PureLink PCR Purification kit (Invitrogen). The amount of immunoprecipitated DNA was analysed in triplicate using human ID-1 (sense: 5'-AGCTTGTGAGAGGCAGTGAT TCT-3'; antisense: 5'-GGAACCCAGAGGTAGGCTGTT-3') promoter primer sequences by real-time PCR with SYBR green PCR kit. Data were analysed using the  $2^{-\Delta\text{Ct}}$  method and normalised with input (HDAC1) samples.

**Statistical analysis.** The differences in mean values among the groups were evaluated and expressed as the means  $\pm$  s.d. Averages were drawn, and statistical calculations were then performed using a Student's *t*-test in Microsoft Excel 2010 (Redmond, WA, USA).

## RESULTS

**DJ-1 expression in breast cancer cells.** To demonstrate the relation between DJ-1 expression and breast cancer cell invasion capacity, we investigated DJ-1 mRNA and protein expression levels in non-metastatic (MCF-7 and T47D) and metastatic (MDA-MB-231 and MDA-MB-435) cancer cells. Interestingly, the expression levels of DJ-1 mRNA (Figure 1A) and protein (Figure 1B) are higher in the invasive breast cancer cells compared with non-metastatic cells.

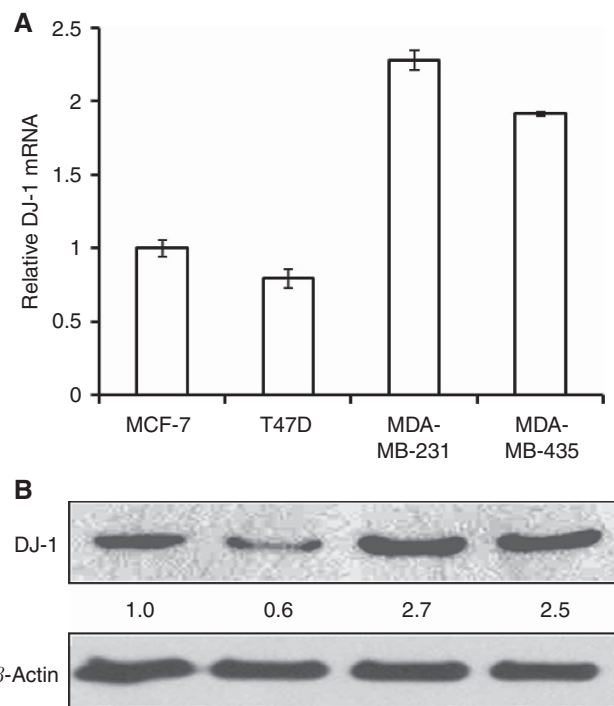


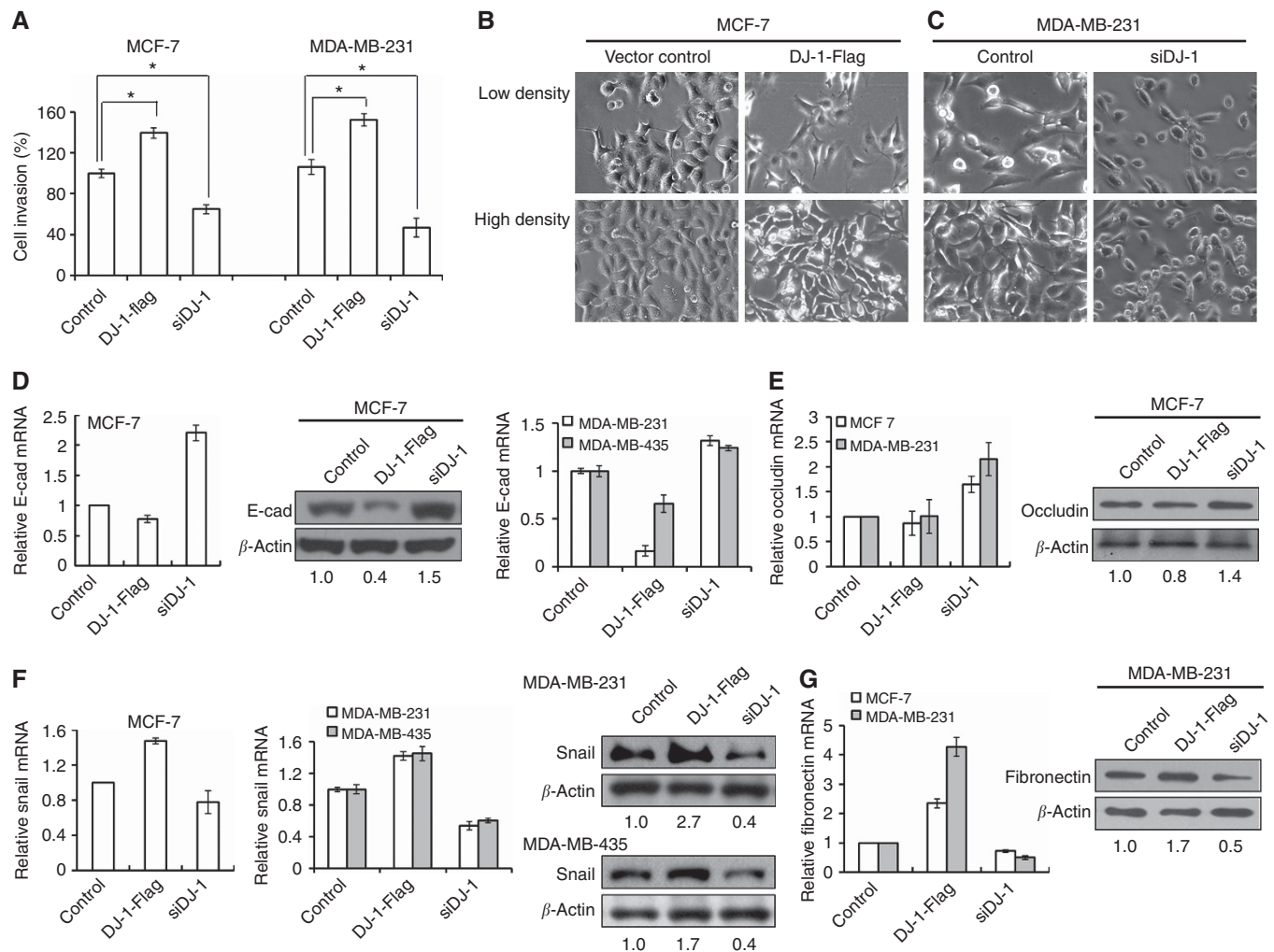
Figure 1. Expression of DJ-1 mRNA (A) and protein (B) in non-metastatic (MCF-7, T47D) and metastatic (MDA-MB-231, MDA-MB-435) breast cancer cells. Real-time qPCR data are from three independent experiments; each line represents s.d.

**Effect of DJ-1 knockdown or overexpression on the EMT of breast cancer cells.** E-cadherin is one of the most important molecules in cell–cell adhesion in epithelial tissues, a hallmark of EMT being the loss of E-cadherin expression. We investigated the effect of DJ-1 on epithelial markers (E-cadherin, occludin) and mesenchymal markers (Snail, fibronectin) in breast cancer cells. DJ-1 mRNA and protein expression levels were confirmed in DJ-1-Flag- or siDJ-1-transfected cells (Supplementary Figure S1). Interestingly, exogenous DJ-1 overexpression increased the invasion capacity, and DJ-1 knockdown by siRNA treatment downregulated cell invasion of MCF-7 and MDA-MB-231 cells as compared with vector-treated cells (Figure 2A).

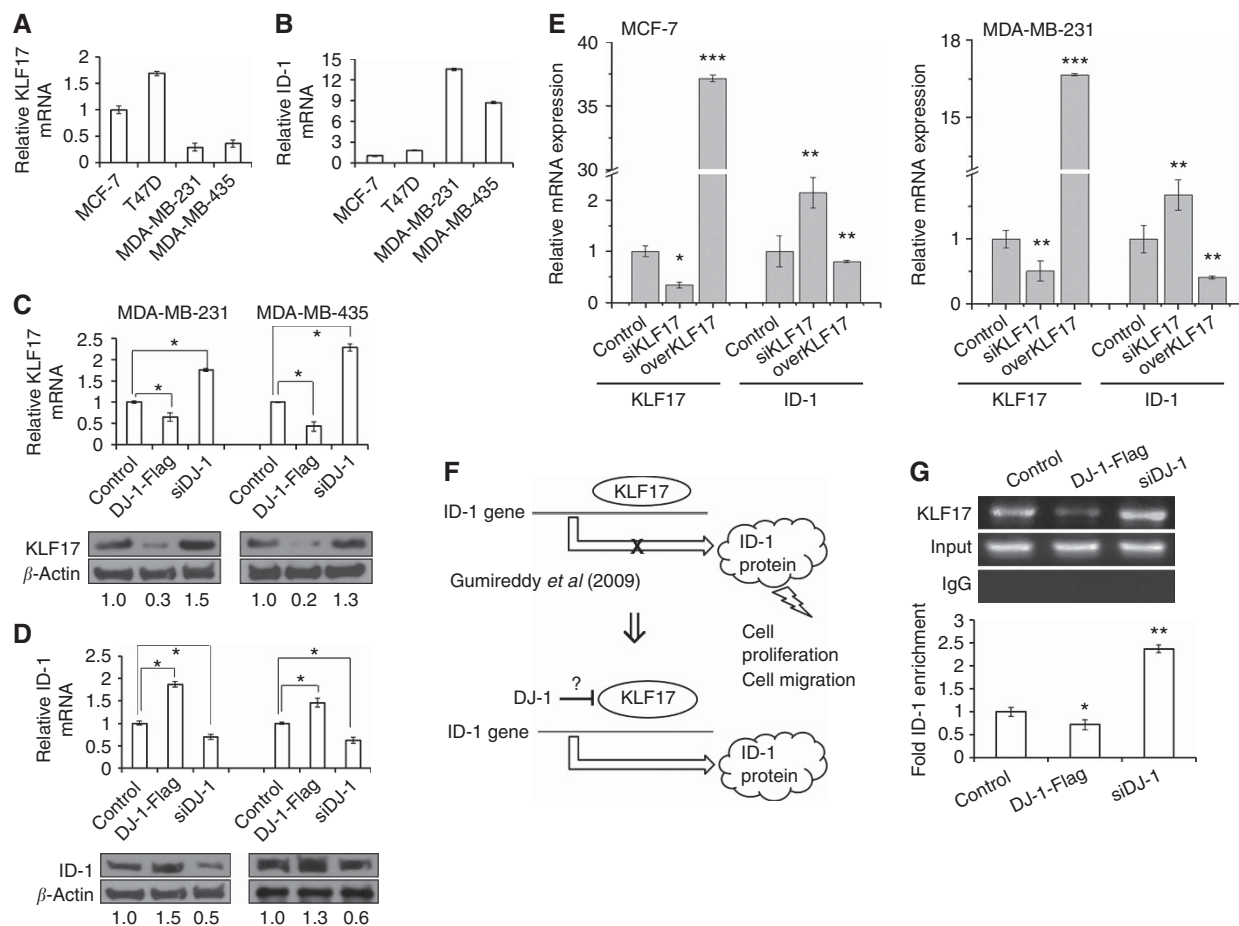
To evaluate the EMT status in relation to DJ-1 expression level, morphological changes were assessed by light microscopy. As shown in Figure 2B, MCF-7 cells overexpressing DJ-1 lost their observable epithelial morphology after 48 h of DJ-1-Flag transfection. On the contrary, siDJ-1-treated MDA-MB-231 cells gained epithelial-like morphology, unlike control. To further confirm EMT by DJ-1, the expression of epithelial and mesenchymal markers was investigated by real-time qPCR and/or western blotting. The downregulation of E-cadherin mRNA and/or protein levels was observed in cells transfected with DJ-1-Flag (Figure 2D).

On the contrary, siDJ-1 transfection increased E-cadherin mRNA and/or protein expression. Occludin mRNA and protein expression was also increased in siDJ-1-transfected cells. There was no remarkable change of occludin expression in DJ-1-Flag-transfected cells (Figure 2E). Conversely, Snail mRNA and/or protein expression levels were upregulated in DJ-1-Flag-transfected cells, whereas siDJ-1 decreased Snail mRNA and/or protein expression levels in breast cancer cells (Figure 2F). The fibronectin mRNA and protein expression patterns were similar to that of Snail in MCF-7 and MDA-MB-231 cells (Figure 2G).

**Effect of DJ-1 and KLF17 on ID-1 gene expression.** The mRNA expression level of KLF17 is higher in non-metastatic breast cancer cells as compared with invasive cells (Figure 3A), ID-1 mRNA expression showing the opposite pattern as compared with KLF17 (Figure 3B). Interestingly, DJ-1-Flag transfection decreased the expression level of KLF17 mRNA and protein, whereas DJ-1 knockdown by siRNA increased KLF17 expression (Figure 3C). On the contrary, DJ-1-Flag and siDJ-1 transfection caused ID-1 expression to increase and decrease, respectively (Figure 3D). To evaluate the effect of KLF17 on ID-1 expression in MCF-7 and MDA-MB-231 cells, siKLF17 or KLF17 overexpression vector was



**Figure 2.** Effect of DJ-1 overexpression and knockdown on cell invasion and EMT of breast cancer cells. (A) For cell invasion assay, DJ-1-Flag or siDJ-1 was transfected for 12 h, 48 h after which transwell invasion assay was performed. Statistical analysis was performed using the Student's *t*-test (\* $P < 0.05$ ). After transfection with DJ-1-Flag or siDJ-1 for 48 h in (B) MCF-7 and (C) MDA-MB-231 cells, respectively, cellular morphological change was evaluated by light microscope. mRNA and/or protein expression of (D, E) epithelial (E-cadherin, occludin) and (F, G) mesenchymal markers (Snail, fibronectin) were analysed by real-time qPCR or western blotting analysis after 24–48 h of transfection. These results are from three independent experiments; each line represents s.d.



**Figure 3.** The effect of DJ-1 overexpression and knockdown on KLF17 and ID-1 expression in breast cancer cells. The basal mRNA expression levels of (A) KLF17 and (B) ID-1 were analysed. MDA-MB-231 and MDA-MB-435 cells were transfected with DJ-1-Flag or siDJ-1 for 24–48 h and mRNA and protein expression levels of (C) KLF17 and (D) ID-1 were analysed using real-time qPCR and western blotting analyses. To evaluate the effect of KLF17 on ID-1 mRNA expression, pCMV6-AC-KLF17-GFP or siKLF17 was transfected for 24 h, after which (E) ID-1 expression was analysed by real-time qPCR. The effect of DJ-1 on KLF17-dependent ID-1 expression (F) was confirmed by ChIP assay. MDA-MB-231 cells were transfected with either DJ-1-Flag or siDJ-1; KLF17 binding to the ID-1 promoter at CACCC nucleotide sequence was then investigated using KLF17 antibody. (G) RT-PCR and real-time qPCR analyses were performed. Input HDAC1 and IgG were used as positive and negative controls, respectively. The statistical significances were evaluated using the Student's *t*-test (\* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ ). These results are from three independent experiments; each line represents s.d.

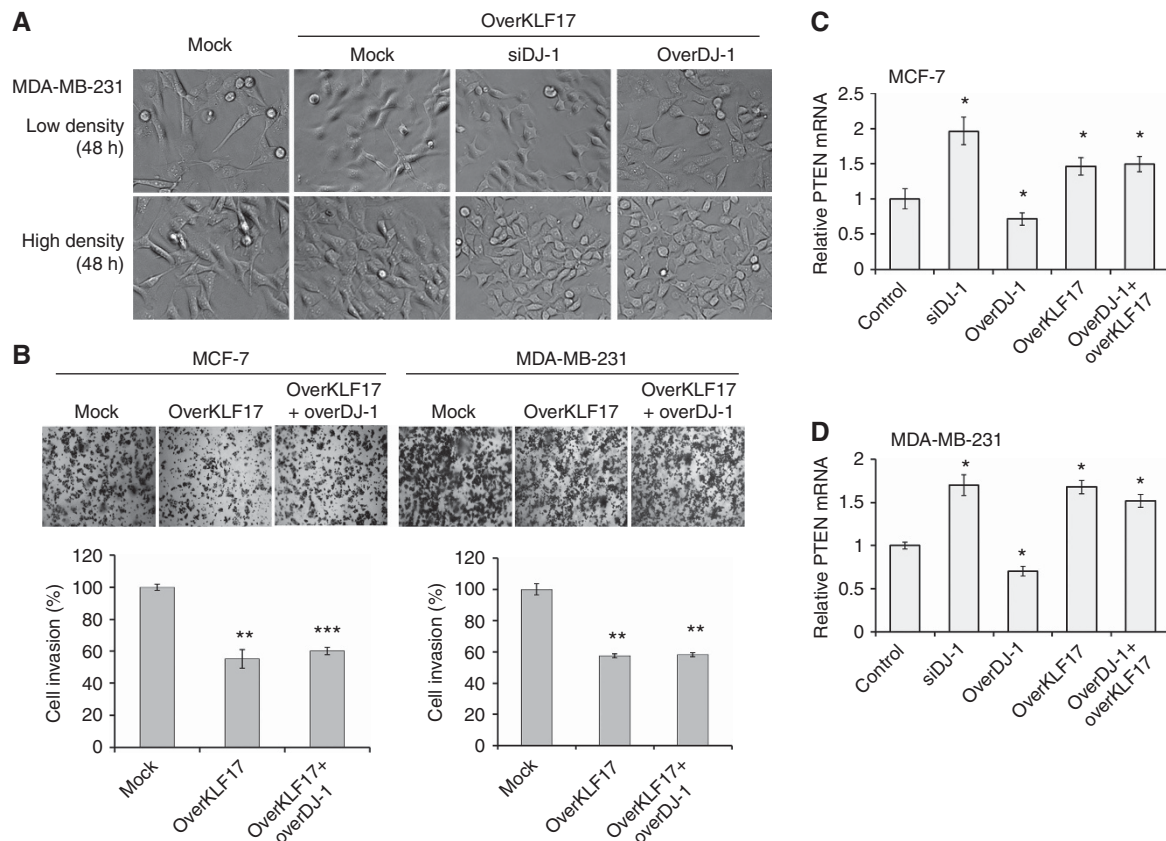
transfected. As shown in Figure 3E, transient KLF17 knockdown increased ID-1 mRNA expression, whereas exogenous KLF17 overexpression inhibited ID-1 gene expression (Figure 3E).

A previous study showed that KLF17 binds directly to the DNA consensus sequence (5'-CACCC-3') of ID-1 promoter to inhibit its transcription (Gumireddy *et al*, 2009). Therefore, ChIP analysis was performed with KLF17 antibody to evaluate whether DJ-1 expression level affects KLF17 binding to the CACCC sequence of ID-1 promoter (Figure 3F). KLF17 mRNA expression was evaluated by RT-PCR, and input and IgG were used as positive and negative controls, respectively. As shown in Figure 3G, DJ-1-Flag transfection decreased KLF17 binding to ID-1 promoter ( $P < 0.05$ ), whereas siDJ-1 transfection upregulated KLF17 binding to ID-1 promoter ( $P < 0.01$ ). The results revealed that DJ-1 regulates KLF17 binding to the ID-1 promoter.

**Epistasis analysis of DJ-1 and KLF17 on the cellular morphology and invasiveness.** To further evaluate that DJ-1 regulates breast cancer invasion and EMT in part through KLF17, we performed double overexpression of DJ-1 and KLF17 in cancer cells. As shown in Figure 4A, MDA-MB-231 cellular morphological changes, KLF17 overexpression showed similar results independent of DJ-1 expression level. When we performed matrigel invasion

assay at the same condition, KLF17 overexpression inhibits cell invasion significantly in both the cells ( $P < 0.005$ ) (Figure 4B). In addition, co-overexpression of KLF17 with DJ-1 results in a similar invasiveness with that of KLF17 single overexpression. To further confirm the effect of DJ-1 and KLF17 on PTEN gene expression, we investigated PTEN mRNA expression by real-time qPCR. DJ-1 knockdown or KLF17 overexpression increased PTEN mRNA expression significantly in both the cells ( $P < 0.05$ ) (Figures 4C and D). However, exogenous DJ-1 overexpression downregulated PTEN mRNA expression, and double overexpression with KLF17 overcomes PTEN inhibition by DJ-1. These data suggest that KLF17 might be one of the downstream signalling proteins of DJ-1.

**Effect of Ras inhibitor on DJ-1-enhanced cell invasion.** It has been shown that DJ-1 cooperates with oncogenic Ras to promote metastasis (Moon *et al*, 2006). Therefore, we examined the role of Ras/Raf/MEK or PI3K/Akt in cell invasion when DJ-1 is overexpressed. DJ-1-Flag overexpression vector was transfected in MCF-7 and MDA-MB-231 bearing a PI3K and BRAF/MEK mutation, respectively, with or without specific Ras inhibitor pretreatment. LY294002 (a specific inhibitor of PI3K) or PD98059 (a specific inhibitor of MEK, which is an immediate downstream kinase target of BRAF) were used to evaluate the effect of Ras in these two cell lines.



**Figure 4.** Epistatic analysis of DJ-1 and KLF17 on cellular morphology, invasion, and PTEN mRNA expression. **(A)** pCMV6-AC-KLF17-GFP was co-transfected with DJ-1-Flag or siDJ-1 for 48 h, after which cellular morphology was analysed with phase-contrast microscope. **(B)** For cell invasion assay, KLF17 overexpression vector was co-transfected with DJ-1-Flag for 12 h, 48 h after which transwell invasion assay was performed. **(C, D)** PTEN mRNA expression was analysed by real-time qPCR after transfection with DJ-1-Flag, siDJ-1, and/or KLF17 overexpression vector. Statistical analysis was performed using the Student's *t*-test (\* $P < 0.05$ , \*\* $P < 0.005$ , \*\*\* $P < 0.001$ ). These results are from two or three independent experiments; each line represents s.d.

First, we evaluated the effect of PD98059 on morphological change due to exogenous DJ-1 overexpression in MDA-MB-231 cells. As shown in Figure 5A, there was no remarkable morphological change in control vector-transfected cells. In DJ-1-Flag transfected cells, however, PD98059 treatment alleviated the mesenchymal morphological change induced by DJ-1 overexpression. To further confirm the role of PI3K or MEK activation in DJ-1-promoted cell invasion, we treated the cells with LY294002 or PD98059 and measured cell invasion in DJ-1-Flag- or control vector-transfected cells. In control vector-transfected cells, LY294002 or PD98059 treatment induced no significant change in MCF-7 and MDA-MB-231 cells' invasion capacity, respectively (Figure 5B). In DJ-1-Flag-transfected cells, however, each Ras inhibitor alleviated DJ-1-promoted cell invasion significantly in both the cells (Figure 5B). We also evaluated Snail protein expression under the same experimental condition. There was no remarkable change in Snail protein expression due to Ras inhibitors in control vector-transfected cells. However, in DJ-1-Flag-transfected cells, LY294002 or PD98059 treatment decreased Snail expression remarkably (Figure 5C).

## DISCUSSION

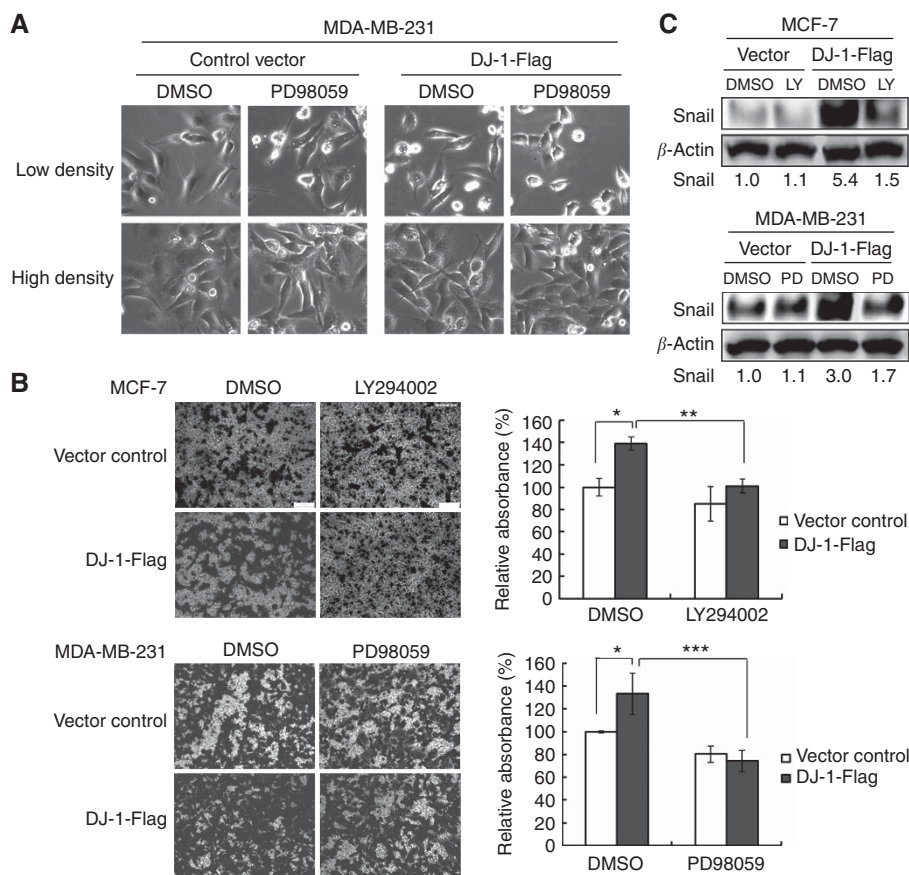
Lee *et al* (2009) showed that ID-1 negatively regulated PTEN at the transcriptional level, and this led to the Akt-mediated canonical Wnt signalling pathway, which may be partly attributed in human

breast carcinogenesis. KLF17 is able to negatively regulate EMT and cell invasion by directly binding to the ID-1 promoter, and KLF17/ID-1 reciprocal expression is a critical pathway in the development of breast cancer metastasis (Gumireddy *et al*, 2009), indicating that regulation of this pathway may serve as a basis for the development of therapies that target early metastatic events.

Kim *et al* (2005) reported that DJ-1 inhibits PTEN/Akt survival pathway inactivation in breast cancer, and this has been supported by their results that DJ-1 expression correlates negatively with PTEN immunoreactivity and positively with PKB/Akt hyperphosphorylation. However, little has been known about how DJ-1 negatively regulates PTEN gene expression.

ID protein (inhibitor of differentiation or DNA binding) is a group of helix-loop-helix (HLH) proteins unable to directly bind DNA. They act as dominant-negative regulators of basic HLH (bHLH) transcription factors via heterodimerisation. As bHLH proteins are involved in tissue-specific differentiation, aberrant ID expression may interfere with cellular differentiation and growth programmes of a wide variety of cell types (Norton, 2000). Upregulation of ID-1 has been found in many types of human cancer, including breast (Lin *et al*, 2000). In addition, overexpression of ID-1 has been indicated as a marker for unfavourable prognosis in breast cancer (Ling *et al*, 2005), and the level of ID-1 protein expression is correlated with a more aggressive clinical behaviour of ovarian and prostate carcinomas (Schindl *et al*, 2003; Schoppmann *et al*, 2003).

Our data showed that DJ-1 accelerates EMT and cell invasion capacity by downregulating E-cadherin and increasing Snail



**Figure 5.** The effect of Ras inhibitors on DJ-1-promoted cell invasion. **(A)** Morphological change was analysed in MDA-MB-231 cells after control vector or DJ-1-Flag transfection for 48 h with or without PD98059 (a specific inhibitor of MEK, which is an immediate downstream kinase target of BRAF). **(B)** Cell invasion was analysed in matrigel-coated transwell. After 48 h of DJ-1-Flag transfection with or without PD98059 (a specific inhibitor of BRAF-MEK) or LY294002 (a specific inhibitor of PI3K), the bottom cells were fixed and stained with a 0.2% solution of crystal violet in 2% ethanol for light microscope image analysis. After washing, the crystal violet stain was solubilised with 10% acetic acid solution, and the cell invasion was quantitated by measuring the absorbance at 595 nm. The statistical significance was evaluated using the Student's *t*-test (\* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ ). These results are from three independent experiments, and each line represents s.d. **(C)** Snail protein expression was also evaluated under the same experimental conditions.

expression. Exogenous DJ-1 overexpression markedly decreased the EMT negative regulator KLF17, as confirmed by investigating ID-1 expression inhibition by direct binding of KLF17 transcription factor. Nagakubo *et al* (1997), who first demonstrated that DJ-1 is a Ras-dependent oncogene, showed that DJ-1 transcription activity was not detected in an artificial promoter binding system. We tried to uncover whether DJ-1 binds directly to KLF17 promoter sequence. The exact mechanisms by which DJ-1 inhibits KLF17 expression remain unclear and will be investigated further.

To further evaluate that KLF17 is one of the downstream signalling proteins of DJ-1 mediating cellular EMT and invasion, Epistasis analysis of DJ-1 and KLF17 was performed. After double overexpression of these two genes, cellular morphology, invasion efficacy, and PTEN gene expression were analysed. Interestingly, KLF17 overexpression overcomes DJ-1-mediated EMT and invasiveness. In addition, KLF17 and DJ-1 co-overexpression recovered decreased PTEN mRNA expression by DJ-1 single overexpression. These data suggest that KLF17 might be one of the mediators of DJ-1 that accelerates EMT and cell invasion through PTEN inhibition.

Although several studies have proposed that DJ-1 is a potent modulator of the PI3K/Akt pathway in both breast cancer cells and neurons (Kim *et al*, 2005; Yang *et al*, 2005), there is little molecular basis for this assumption. Based on the present evidence that DJ-1 inhibits KLF17 expression, which is correlated negatively with

PTEN immunoreactivity in primary breast cancer tissue (Kim *et al*, 2005), DJ-1 seems to modulate the PI3K/Akt pathway through KLF17 downregulation. A previous study finding that PTEN is required for the regulation of the PI3K/Akt pathway by DJ-1 (He *et al*, 2012) supports this suggestion. In addition, Nagakubo *et al* (1997) suggested that DJ-1 does not bind to Ras directly, suggesting that the signalling molecule(s) downstream of DJ-1 interact with Ras. Our data showed that Ras inhibitors alleviated acceleration of cell invasion by DJ-1, supporting that DJ-1 works with Ras to increase cell invasion. A previous study showed that KLF17 downregulation enhances the key role played by Ras in tumour cell invasion (Gumireddy *et al*, 2009). Furthermore, coexpression of ID-1 and Ras produces an abundance of rapidly growing aggressive tumours and metastasis (Swarbrick *et al*, 2008). Straume and Akslen (2005) also showed a significant increase in ID-1 expression in cases with BRAF mutations. These data partially explain how DJ-1 regulate cell invasion in a Ras-dependent manner.

Altogether, these data suggest for the first time that DJ-1 promotes breast cancer cell invasion and EMT in part through the downregulation of KLF17, resulting in ID-1 upregulation, thus accounting for DJ-1 inhibition of PTEN expression. Furthermore, considering the synergetic effects of ID-1 and Ras in stimulating cancer metastasis (Swarbrick *et al*, 2008), our data reveal ID-1 as an intermediary between DJ-1 and a Ras-related signal transduction pathway.

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## CONFLICT OF INTEREST

The authors declare no conflict of interest.

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