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#### **Minireview**

# MicroRNA dysregulation in colorectal cancer: a clinical perspective

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Recent researches have shed light on the biological importance of microRNAs (miRNAs) in colorectal cancer (CRC) genesis, progression and response to treatments. The potential utility of miRNAs in the preclinical stage have been explored and investigated. In this review, we explored the literature and reviewed the cutting edge progress in the discovery of noninvasive plasma and faecal miRNAs for CRC early diagnosis, as well as their measurability and predictability. We also discussed the utility of miRNAs as novel prognostic and predictive markers, and their association with CRC clinical phenotypes including recurrence, metastasis and therapeutic outcomes. Finally, we summarised miRNA-related single-nucleotide polymorphisms and their potential influence on sporadic CRC susceptibility and therapeutic response. In conclusion, the use of miRNAs as biomarker for CRC is still in its infancy and need further characterisation and evaluation.

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MicroRNAs (miRNAs), a class of highly conserved ~22-nucleotides single-stranded RNAs, can act as trans-acting factors that suppress translation or induce messenger RNA (mRNA) degradation of target genes (Bartel, 2004). A spectrum of dysregulated miRNAs was identified to be associated with colorectal cancer (CRC) genesis, progression and therapeutic response. Herein, we summarise recent findings and discuss the potential value of miRNAs as biomarkers for CRC diagnosis, prognosis and susceptibility.

#### mirnas as diagnostic markers

Approximately 1.23 million individuals worldwide will develop CRC and 0.6 million people will die of it annually (Ferlay et al, 2010). Fortunately, the mortality and morbidity have modestly declined and CRC has become a preventable disease if preneoplastic lesions can be detected with modern screening methods (Lieberman, 2009). Among the available screening approaches, colonoscopy is recommended as the gold standard for high-risk people; however, its invasive nature and expensive cost have hampered the worldwide application. This setting provides the foundation for defining new biomarkers in blood and stool, which can offer more acceptable and practical benefits.

#### PLASMA mirnas as diagnostic markers

Surprisingly high concentration of miRNAs is discovered in highly stable, cell-free form in the peripheral blood (Mitchell *et al*, 2008).

\*Correspondence: Dr J Yu or Professor SSM Ng; E-mails: junyu@cuhk.edu.hk (J Yu) or simonng@surgery.cuhk.edu.hk (SSM Ng) Received 6 October 2010; revised 13 December 2010; accepted 14 January 2011; published online 1 March 2011 Circulating miRNAs are packed in complexes, either called exosomes or microvesicles, and released by normal and tumour cells through unknown mechanisms. Emerging evidence has indicated that such external miRNAs are also involved in cell-to-cell signal transduction and genetic information exchange (Kosaka et al, 2010). Given that aberrantly expressed miRNAs vary among different tumour types and some of them are secreted into blood (Mitchell et al, 2008), circulating miRNAs can potentially serve as noninvasive markers for CRC detection.

We have previously discovered that miR-17-3p and miR-92a (previously named miR-92), both belonging to the miR-17-92 cluster, were significantly elevated in plasma and CRC tissues, but reduced in the postoperative samples when compared with preoperative samples. These data support the idea that CRC cells can release aberrantly expressed miRNAs into the circulatory system. Furthermore, promising discriminatory power of the two miRNAs was verified in an independent larger cohort: at a cut-off value of 3.6 for miR-17-3p (relative expression in comparison with RNU6B), the sensitivity was 64% and the specificity was 70%; at a cut-off value of 240 for miR-92a, the sensitivity was 89% and the specificity was 70% (Table 1). In addition, miR-92a could distinguish CRC from other gastrointestinal cancers and inflammatory bowel diseases (Ng et al, 2009). Diagnostic potential of miR-92a was further verified by Huang et al. They confirmed that miR-92a was significantly increased in CRC plasma compared with healthy control, and the sensitivity and specificity were consistent with our previous study (Table 1). It was also reported that miR-92a could distinguish advanced adenoma from normal control, with a sensitivity of 64.9% and a specificity of 81.4%, whereas its expression levels were not correlated with tumour-node-metastasis (TNM) stages (Ng et al, 2009; Huang et al, 2010).

Tumour-derived plasma miR-29a is another potential CRC diagnostic marker. It was demonstrated that plasma miR-29a could distinguish CRC and advanced adenoma from normal group with



**Table I** miRNAs as potential diagnostic markers for CRC

miRNA	Sensitivity,	Specificity,	AUC	Sample size, n	Endogenous control	Remarks	Reference
Plasma miRNA	1						
miR-17-3p	64	70	CRC: 0.717 (95% CI: 0.63-0.80)	140 (90 CRC, 50 control)	RNU6B	Upregulated in CRC plasma	Ng et al (2009)
miR-92a	89	70	CRC: 0.885 (95% CI: 0.83-0.94)	140 (90 CRC, 50 control)	RNU6B	Upregulated in CRC plasma, could distinguish CRC from other GI cancer and IBD, not associated with TNM stages	Ng et al (2009)
miR-92a	84	71.2	CRC: 0.838 (95% Cl: 0.775-0.900)	196 (100 CRC, 37 advanced adenomas, 59 control)	miR-16	Upregulated in CRC plasma, not associated with TNM stages	Huang et al (2010)
	64.9	81.4	Advanced adenomas: 0.749 (95% CI: 0.642-0.856)				
miR-29a	69	89.1	CRC: 0.844 (95% Cl: 0.786-0.903)	196 (100 CRC, 37 advanced adenomas, 59 control)	miR-16	Upregulated in CRC plasma, associated with advanced TNM stages	Huang et al (2010)
	62.2	84.7	Advanced adenomas: 0.769 (95% CI: 0.669 – 0.869)	,			
Faecal miRNA							
miR-92a	50	80	_	133 (59 CRC, 74 control)	Equal amount of total RNA	Upregulated in stool of CRC patients	Wu et al (2010)
miR-21	50	83	_	133 (59 CRC, 74 control)	Equal amount of total RNA		Wu et al (2010)
miR-17-92 cluster	69.5	81.5	_	316 (197 CRC, 119 control)	U6 snRNA	Upregulated in stool of CRC patients	Koga et al (2010)
miR-135	46.2	95	_	316 (197 CRC, 119 control)	U6 snRNA	Upregulated in stool of CRC patients	Koga et al (2010)

Abbreviations: AUC = area under the ROC curve; CI = confidence interval; CRC = colorectal cancer; GI cancer = gastrointestinal cancer; IBD = inflammatory bowel disease; miRNA = microRNA; snRNA = small nuclear RNA; TNM = tumour-node-metastasis.

sensitivities of 69 and 62.2%, and specificities of 89.1 and 84.7%, respectively (Table 1). Further studies indicated that the level of miR-29a was significantly increased in CRC compared with adenoma, and that its expression was associated with more advanced TNM stages (Huang et al, 2010).

Employing high-throughput transcriptomic sequencing technique, Chen et al (2008) found that the serum miRNA expression profiles of CRC and healthy controls were significantly different: 69 miRNAs were only detected in CRC but not in the control group. Nevertheless, it is most likely that only a minor portion of these dysregulated miRNAs can be finally developed into CRC-specific markers, since roughly 80% of the aberrantly expressed miRNAs are also present in the serum of lung cancer patients (Chen et al, 2008). It is possible that common carcinogenesis-related miRNAs are shared by different types of tumours or it is merely a tumourinduced 'systemic reaction' generated from other sources, such as immune cells. If the latter is the case, future studies should cautiously verify the applicability of a single plasma miRNA, or the use of a combination of multiple plasma miRNAs for specific CRC diagnosis should be recommended.

In previous studies, SYBR green-based real-time polymerase chain reaction (PCR) was generally adopted to detect the presence of miRNA. However, this method has a lower specificity and causes high false-positive and false-negative rates in diagnosis. Recently, Kroh et al (2010) described an optimised method of plasma/serum miRNA detection and data analysis. It is expected that the improved protocol can increase the detection accuracy and help to uncover novel miRNA markers in the plasma.

#### FAECAL mirnas as Diagnostic Markers

Stool-based test is widely adopted as noninvasive screening methods for CRC diagnosis. Currently, the most commonly used

stool test for CRC screening is the guaiac-based faecal occult blood test (FOBT). However, the test itself has some crucial limitations. Owing to the irregular bleeding nature of colorectal tumours, presence of red meat in digested residues, and other causes of bleeding in the gastrointestinal tract, the specificity and sensitivity of FOBT for CRC are relatively poor. Some advanced adenomas with diameter larger than 1 cm are missed by FOBT (Lieberman, 2009). Other stool-based detections depend on colonocytes that are constantly exfoliated from the intestinal tract as well as from the neoplastic site, which theoretically carry important genetic and epigenetic information for subsequent testing, such as detection of mutation genes or dysregulated mRNAs, proteins or miRNAs.

In contrast with the fast degradation of mRNA and protein, endogenous miRNAs are packed and protected from RNase. They are therefore more likely to be detected. Nonetheless, the stool environment is much more complex and hostile than plasma, and human RNA is expected to constitute < 1% of totally stool RNA. In this regard, several criteria, including measurability, reproducibility and predictability, must be met if stool miRNAs have to be developed as diagnostic markers. It is a ground-breaking attempt that Ahmed et al (2009a) evaluated the feasibility of using miRNA from faecal specimens as screening markers for CRC. They developed a novel detection protocol including stool preparation, stool miRNA extraction and quantitative analysis. Commercial miRNA extraction kit is also available for stool miRNA purification, which can yield total RNA of high quality and integrity for further assays. Taqman-based quantitative real-time PCR is performed for miRNA quantification, which can eliminate the interference from food and intestinal bacteria. At present, this analysis protocol is most widely adopted and further optimised by other studies. For example, Koga et al (2010) used immunomagnetic beads conjugated with EpCAM monoclonal antibody to isolate colonocytes from stool, rather than performing colonocytes scrapping from the stool mucinous layer; several other studies



used homogenised stool specimens to reduce selection bias. A simpler method known as direct miRNA analysis, which analysed extracellular miRNA in stool without RNA extraction has also been recently developed (Link et al, 2010). It is necessary to compare different methods in future studies and an appropriate balance between precision and costs should be considered for further clinical use.

A key issue of miRNA detection in stool is the selection of endogenous control. Although several small RNA species are recommended for normalising miRNA expression in tissues and cell lines (Wong et al, 2007), the widely used RNU6B expression is not correlated with total RNA concentration in stool (Link et al., 2010). In addition, RNU6B is rapidly degraded and the detection rate was relatively low in stool specimens. As there is no consensus on suitable control for stool testing, it is suggested that quantification should be performed with equal amount of starting total RNA or adopting other stably expressed small RNAs as normaliser, such as the classical 18S rRNA control and miR-16.

It has been reported that the expression levels of several miRNAs can distinguish healthy control and ulcerative colitis from CRC, as well as to differentiate among CRC with different Dukes' stages. miR-21, miR-106a, miR-96, miR-203, miR-20a, miR-326 and miR-92 showed a higher expression level in stool of CRC patients with advanced Dukes' stages, while miR-320, miR-126, miR-484-5p, miR-143, miR-145, miR-16 and miR-125b exhibited lower expression in stool of CRC patients with advanced Dukes' stages. It is noteworthy that Ahmed et al (2009a) cannot provide a sufficient statistical power for each miRNA because of small sample size used in the study. Moreover, their conclusion was challenged by other studies. Link et al (2010) reported that the levels of stool miR-21 and miR-106a were significantly increased in patients with CRC and adenoma compared with normal controls, but were negatively associated with tumour stages. On the other hand, the expression levels of miR-143 as well as miR-17, miR-622 and miR-654-3p were found to have no significant difference between colorectal neoplasia and normal control.

On the basis of screening of 59 patients and 74 normal controls, we previously reported that stool miR-92a and miR-21 levels were significantly higher in CRC compared with normal controls. At a cut-off value of 430 copies per ng of extracted stool RNA, miR-92a had a sensitivity of 50% and a specificity of 80% towards CRC. At a cut-off value of 820 000 copies per ng, miR-21 had a sensitivity of 50% and a specificity of 83% (Table 1). However, stool miR-92a cannot distinguish adenoma and CRC, or different CRC stages (Wu et al, 2010). In the largest cohort to date, Koga et al (2010) reported the stool miR-17-92 cluster and miR-135 significantly increased in CRC patients, with sensitivities of 69.5 and 46.2%, and specificities of 81.5 and 95.0%, respectively (Table 1). In addition, the sensitivities of the stool miR-17-92 cluster and miR-135 were lower for the proximal region tumour detection. Although the expression level of miR-21 increased in cancer tissue, stool miR-21 showed no difference between CRC patients and healthy volunteers.

It has been postulated that tumour cells and most tumour markers can be readily detected in stool at earlier stages of CRC than in blood. Thus, stool miRNA testing has the advantage for pre-cancerous lesion screening (Ahlquist, 2010). On the other hand, colonocytes shed from proximal region will transit a longer distance and will be more exposed to cytolytic agents, thus less likely to be preserved and sampled. If this is a common phenomenon for stool miRNA markers, right-sided CRC will have less chance to be detected by this approach. Although the use of stool miRNAs as biomarkers is still in its infancy and none of the miRNA discussed above has progressed beyond the preclinical stage, the available data indicated that higher specificity, sensitivity and reproducibility can be achieved for stool miRNA detection. Further studies of stool miRNA characterisation and validation are needed for CRC diagnosis.

#### mirnas as prognostic markers for CRC

In spite of the improvement in surgical technique, CRC patients who have an operable tumour will still have a high risk of subsequent locoregional relapse. On the other hand, the clinical responses to adjuvant therapy vary among individuals. Recently, the utility of miRNA as prognostic markers for CRC has been extensively studied.

Several preclinical and clinical studies have highlighted the potential of miR-21 as a highly promising prognostic marker. Slaby et al (2007) reported that miR-21 upregulation in CRC patients were associated with lymph node positivity and development of distant metastasis based on their study of 29 patients. Later, a larger scale study using miRNA array was conducted with two independent cohorts with different races and geographical distributions. Schetter et al (2008) identified 37 aberrantly expressed miRNA in CRC, among which 5 highly expressed miRNA (miR-20a, miR-21, miR-106a, miR-181b and miR-203) were associated with poor survival in the test cohort. Further validation confirmed the statistically significant association between poor prognosis and high tumour level of miR-21 in Asian CRC patients. In addition, high miR-21 expression level was found to be associated with poor response to adjuvant therapy and more rapid recurrence in patients with stage III disease. Kulda et al (2010) confirmed that higher miR-21 level in primary tumour was correlated with shorter disease-free interval (DFI), but not with overall survival (OS). Although significantly higher level of miR-21 was also found in liver metastasis of CRC, its level was not associated with DFI or OS. miR-31 is another crucial factor for tumour metastasis and it is dysregulated in both CRC cell lines and tissues. Bandrés et al (2006) found that the expression level of miR-31 was significantly increased in stage IV compared with stage II CRC. Wang et al (2009) confirmed the association between miR-31 level and pathological stages, including local invasion. Numerous other miRNAs (Table 2) are also shown to have potential predictive value for prognosis, but further large-scale studies are needed for their validation.

It is known that microsatellite status is associated with CRC disease course and response to adjuvant therapy. Lanza et al (2007) first suggested a link between microsatellite status and different miRNA profiles, and identified 14 differentially expressed miRNA between microsatellite stable tumour (MSS) and microsatellite unstable tumour-high (MSI-H). Employing miRNA array on 49 specimens from TNM stage II CRC patients, Schepeler et al (2008) showed a signature consisting of 4 miRNAs (miR-142-3p, miR-212, miR-151 and miR144) could significantly separate MSI from MSS tumours, with 92% sensitivity and 81% specificity. Furthermore, they reported that 17 miRNAs could separate the MSS subtype according to the metastatic recurrence status, resulting in 81% accuracy, 77% sensitivity and 83% specificity. Among the 17 miRNAs, high levels of miR-320 and miR-498 were associated with longer progression-free survival. Moreover, subgroups of MSI (MSI-high (MSI-H) and MSI-low (MSI-L)) could be distinguished with miRNAs. Earle et al (2010) later reported that upregulation of miR-92, let-7a and miR-145 were associated with MSI-L status whereas increased level of miR-155, miR-223, miR-31 and miR-26b were correlated with MSI-H status.

#### mirnas as predictive markers for TREATMENT OUTCOME IN CRC

Fluoropyrimidine-based adjuvant therapy is commonly used in CRC treatment. Patients who respond to S-1, the fourth-generation Fluoropyrimidine, showed lower levels of miR-181b and let-7g. However, neither miR-181b nor let-7g was associated with survival (Nakajima et al, 2006). Moreover, by induction G2 phase arrest through downregulation of denticleless protein homolog in a p53dependent manner, miR-215 increased chemoresistance of



Table 2 miRNAs as potential prognostic and predictive markers for CRC

miRNA	Dysregulation <sup>a</sup>	Clinical-related phenotypes	Reference
let-7g	Upregulated in CRC	Higher level associated with poor S-I response	Nakajima et al, 2006
miR-18a	Upregulated in CRC	Higher level associated with poor overall survival	Motoyama et al, 2009
miR-21	Upregulated in adenoma, CRC, and	Higher level associated with lymph node positivity, metastasis;	Slaby et al, 2007; Schetter et al, 2008;
	liver metastasis tissue	poor survival, poor therapeutic outcomes, rapid recurrence; shorter disease-free interval	Kulda et al, 2010
miR-31	Upregulated in CRC	Higher level associated with higher TNM stages and local invasion	Bandrés et al, 2006; Slaby et al, 2007; Wang et al, 2009
miR-106a	Upregulated in colon cancer	Higher level associated with longer disease-free survival and overall survival	Díaz et al, 2008; Schetter et al, 2008
miR-143	Downregulated in colon cancer and liver metastasis tissue	Lower level associated with larger tumour size and longer disease-free interval	Slaby et al, 2007; Motoyama et al, 2009; Wang et al, 2009; Kulda et al, 2010;
miR-145	Downregulated in CRC	Lower level associated with large tumour size; related with tumour location	Slaby et al, 2007; Motoyama et al, 2009 Wang et al, 2009;
miR-181b	Upregulated in CRC	Higher level associated with poor S-1 response	Nakajima et al, 2006
miR-320	Downregulated in MSS tumour	Lower level associated with shorter progression-free survival	Schepeler et al, 2008
miR-498	Downregulated in MSS tumour	Lower level associated with shorter progression-free survival	Schepeler et al, 2008

Abbreviations: CRC = colorectal cancer; miRNA = microRNA; MSS = microsatellite stable tumour; TNM = tumour-node-metastasis. <sup>a</sup>Up- or downregulation is stated as miRNA expression relative to normal colon tissue, or MSS relative to MSI.

HCT116 to methotrexate and tomudex, but had no impact on cisplatin and doxorubicin (Song et al, 2010). Remarkably, although miR-215 was downregulated in clinical colon cancer specimen, high level of miR-215 was found in colon cancer stem cells. Cetuximab, the monoclonal antibody that inhibits epidermal growth factor receptor signal transduction, is used to treat metastatic or advanced CRC with wild-type KRAS. Ragusa et al (2010) found that miR-146b-3p and miR-486-5p were more abundant in mutant KRAS patients compared with wild-type ones. Moreover, they suggested that the downregulation of let-7b and let-7e and upregulation of miR-17-3p were potential predictive markers of cetuximab resistance.

Several miRNAs are also implicated in the modulation of radiosensitivity. Svoboda *et al* (2008) reported that median levels of miR-125b and miR-137 were upregulated in rectal cancer patients after a short-course of capecitabine-based chemoradiotherapy, and higher induction of miR-125b and miR-137 were associated with worse response to the treatment. Moreover, Ahmed *et al* (2009b) found different miRNA expression patterns of colon cancer cell line HT29 treated with two clinical X-irradiation modalities. Nonetheless, their clinical values remain unexplored.

#### mirna-related single-nucleotide Polymorphisms influenced risk of sporadic CRC and therapy response

The whole genome is constantly evolving and generates many germ-line nucleotide changes known as single-nucleotide polymorphisms (SNPs). SNPs that fall into the protein coding sequence may have a biological effect by changing the amino acid sequence or yielding a premature stop code. Although SNPs residing in the untranslated region (UTR) do not alter the function of the protein, they may occasionally perturb the protein expression level and may have pathogenic consequences (Yu et al, 2007). As a result of the stringent sequence requirement of miRNA, it is speculated that SNPs residing in miRNA-associated motifs in the 3' UTR, such as the seed region and position 13-16, will potentially interfere with the formation of miRNA::mRNA complex. It raises the possibility that individuals with altered miRNA target site within tumourrelated genes are more prone to develop CRC. The initial study of association between SNPs at miRNA-binding sites and CRC risk was reported by Landi et al (2008). The selected 104 candidates are well-known genes that are related to inflammation, prostaglandins

and thromboxane synthesis, obesity and insulin resistance. By in silicon searching for the SNPs in 3' UTR and assessing the variation of Gibbs energy between the alleles and validation in a case-control study, they reported that both heterozygotes and homozygotes of SNP (rs17281995) within CD86 showed a statistically significant association with increased risk of CRC in Caucasians (especially the risk of rectal cancer), and that the G>C substitution would weaken the affinity of miR-337, miR-582 and miR-200a\* (Landi et al, 2008). Another SNP that less strongly associated with CRC risk is rs1051690 within insulin receptor (INSR), which would theoretically impact miR-612 and miR-618 (Landi et al, 2008) (Table 3). One more thing that should be addressed is the predicted miRNAs may possibly be false-positive errors, because the algorithm is updating and all the five miRNAs predicted in Landi's paper are not found in the latest miRNA online databases. Thus, whether illegitimate miRNA target site can cause higher CRC risk needs to be experimentally verified in the future.

Ultraconserved region (UCR) encodes a class of non-coding RNAs and their expression can be regulated by miRNA *in vivo* (Calin *et al*, 2007). Previous studies indicated that SNPs at UCR loci are related to cancer susceptibility (Yang *et al*, 2008). Wojcik *et al* (2010) randomly selected and sequenced 28 UCRs in Caucasians and identified 4 UCRs harboured sequence variations in 35 CRC patients, but none of them existed in a cohort of 175 cancer-free control. The A > G substitution at position 90 of *uc.*276 in CRC patients would putatively disturb the interaction with miR-214 and miR-887.

Single-nucleotide polymorphisms located within the miRNA biogenesis machinery or the miRNA itself may also have possible influences on clinical outcomes. Exportin 5 (XPO5) is responsible for the miRNA::miRNA duplex cytoplasm translocation. Inactivated XPO5 trapped many precursor miRNA that had potential tumour-suppressor feature in the nucleus, and promoted cancer growth in vitro and in vivo (Melo et al, 2010). Boni et al (2010) discovered a trend of association between SNP (rs11077) in 3'UTR of XPO5 and disease control rate. It is not clear whether this SNP influence protein level of XPO5 as well as precursor miRNA shuttling. Lee et al (2010) reported that 23 SNPs (including rs11077) in the functional regions of miRNA biogenesis machinery did not influence survival of CRC patients. In addition, SNP (rs1834306) in 5'UTR of pri-miR-100 was weakly associated with CRC progression while C>T substitution (rs7372209) in 5'UTR of pri-miR-26a-1 was significantly associated with disease progres-



Table 3 Nucleotide polymorphisms in CRC patients and potentially related miRNAs

db SNP ID	Location	Variation (M/m)	Potentially related miRNA	Remark	Reference
rs17281995	3'UTR of CD86	G/C	miR-337 miR-582 miR-200a*	Associated with increased risk of CRC in Caucasians	Landi et al (2008)
rs1051690	3'UTR of INSR	G/A	miR-612 miR-618	Associated with increased risk of CRC in Caucasians	Landi et al (2008)
_	Position 90 in uc.276	A/G	miR-214 miR-887	Rare polymorphism found in CRC patients	Wojcik et al (2010)
rs11077	3'UTR of XPO5	A/C	_	Weak associated with DCR; not associated with survival	Boni et al (2010); Lee et al (2010)
rs7372209	5'UTR of pri-miR-26a-1	C/T	miR-26a-1	Significantly associated with ORR and TTP; not associated with survival	Boni et al (2010); Lee et al (2010)
rs1834306	5'UTR of pri-miR-100	C/T	miR-100	Weak associated with TTP; not associated with survival	Boni et al (2010); Lee et al (2010)

Abbreviations: CRC = colorectal cancer; DCR = disease control rate; ORR = overall response rate; M = major allele; m = minor allele; miRNA = microRNA; SNP = single-nucleotide polymorphism; TTP = time to progression; UTR = untranslated region.

sion and therapy response (Boni *et al*, 2010). However, 17 SNP located in miRNA precursors, with inclusion of rs1834306 and rs7372209, did not influence the OS rate in a study of 426 surgically treated Korea CRC patients (Lee *et al*, 2010).

Single-nucleotide polymorphisms and their mechanisms related to CRC progression remain to be investigated. Since SNPs in the miRNA-binding motifs are sometimes deleterious, natural selection keeps the SNP density in conserved seed region at lower level than expected, approximately 0.50 SNP per kb (Chen and Rajewsky, 2006). Nevertheless, genome-wide screening for miRNA-related SNPs and understanding of their biological relevance can make further advances in the evaluation of individual CRC susceptibility, expending the list of CRC-associated genes, and ultimately help to develop patient-tailored medicine.

## FUTURE PERSPECTIVE

Genome-wide searching can help identify many oncogenic and tumour-suppressing miRNAs, which are potential therapeutic targets for CRC treatment. Theoretically, the downregulated miRNAs can be replaced by chemically synthesised mimics. Overexpressed miRNAs can also be targeted by anti-miRNA

oligonucleotides. Recently, Idogawa et al (2009) reported that adenovirus mediated cocistronic expression of the p53 protein and artificial miRNA that targeted p21 increased the chemosensitivity of infected CRC cells. However, without fully understanding their biological functions, manipulation of miRNA can potentially induce severe side-effects as miRNA will influence a spectrum of genes. In addition, the technique of target-specific delivery of miRNA is immature. Many issues must be addressed before the preclinical findings can be translated into clinical benefits.

In conclusion, those studies mentioned above highlight the potential value of miRNAs in CRC diagnosis, prognosis and susceptibility. Further large-scale evaluation in multiple independent cohorts is indispensable for determining their realistic expectation.

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#### Conflict of interest

The authors declare no conflict of interest.

#### REFERENCES

Ahlquist DA (2010) Molecular detection of colorectal neoplasia. *Gastro-enterology* 138: 2127-2139

Ahmed FE, Jeffries CD, Vos PW, Flake G, Nuovo GJ, Sinar DR, Naziri W, Marcuard SP (2009a) Diagnostic microRNA markers for screening sporadic human colon cancer and active ulcerative colitis in stool and tissue. *Cancer Genomics Proteomics* 6: 281 – 295

Ahmed FE, Vos PW, Jeffries C, Wiley JE, Weidner DA, Mota H, Bonnerup C, Sibata C, Allison RR (2009b) Differences in mRNA and microRNA microarray expression profiles in human colon adenocarcinoma HT-29 cells treated with either intensity-modulated radiation therapy (IMRT), or conventional radiation therapy (RT). *Cancer Genomics Proteomics* 6: 109–127

Bandrés E, Cubedo E, Agirre X, Malumbres R, Zárate R, Ramirez N, Abajo A, Navarro A, Moreno I, Monzó M, García-Foncillas J (2006) Identification by real-time PCR of 13 mature microRNAs differentially expressed in colorectal cancer and non-tumoral tissues. *Mol Cancer* 5: 29 Bartel DP (2004) MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 116: 281-297

Boni V, Zarate R, Villa JC, Bandrés E, Gomez MA, Maiello E, Garcia-Foncillas J, Aranda E (2010) Role of primary miRNA polymorphic variants in metastatic colon cancer patients treated with 5-fluorouracil and irinotecan. *Pharmacogenomics J*, e-pub ahead of print 29 June 2010; doi:10.1038/tpj.2010.58

Calin GA, Liu CG, Ferracin M, Hyslop T, Spizzo R, Sevignani C, Fabbri M, Cimmino A, Lee EJ, Wojcik SE, Shimizu M, Tili E, Rossi S, Taccioli C, Pichiorri F, Liu X, Zupo S, Herlea V, Gramantieri L, Lanza G, Alder H, Rassenti L, Volinia S, Schmittgen TD, Kipps TJ, Negrini M, Croce CM (2007) Ultraconserved regions encoding ncRNAs are altered in human leukemias and carcinomas. *Cancer Cell* 12: 215–229

Chen K, Rajewsky N (2006) Natural selection on human microRNA binding sites inferred from SNP data. *Nat Genet* **38:** 1452 – 1456

Chen X, Ba Y, Ma L, Cai X, Yin Y, Wang K, Guo J, Zhang Y, Chen J, Guo X, Li Q, Li X, Wang W, Zhang Y, Wang J, Jiang X, Xiang Y, Xu C, Zheng P, Zhang J, Li R, Zhang H, Shang X, Gong T, Ning G, Wang J, Zen K, Zhang J, Zhang CY (2008) Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* 18: 997 – 1006

Díaz R, Silva J, García JM, Lorenzo Y, García V, Peña C, Rodríguez R, Muñoz C, García F, Bonilla F, Domínguez G (2008) Deregulated expression of miR-106a predicts survival in human colon cancer patients. Genes Chromosomes Cancer 47: 794-802



- Earle JS, Luthra R, Romans A, Abraham R, Ensor J, Yao H, Hamilton SR (2010) Association of microRNA expression with microsatellite instability status in colorectal adenocarcinoma. *J Mol Diagn* 12: 433-440
- Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM (2010) Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 127: 2893–2917
- Huang Z, Huang D, Ni S, Peng Z, Sheng W, Du X (2010) Plasma microRNAs are promising novel biomarkers for early detection of colorectal cancer. Int J Cancer 127: 118-126
- Idogawa M, Sasaki Y, Suzuki H, Mita H, Imai K, Shinomura Y, Tokino T (2009) A single recombinant adenovirus expressing p53 and p21-targeting artificial microRNAs efficiently induces apoptosis in human cancer cells. Clin Cancer Res 15: 3725-3732
- Koga Y, Yasunaga M, Takahashi A, Kuroda J, Moriya Y, Akasu T, Fujita S, Yamamoto S, Baba H, Matsumura Y (2010) MicroRNA expression profiling of exfoliated colonocytes isolated from feces for colorectal cancer screening. Cancer Prev Res (Phila) 3: 1435-1442
- Kosaka N, Iguchi H, Ochiya T (2010) Circulating microRNA in body fluid: a new potential biomarker for cancer diagnosis and prognosis. *Cancer Sci* 101: 2087 – 2092
- Kroh EM, Parkin RK, Mitchell PS, Tewari M (2010) Analysis of circulating microRNA biomarkers in plasma and serum using quantitative reverse transcription-PCR (qRT-PCR). *Methods* **50:** 298-301
- Kulda V, Pesta M, Topolcan O, Liska V, Treska V, Sutnar A, Rupert K, Ludvikova M, Babuska V, Holubec Jr L, Cerny R (2010) Relevance of miR-21 and miR-143 expression in tissue samples of colorectal carcinoma and its liver metastases. Cancer Genet Cytogenet 200: 154-160
- Landi D, Gemignani F, Naccarati A, Pardini B, Vodicka P, Vodickova L, Novotny J, Forsti A, Hemminki K, Canzian F, Landi S (2008) Polymorphisms within micro-RNA-binding sites and risk of sporadic colorectal cancer. *Carcinogenesis* 29: 579 – 584
- Lanza G, Ferracin M, Gafa R, Veronese A, Spizzo R, Pichiorri F, Liu CG, Calin GA, Croce CM, Negrini M (2007) mRNA/microRNA gene expression profile in microsatellite unstable colorectal cancer. *Mol Cancer* 6: 54
- Lee HC, Kim JG, Chae YS, Sohn SK, Kang BW, Moon JH, Jeon SW, Lee MH, Lim KH, Park JY, Choi GS, Jun SH (2010) Prognostic impact of microRNA-related gene polymorphisms on survival of patients with colorectal cancer. J Cancer Res Clin Oncol 136: 1073 – 1078
- Lieberman DA (2009) Clinical practice. Screening for colorectal cancer. N Engl J Med 361: 1179–1187
- Link A, Balaguer F, Shen Y, Nagasaka T, Lozano JJ, Boland CR, Goel A (2010) Fecal microRNAs as novel biomarkers for colon cancer screening. Cancer Epidemiol Biomarkers Prev 19: 1766-1774
- Melo SA, Moutinho C, Ropero S, Calin GA, Rossi S, Spizzo R, Fernandez AF, Davalos V, Villanueva A, Montoya G, Yamamoto H, Schwartz Jr S, Esteller M (2010) A genetic defect in exportin-5 traps precursor microRNAs in the nucleus of cancer cells. *Cancer Cell* 18: 303–315
- Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, Peterson A, Noteboom J, O'Briant KC, Allen A, Lin DW, Urban N, Drescher CW, Knudsen BS, Stirewalt DL, Gentleman R, Vessella RL, Nelson PS, Martin DB, Tewari M (2008) Circulating microRNAs as stable blood-based markers for cancer detection. *Proc* Natl Acad Sci USA 105: 10513 – 10518
- Motoyama K, Inoue H, Takatsuno Y, Tanaka F, Mimori K, Uetake H, Sugihara K, Mori M (2009) Over- and under-expressed microRNAs in human colorectal cancer. *Int J Oncol* **34:** 1069 1075

- Nakajima G, Hayashi K, Xi Y, Kudo K, Uchida K, Takasaki K, Yamamoto M, Ju J (2006) Non-coding MicroRNAs hsa-let-7 g and hsa-miR-181b are associated with chemoresponse to S-1 in colon cancer. *Cancer Genomics Proteomics* 3: 317 324
- Ng EK, Chong WW, Jin H, Lam EK, Shin VY, Yu J, Poon TC, Ng SS, Sung JJ (2009) Differential expression of microRNAs in plasma of patients with colorectal cancer: a potential marker for colorectal cancer screening. *Gut* 58: 1375–1381
- Ragusa M, Majorana A, Statello L, Maugeri M, Salito L, Barbagallo D, Guglielmino MR, Duro LR, Angelica R, Caltabiano R, Biondi A, Di Vita M, Privitera G, Scalia M, Cappellani A, Vasquez E, Lanzafame S, Basile F, Di Pietro C, Purrello M (2010) Specific alterations of microRNA transcriptome and global network structure in colorectal carcinoma after cetuximab treatment. Mol Cancer Ther 9: 3396-3409
- Schepeler T, Reinert JT, Ostenfeld MS, Christensen LL, Silahtaroglu AN, Dyrskjøt L, Wiuf C, Sørensen FJ, Kruhøffer M, Laurberg S, Kauppinen S, Ørntoft TF, Andersen CL (2008) Diagnostic and prognostic microRNAs in stage II colon cancer. Cancer Res 68: 6416-6424
- Schetter AJ, Leung SY, Sohn JJ, Zanetti KA, Bowman ED, Yanaihara N, Yuen ST, Chan TL, Kwong DL, Au GK, Liu CG, Calin GA, Croce CM, Harris CC (2008) MicroRNA expression profiles associated with prognosis and therapeutic outcome in colon adenocarcinoma. *JAMA* 299: 425-436
- Slaby O, Svoboda M, Fabian P, Smerdova T, Knoflickova D, Bednarikova M, Nenutil R, Vyzula R (2007) Altered expression of miR-21, miR-31, miR-143 and miR-145 is related to clinicopathologic features of colorectal cancer. Oncology 72: 397 – 402
- Song B, Wang Y, Titmus MA, Botchkina G, Formentini A, Kornmann M, Ju J (2010) Molecular mechanism of chemoresistance by miR-215 in osteosarcoma and colon cancer cells. *Mol Cancer* 9: 96
- Svoboda M, Izakovicova Holla L, Sefr R, Vrtkova I, Kocakova I, Tichy B, Dvorak J (2008) Micro-RNAs miR125b and miR137 are frequently upregulated in response to capecitabine chemoradiotherapy of rectal cancer. *Int J Oncol* 33: 541 547
- Wang CJ, Zhou ZG, Wang L, Yang L, Zhou B, Gu J, Chen HY, Sun XF (2009) Clinicopathological significance of microRNA-31, -143 and -145 expression in colorectal cancer. *Dis Markers* 26: 27-34
- Wong L, Lee K, Russell I, Chen C (2007) Endogenous controls for real-time quantitation of miRNA using TaqMan microRNA assays. *Applied Biosystems Application Note*, Publication 127AP11-01
- Wojcik SE, Rossi S, Shimizu M, Nicoloso MS, Cimmino A, Alder H, Herlea V, Rassenti LZ, Rai KR, Kipps TJ, Keating MJ, Croce CM, Calin GA (2010) Non-codingRNA sequence variations in human chronic lymphocytic leukemia and colorectal cancer. *Carcinogenesis* 31: 208-215
- Wu CW, Ng SS, Leung WW, Lee CW, Wong CY, Yu J, Sung JJ (2010) MicroRNAs in stool as screening marker for colorectal cancer. Gastroenterology 138: S-194
- Yang R, Frank B, Hemminki K, Bartram CR, Wappenschmidt B, Sutter C, Kiechle M, Bugert P, Schmutzler RK, Arnold N, Weber BH, Niederacher D, Meindl A, Burwinkel B (2008) SNPs in ultraconserved elements and familial breast cancer risk. Carcinogenesis 29: 351 355
- Yu Z, Li Z, Jolicoeur N, Zhang L, Fortin Y, Wang E, Wu M, Shen SH (2007) Aberrant allele frequencies of the SNPs located in microRNA target sites are potentially associated with human cancers. *Nucleic Acids Res* 35: 4535-4541