

Original Article

MiR-506 suppresses proliferation of hepatoma cells through targeting YAP mRNA 3'UTR

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Aim: MiR-506 is a miRNA involved in carcinogenesis of several kinds of cancer. In this study, we explored whether miR-506 played a critical role in hepatocellular carcinoma (HCC).

Methods: Twenty HCC and adjacent normal liver tissue samples were collected. Human hepatoma cell lines HepG2 and H7402 were used for *in vitro* studies. The expression of miR-506 and transcriptional co-activator YAP was examined using qRT-PCR. Western blot analysis was used to measure the expression of YAP and its target genes. Luciferase reporter gene assay was used to identify YAP as a target gene of miR-506. MTT and EdU assays were carried out for functional analysis.

Results: The expression of miR-506 was significantly lower in HCC than in adjacent normal liver tissues. Bioinformatics analysis revealed that YAP mRNA might be one of the targets of miR-506, and miR-506 in HCC tissues was found to be negatively correlated with YAP ($r=-0.605$). In both HepG2 and H7402 cells, miR-506 dose-dependently down-regulated YAP and its target genes c-Myc and the connective tissue growth factor (CTGF). Luciferase reporter gene assays demonstrated that miR-506 targeted the wild type 3'UTR of YAP mRNA, but not a 3'UTR with a mutant seed site. Furthermore, miR-506 significantly inhibited the proliferation of HepG2 and H7402 cells, while anti-miR-506 enhanced the cell proliferation, which was blocked by YAP siRNA.

Conclusion: MiR-506 suppresses the proliferation of hepatoma cells by targeting YAP mRNA.

Keywords: microRNA; miR-506; hepatocellular carcinoma; cell proliferation; YAP; c-Myc; connective tissue growth factor

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Introduction

MicroRNAs (miRNAs) are small, non-coding RNAs of approximately 19–25 nucleotides. MiRNAs negatively modulate gene expression through RNA-induced silencing complex-mediated mRNA cleavage or translational repression when partially complementary sequences are present in the 3' un-translated regions (3'UTRs) of target mRNAs^[1, 2]. Most importantly, numerous miRNAs are involved in cancers, and they can take on opposing roles as oncogenes or tumor suppressor genes depending on the target genes^[3–5]. Hepatocellular carcinoma (HCC) is the sixth most common neoplasm and the third leading cause of cancer death worldwide^[6]. The development and progression of HCC is associated with the accumulation of additional genetic alterations^[7]. Many studies have emphasized the causal links between miRNA deregulation and cancer development. Our laboratory has reported that miR-

520e (or miR-205) suppresses the growth of hepatoma cells through the targeting of NF- κ B-inducing kinase (NIK) mRNA (or hepatitis B virus X protein mRNA)^[8, 9]. Recently, it has been reported that miR-506 inhibits epithelial mesenchymal transition (EMT) in ovarian cancer and breast cancer^[10, 11]. In addition, miR-506 can significantly restrain the proliferation of lung cancer cells^[12]. All these reports suggest that miR-506 is a novel microRNA that is important in the development of cancer. However, the functions of miR-506 in liver cancer have not been well documented.

The Hippo pathway is closely associated with hepatocarcinogenesis^[13–15]. The most critical effector of the Hippo pathway is the transcriptional co-activator YAP. By co-activating key transcriptional factors, YAP plays a considerable role in regulating Hippo pathway-responsive gene^[16, 17]. Deregulation of this pathway leads to massive overgrowth of tissue and induces cancer^[18]. More importantly, liver-specific overexpression of YAP results in a greater than 5-fold size enlargement that is reversible after cessation of YAP expression^[16, 19]. Previously, our group found that the expression levels of YAP were

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dramatically elevated in clinical HCC tissues, and that YAP was a key driver gene in HBx-induced hepatocarcinogenesis in a CREB-dependent manner^[20]. However, understanding of the post-transcriptional control of the YAP gene in liver cancer remains elusive.

In this study, we investigated the role of miR-506 in hepatocarcinogenesis. Interestingly, our findings show that miR-506 is able to suppress the proliferation of hepatoma cells through direct targeting of YAP mRNA. Our findings provide new insights into the mechanism by which miR-506 modulates cell proliferation in HCC.

Materials and methods

Patient samples

Twenty paired HCC and adjacent non-tumorous liver tissues were collected from patients undergoing resection of HCC in Tianjin First Center Hospital (Tianjin, China). The relevant characteristics of the studied subjects are shown in Supplementary Table 1. Written consents approving the use of their tissues for research purposes were obtained from the patients. The study protocol was approved by the Institute Research Ethics Committee at the Nankai University.

Cell lines and cell culture

Human hepatoma cell lines, HepG2 and H7402^[20], were maintained in Dulbecco's modified Eagle's medium and RPMI medium 1640, respectively (Gibco, CA, USA). Media were supplemented with heat inactivated 10% fetal bovine serum (FBS, Gibco, CA, USA), 100 U/mL penicillin and 100 mg/mL streptomycin in 5% CO₂ at 37°C.

Plasmid construction

A 294 bp fragment of the YAP 3'UTR was sub-cloned into a pGL3-Control vector (Promega, Madison, WI, USA) immediately downstream of the stop codon of the luciferase gene to generate pGL3-YAP. A mutant construct of the YAP 3'UTR (named pGL3-YAP-MUT), carrying a substitution of 8 nucleotides within the core seed sequence of miR-506, was constructed using overlapping extension PCR. The primers used in this study for plasmid construction were as follows: pGL3-YAP forward, 5'-TGCTCTAGAGGGTATTCTTGAATTGCTTT-3', reverse, 5'-GGGGGCCCGCCGTCTTTCTGC-CATGTTCCA-3'; and pGL3-YAP-MUT forward, 5'-TTTTC-TAAATGTACACGGAAAAAAGGAAAAATGAACACA-3', reverse, 5'-TGTGTTTCATTTTTCCTTTTCCGTGTACATT-TAGAAAA-3'.

Cell transfection

The cells were cultured in a 6-well or 24-well plate for 24 h and were then transfected with plasmids or miRNAs. All transfections were performed using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. YAP siRNA, miR-506 (anti-miR-506), and miRNA control (miRNA Ctrl or anti-miRNA Ctrl) were synthesized by RiboBio (Guangzhou, China). The siRNA duplex sequences

used were as follows: YAP siRNA, 5'-GACAUCUUCUGGU-CAGAGAdTdT-3'^[20].

Quantitative real-time polymerase chain reaction (qRT-PCR)

Total RNA was extracted from the cells or tissues using TRIzol (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. For mature miR-506 detection, total RNA was polyadenylated by poly(A) polymerase (Ambion, Austin, TX, USA) as described previously^[8]. Reverse transcription was performed using poly(A)-tailed total RNA and reverse transcription primer with ImPro-II Reverse Transcriptase (Promega, Madison, WI, USA) according to the manufacturer's instructions. The qRT-PCR was performed as described in the method of Fast Start Universal SYBR Green Master (Rox) (Roche Diagnostics GmbH Mannheim, Germany). The primers used were as follows: YAP forward, 5'-TATCAATCCCAGCACAG-3', reverse, 5'-GGAATGGCTTCAAGGTAG-3'; GAPDH forward, 5'-CATCAC-CATCTTCCAGGAGCG-3', reverse, 5'-TGACCTTGCCCA-CAGCCTTG-3'; miR-506 forward, 5'-TAAGGCACCCTTCT-GAGTAGA-3', reverse, 5'-GCGAGCACAGAAATTAATAC-GAC-3'; and U6 forward, 5'-AGAGCCTGTGGTGTCCG-3', reverse, 5'-CATCTTCAAAGCACTTCCCT-3'.

Luciferase reporter gene assays

Luciferase reporter gene assays were performed using the Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA) according to the manufacturer's instructions. Cells were transferred into 24-well plates at 3×10⁴ cells per well. After 24 h, the cells were transiently co-transfected with the pRL-TK plasmid (Promega, Madison, WI, USA) containing the Renilla luciferase gene, which was used for internal normalization, and with various constructs containing the seed sequence or mutant seed sequence of YAP 3'UTR, or pGL3-Control.

Western blot analysis

The Western blot analysis protocol has been described previously^[20]. The primary antibodies used were mouse anti-β-actin (Sigma, St Louis, MO, USA), rabbit anti-YAP (Proteintech Group, USA), rabbit anti-CTGF (Proteintech Group, USA), and mouse anti-c-Myc (Santa Cruz Biotechnology, Santa Cruz, CA, USA). Protein bands were quantified using Quantity One software (Bio-Rad).

Analysis of cell proliferation

HepG2 cells were seeded onto 96-well plates (1000 cells/well) for 24 h before transfection, and the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay was used to assess cell proliferation every day from the first day until the third day after transfection. The protocol has been described previously^[21]. The 5-ethynyl-2'-deoxyuridine (EdU) incorporation assay was carried out using the Cell-Light™ EdU imaging detecting kit according to the manufacturer's instructions (RiboBio).

Statistical analysis

Statistical significance was assessed by comparing mean values (6 standard deviation; SD) using Student's *t* test for independent groups and was assumed at $P < 0.05$ and $P < 0.01$ or reported as not significant (NS). The expression levels of miR-506 in tumor tissues and matched adjacent non-tumor tissues were compared using Wilcoxon's signed-rank test. Pearson's correlation coefficient was used to determine the correlation between the expression levels of each gene in tumor tissues. Each experiment was repeated at least three times.

Results

MiR-506 is down-regulated and negatively correlated with YAP in HCC tissues

Deregulation of miRNA expression has been linked to tumor development and progression. It has been reported that miR-506 shows a significant association with distant-relapse-free survival (DRFS) in more than 98% of breast cancer patients^[11]. However, the expression of miR-506 in HCC remains unclear. First, we analyzed the expression of miR-506 in 20 paired clinical HCCs and adjacent non-tumor liver tissues using qRT-PCR with normalization against an endogenous control (U6 RNA). Compared with matched non-tumor tissues, all 20 HCC samples showed significant down-regulation of miR-506 (Figure 1A), suggesting that reduced miR-506 expression is a frequent event in HCC tissues and may be involved in hepatocarcinogenesis. To explore the role of miR-506 in the development of liver cancer, we screened the target genes of miR-506 using Targetscan and microRNA.org (<http://www.targetscan.org/> and <http://www.microrna.org>). YAP was identified as a candidate, as it is an important onco-protein in liver cancer. Next, we examined the correlation between the expression of miR-506 and YAP in clinical HCC tissues. MiR-506 levels were markedly inversely correlated with YAP expression (Figure 1B), suggesting miR-506 might target YAP mRNA in HCC tissues. Therefore, we conclude that miR-506 is negatively correlated with YAP and is down-regulated in HCC tissues.

MiR-506 is able to down-regulate YAP in hepatoma cells

Next, we investigated the effect of miR-506 on YAP in hepatoma cells. HepG2 and H7402 cells were transiently transfected with miR-506. Interestingly, we observed that miR-506 was capable of down-regulating YAP in the cells at the protein level in a dose-dependent manner (Figure 2A). The opposite result was obtained when the cells were treated with anti-miR-506 (Figure 2B), suggesting that miR-506 is able to suppress the expression of YAP in hepatoma cells. The transfection efficiency of miR-506 and anti-miR-506 was validated by qRT-PCR (Figure 2A and 2B). Furthermore, our data demonstrated that miR-506 down-regulated the expression levels of c-Myc and connective tissue growth factor (CTGF), two downstream target genes of YAP, in HepG2 and H7402 cells (Figure 2C)^[22, 23]. Conversely, anti-miR-506 heightened the expression levels of c-Myc and CTGF in the cells (Figure 2D). Therefore, we conclude that miR-506 is able to down-regulate YAP in hepatoma cells.

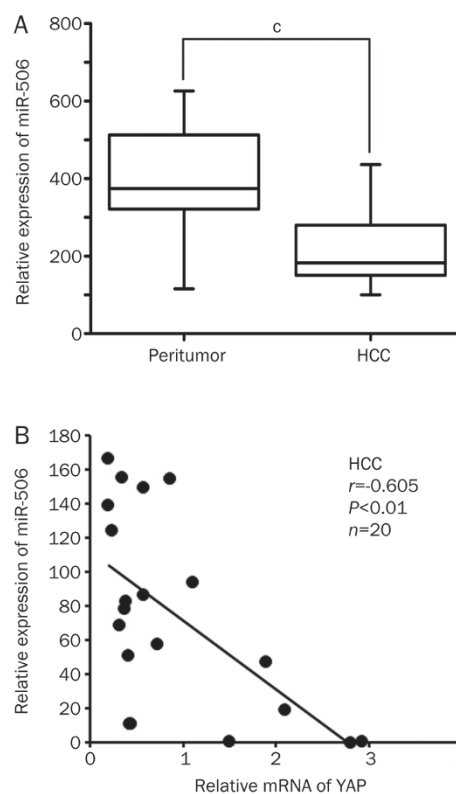


Figure 1. MiR-506 is down-regulated and negatively correlated with YAP in HCC tissues. (A) Relative expression levels of miR-506 were assessed by qRT-PCR in 20 pairs of clinical HCC tissues and corresponding non-tumorous tissues ($P < 0.01$, Wilcoxon's signed-rank test). (B) Correlation of miR-506 levels with YAP mRNA levels was examined by qRT-PCR in 20 cases of clinical HCC tissues ($r = -0.605$, $P < 0.01$, Pearson's correlation).

MiR-506 restrains the expression of YAP through direct targeting of YAP mRNA 3'UTR

To gain insight into the mechanism by which miR-506 inhibits YAP, we identified the miR-506 binding site in the YAP mRNA 3'UTR (Figure 3A) and constructed pGL3-YAP and pGL3-YAP-MUT plasmids (Figure 3B). Our data showed that the co-transfection of miR-506 significantly suppressed the firefly luciferase activity of pGL3-YAP but failed to influence the luciferase activity of pGL3-YAP-MUT in HepG2 and H7402 cells (Figure 3C). Furthermore, inhibition of endogenous miR-506 in the cells with anti-miR-506 resulted in increased firefly luciferase activity of the wild-type reporter but the mutant reporter (Figure 3D). Thus, our data indicate that miR-506 can attenuate the expression of YAP through direct targeting of its mRNA 3'UTR in hepatoma cells.

MiR-506 suppresses the proliferation of hepatoma cells by inhibiting YAP

To examine the potential role of miR-506 in tumorigenesis, we sought to determine whether miR-506 affects the proliferation of hepatoma cells using MTT and EdU assays. Our data indicated that the proliferation of HepG2 and H7402 cells was decreased when the cells were treated with miR-506, while the

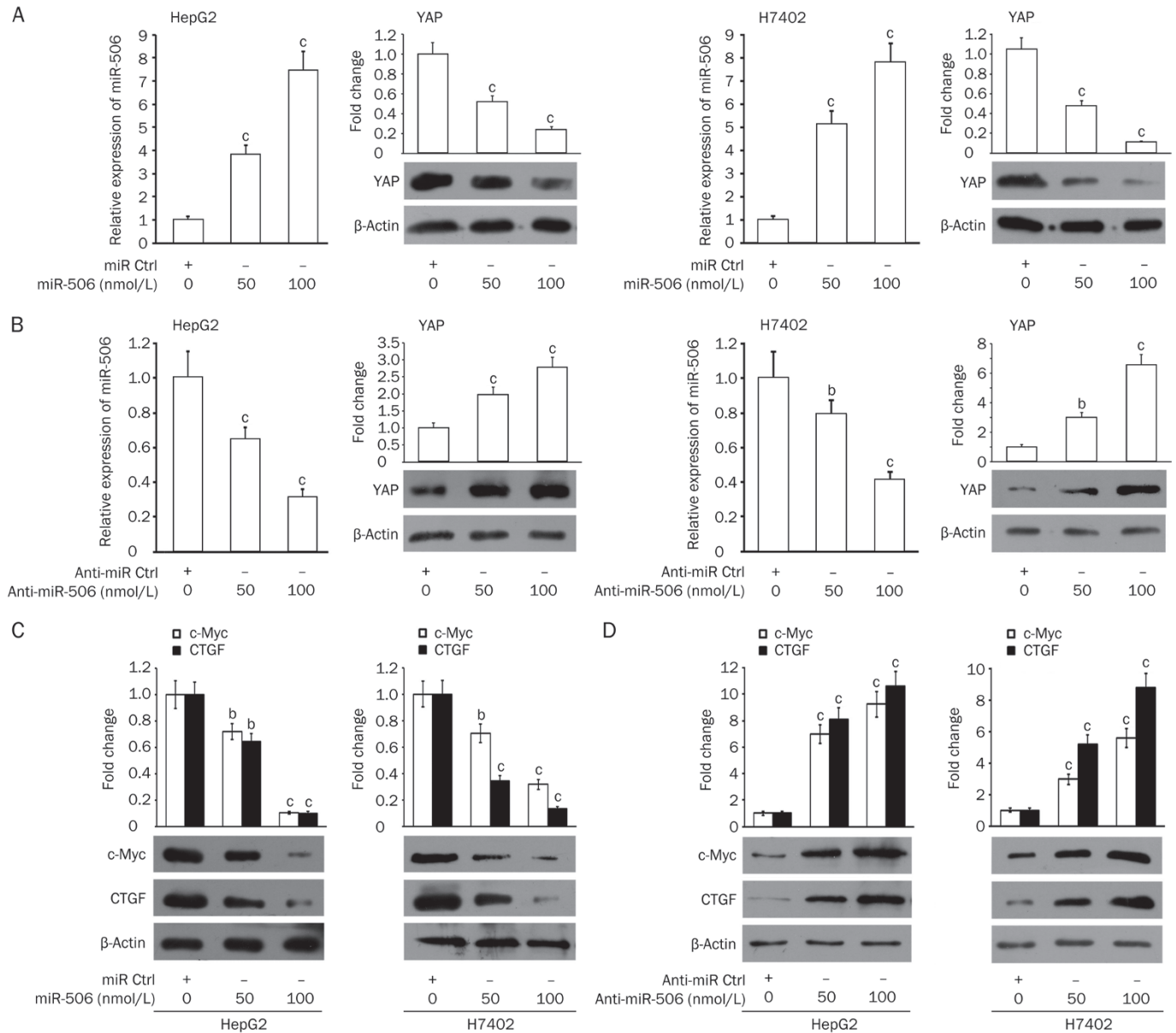


Figure 2. MiR-506 is able to down-regulate YAP in hepatoma cells. (A) The effect of miR-506 on YAP was examined by Western blot analysis in HepG2 and H7402 cells. Levels of miR-506 in the cells were validated by qRT-PCR. (B) The effect of anti-miR-506 on YAP was examined by Western blot analysis in HepG2 and H7402 cells. Levels of miR-506 in the cells were validated by qRT-PCR. (C) The effect of miR-506 on c-Myc and CTGF was examined by Western blot analysis in HepG2 and H7402 cells. (D) The effect of anti-miR-506 on c-Myc and CTGF was examined by Western blot analysis in HepG2 and H7402 cells. Statistically significant differences are indicated: ^b $P < 0.05$; ^c $P < 0.01$; Student's *t* test.

treatment with anti-miR-506 increased the proliferation. The treatment with YAP siRNA was able to block the anti-miR-506-enhanced proliferation (Figure 4), suggesting that miR-506 suppresses the proliferation of hepatoma cells by modulating YAP. Therefore, our observations suggest that miR-506 is able to inhibit the proliferation of hepatoma cells through direct targeting of YAP mRNA.

Discussion

As a heterogeneous tumor, HCC develops through the activation of multiple pathways and molecular alterations^[24, 25]. The

development and progression of HCC is a complicated process that involves the deregulation of multiple genes that are essential to cellular biological processes. In this study, we are interested in the role of miR-506 in hepatocarcinogenesis.

We first investigated the expression levels of miR-506 in 20-paired clinical HCCs and adjacent non-tumor liver tissues. Interestingly, we observed that the expression levels of miR-506 were remarkably decreased in all 20 HCC samples relative to paired non-tumor tissues. Based on this finding, we hypothesized that miR-506 might be a novel tumor-suppressor miRNA in liver cancer. We screened the target genes

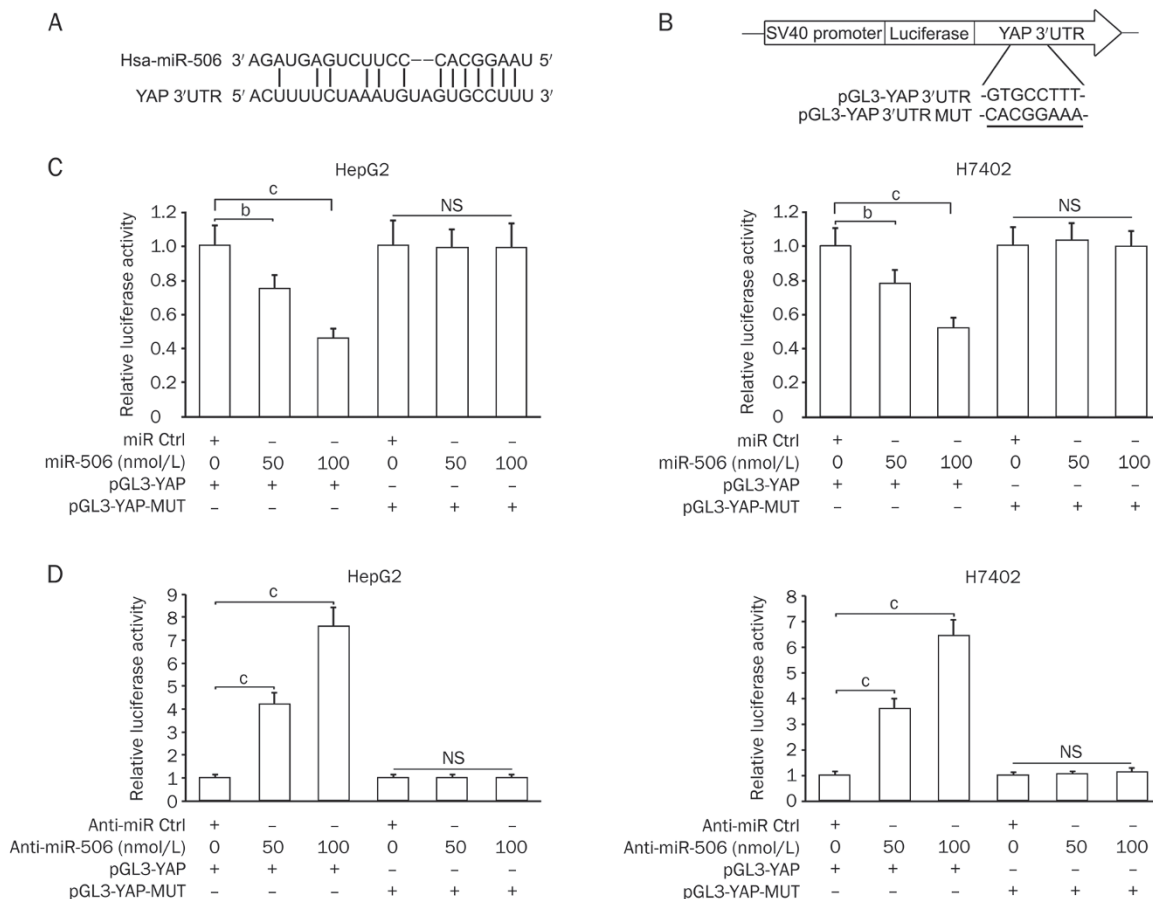


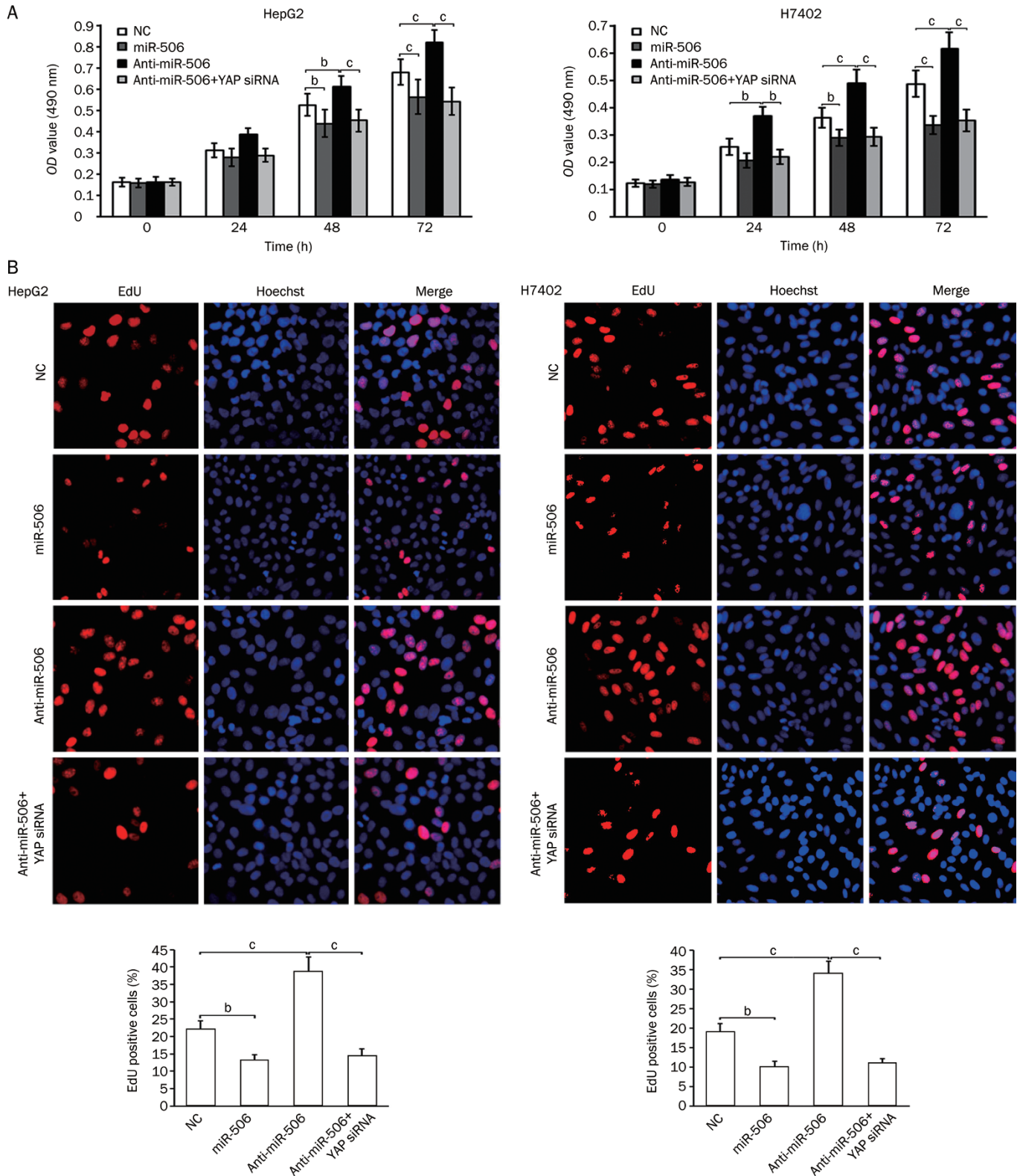
Figure 3. MiR-506 restrains the expression of YAP through direct targeting of YAP mRNA 3'UTR. (A and B) Model shows the binding site of miR-506 in the YAP mRNA 3'UTR by bioinformatics prediction. Schematic diagram shows the generated mutant site in the YAP 3'UTR seed region binding to miR-506 and the insertion sites of wild type YAP 3'UTR (or mutant) downstream of the luciferase reporter gene in pGL3-Control vector. (C) The effect of miR-506 on pGL3-YAP and pGL3-YAP-MUT reporter plasmids in HepG2 and H7402 cells was measured by luciferase reporter gene assays. (D) The effect of anti-miR-506 on pGL3-YAP and pGL3-YAP-MUT reporter plasmids in HepG2 and H7402 cells was measured by luciferase reporter gene assays. Statistically significant differences are indicated: ^b $P < 0.05$, ^c $P < 0.01$; Student's *t* test.

of miR-506 using bioinformatics tools. From this screening, YAP stood out based on its particular functions and expression patterns. However, it had not been reported whether miR-506 could directly target YAP mRNA in HCC. Based on the bioinformatics analysis, we examined the correlation between the expression levels of miR-506 and YAP in clinical HCC tissues. As expected, we observed that miR-506 levels were inversely correlated with YAP expression in the tissues. We then further investigated the effects of miR-506 on YAP expression in hepatoma cells. Our data demonstrated that miR-506 was able to down-regulate YAP and its direct target genes (c-Myc and CTGF) in hepatoma cells. Moreover, we identified that miR-506 could directly bind to the 3'UTR of YAP. We demonstrated that miR-506 could suppress the proliferation of hepatoma cells through direct targeting of YAP mRNA in functional assays.

YAP, an essential downstream effector of the Hippo pathway, is frequently highly expressed in a wide spectrum of human solid tumors^[26, 27]. It has been reported that YAP promotes proliferation in liver by regulating the transcrip-

tion of certain target genes, including Ki-67, CTGF, c-Myc, sex-determining region Y-related high-mobility group box 4 (SOX4), H19, and alpha-fetoprotein (AFP)^[19, 20, 28]. In clinical studies, YAP is an independent predictor of poor disease free survival (DFS) and overall survival (OS) in HCC^[29]. Identifying the underlying mechanisms of YAP regulation will help facilitate the study of YAP as a novel target for the treatment of cancer. It has been reported that mitogen-activated protein kinase kinase 1 (MEK1) promotes YAP expression and that their interaction is critical for tumorigenesis in liver cancer^[30]. In addition, there is a novel positive auto-regulatory feedback loop that underlies the interaction between YAP and c-Myc link the Hippo-YAP and c-Myc pathways^[13]. Therefore, as an oncogene, YAP plays crucial roles in the development of HCC. In this study, we enriched the understanding of the regulation mechanism of YAP at the post-transcriptional level.

Growing evidence suggests that alteration of microRNAs contributes to the development of cancer^[31-33]. Recently, it has been reported that ABCC4, which is involved in the regula-



tion of drug response, is regulated by miR-506^[34]. In addition, miR-506 regulates expression of PPAR alpha in hydroxycampothecin-resistant human colon cancer cells^[35]. MiR-184 post-transcriptionally regulates SOX7 expression and promotes proliferation in human hepatocellular carcinoma cells^[36]. Our group has reported that miR-205 modulates abnormal lipid metabolism of hepatoma cells via targeting acyl-CoA synthetase long-chain family member 1 and 4 (ACSL1 and ACSL4) mRNA^[37, 38]. Based on our findings in this study, we conclude that miR-506 may act as a tumor suppressor miRNA in liver cancer by directly targeting YAP mRNA. Thus, our finding provides new insights into the mechanism by which miR-506 modulates proliferation of hepatoma cells.

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Author contribution

Li-hong YE, Xiao-dong ZHANG, and Yue WANG designed the research, analyzed the data and wrote the paper; Yue WANG, Ming CUI, Bao-di SUN, and Fa-bao LIU performed the research; Li-hong YE and Xiao-dong ZHANG contributed new reagents or analytic tools.

Abbreviations

miRNA, microRNA; YAP, Yes-associated protein; HCC, hepatocellular carcinoma; CTGF, connective tissue growth factor; 3'UTR, 3' untranslated region; MTT, methylthiazolylidiphenyl-tetrazolium bromide; EdU, ethynyldeoxyuridine; qRT-PCR, quantitative real-time polymerase chain reaction; siRNAs, small interfering RNAs.

Supplementary information

Supplementary information is available at *Acta Pharmacologica Sinica's* website.

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