

Original Article

In-silico identification of the binding mode of synthesized adamantyl derivatives inside cholinesterase enzymes

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Aim: To investigate the binding mode of synthesized adamantyl derivatives inside of cholinesterase enzymes using molecular docking simulations.

Methods: A series of hybrid compounds containing adamantane and hydrazide moieties was designed and synthesized. Their inhibitory activities against acetylcholinesterase (AChE) and (butyrylcholinesterase) BChE were assessed *in vitro*. The binding mode of the compounds inside cholinesterase enzymes was investigated using Surflex-Dock package of Sybyl7.3 software.

Results: A total of 26 adamantyl derivatives were synthesized. Among them, adamantane-1-carboxylic acid hydrazide had an almost equal inhibitory activity towards both enzymes, whereas 10 other compounds exhibited moderate inhibitory activity against BChE. The molecular docking studies demonstrated that hydrophobic interactions between the compounds and their surrounding residues in the active site played predominant roles, while hydrophilic interactions were also found. When the compounds were docked inside each enzyme, they exhibited stronger interactions with BChE over AChE, possibly due to the larger active site of BChE. The binding affinities of the compounds for BChE and AChE estimated were in agreement with the experimental data.

Conclusion: The new adamantyl derivatives selectively inhibit BChE with respect to AChE, thus making them good candidates for testing the hypothesis that BChE inhibitors would be more efficient and better tolerated than AChE inhibitors in the treatment of Alzheimer's disease.

Keywords: N-arylidene-1-adamantylcarbohydrazides; adamantane; hydrazide; cholinesterase inhibitor; Alzheimer's disease; acetylcholinesterase; butyrylcholinesterase; molecular docking simulation

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Introduction

Alzheimer's disease (AD), the most common form of dementia, is an increasing medical, social and public health concern^[1]. AD is characterized by deficits in the cholinergic system and the deposition of β -amyloid (A β)^[2]. The cholinergic system has been targeted for the design of anti-Alzheimer's drugs. Cholinesterase inhibitors increase both the level and duration of action of acetylcholine. In addition, it has been demonstrated that both acetylcholinesterase and butyrylcholinesterase (BChE) play an important role in A β -aggregation during

plaque formation^[3]. Many research papers published in the last decade have indicated that the structures of compounds that exhibit cholinesterase inhibitory activity are diverse^[4,5].

Adamantane derivatives are known to exhibit bioactivities that include antiviral^[6], antibacterial^[7,8], and anti-inflammatory^[9] activities and inhibition of the central nervous system^[10] and 11 β -HSD1^[11].

Hydrazones are one of the most studied classes of compounds owing to their ease of synthesis, availability and diversity. Many hydrazones show inhibitory activities towards enzymes^[12]. Various hydrazones have been shown to be strong antimicrobial, anti-inflammatory and antifungal agents^[13,14]. The antimicrobial agents, (3 α ,5 β ,7 α ,12 α)-3,7,12-trihydroxy-N-[(1E)-4-chlorophenylmethylene] cholan-24-hydrazide were introduced recently by our group^[15]. The supe-

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riority of the designed compounds over the drugs Cefaclor and Cefixime, when tested against various bacterial strains, encouraged us to further investigate various hydrazone derivatives for new targets.

Inspired by the diverse bioactivities of molecules containing adamantane and hydrazone moieties, and as a continuation of our work on new cholinergic inhibitors^[16], we designed new hybrid compounds containing both moieties and tested their ability to inhibit acetylcholinesterase and butyrylcholinesterase.

Molecular docking is a powerful computational tool that can predict the interaction energy between two molecules (receptor and ligand) and determine which orientation of a ligand would form the lowest energy complex within the receptor's binding pocket^[17-21]. In the Surflex docking methodology, Hammerhead's empirical scoring function is paired with a molecular similarity method to generate putative poses of ligand fragments. This process implements an incremental construction search approach and a new fragment assembly methodology; each is faster and more accurate. This new fragment assembly method is loosely related to the genetic algorithm approach, although it is a deterministic method.

Our docking results were in good agreement with experimental findings and demonstrated that the bulky adamantyl derivatives are more potent against BChE than AChE because BChE has a larger binding pocket. BChE also has wider substrate specificity and interacts with a broad range of inhibitors. As a result, the binding site of BChE accommodated adamantyl derivatives (**1-26**) relatively well, compared to AChE. Adamantyl derivatives synthesized recently by our group could be considered as lead compounds for developing AD treatments, as some selective BChE inhibitors have already been reported to increase acetylcholine levels and to reduce the formation of abnormal amyloid found in Alzheimer's disease^[22, 23].

Materials and methods

Materials and equipments

¹H NMR spectra were recorded on a Bruker Avance III-500 MHz spectrometer using tetramethyl silane as an internal standard. ¹³C NMR spectra were recorded at 125 MHz using a Bruker Avance III-500 MHz spectrometer and tetramethyl silane as an internal standard. Chemical shifts were reported as δ -values in ppm. The multiplicities of carbon atoms were determined from DEPT experiments. High resolution mass spectra (HRMS) were recorded in positive ion mode by electrospray ionization using a Bruker Daltonics Apex IV, 7.0 T Ultra Shield Plus. Samples were dissolved in chloroform, diluted in spray solution (methanol/water 1:1 *v/v* +0.1 % formic acid) and infused using a syringe pump with a flow rate of 2 μ L/min. External calibration was conducted using an arginine cluster in a mass range *m/z* 175-871. For all HRMS data, mass error: 0.00-0.50 ppm. Melting points (mp) were determined on an Electrothermal Melting point Apparatus and are reported uncorrected in °C. Solvents used in this study were obtained from Scharlau, Fluka or Aldrich<. All reactions were monitored by thin layer chromatography (TLC) using Merck

aluminum plates pre-coated with silica gel (PF254; 20×20×0.25 mm) and were detected by visualization of the plate under UV illumination (λ =254 or 365 nm). Spots were also detected by spraying with anisaldehyde-sulfuric acid in ethanol, followed by heating to 140°C.

Cholinesterase inhibition assay

Inhibition of AChE and BChE was measured *in vitro* using Ellman's spectrophotometric method^[24]. Electric-eel AChE (Torpedo californica type VI-S, Sigma, EC 3.1.1.7), horse-serum BChE (EC 3.1.1.8), acetylthiocholine iodide (ATCh), butyrylthiocholine chloride (BTCh), 5,5-dithiobis[2-nitrobenzoic acid (DTNB) and Tacrine were purchased from Sigma (St Louis, MO, USA). Buffers and other chemicals were of analytical grade. Acetylthiocholine iodide and butyrylthiocholine chloride were used as substrates to assay AChE and BChE, respectively. All inhibition studies were performed in 96-well microtiter-plates using a Bio Tek μ Quant™ microplate spectrophotometer.

Methyl adamantane-1-carboxylate (**2**)

Thionyl chloride (15 mL) was added drop wise with continuous stirring to a solution of adamantane-1-carboxylic acid (5 g, 28 mmol) in dry methanol (60 mL) at 0°C. The mixture was refluxed for 15 h. Upon cooling, a crystalline solid precipitated and was collected via suction filtration and dried, yielding 4.8 g (96%) of methyl adamantane-1-carboxylate (mp 37°C).

¹H NMR (500 MHz, CDCl₃): δ =1.72 (s, 6H, adamantane-H), 1.89 (s, 3H, adamantane-H), 1.92 (s, 6H, adamantane-H), 3.68 (s, CH₃). ¹³C NMR (CDCl₃) δ =27.9 (C-1, C-5, and C-7 of adamantane), 36.5 (C-6, C-8, and C-9 of adamantane), 38.8 (C-2, C-4, and C-10 of adamantane), 40.7 (C-3), 51.52 (CH₃), 178.2 (C-11). HRMS (ESI): calcd for C₁₁H₁₈O₂ [M+H]⁺: 217.11990 found: 217.11960.

Adamantane-1-carboxylic acid hydrazone (**3**)

A mixture of methyl adamantane-1-carboxylate (4.8 g, 25 mmol) and 98% hydrazine hydrate (12 mL) in absolute ethanol (18 mL) was refluxed with stirring overnight. Upon the completion of the reaction (as checked by TLC), the reaction mixture was poured onto crushed ice to produce a white crystalline solid, which was filtered and washed with cold water to yield 4.4 g (93%) of adamantane-1-carboxylic acid hydrazone. (mp=150°C).

N-arylidene-1-adamantylcarbohydrazides (**4a-w**)

The corresponding aromatic aldehyde (1.1 mmol) was added to a solution of 1 mmol of adamantane-1-carboxylic acid hydrazone in absolute ethanol containing 6 drops of glacial acetic acid. The mixture was refluxed for 4 h, poured into cold water, and left standing for 1 h to complete precipitation. The precipitate formed was separated by filtration, washed with petroleum ether and air dried. Products were purified by silica gel column chromatography using a gradient of dichloromethane/ethylacetate solution as an eluent, followed by

recrystallization from ethanol to yield pure compounds.

Enzyme inhibition assay

One hundred and thirty microliters of 0.1 $\mu\text{mol/L}$ sodium phosphate buffer (pH 8.0), 10 μL of test compound solution with different concentrations, and 20 μL of a 4×10^{-7} mg/mL solution of acetylcholinesterase were mixed and incubated for 15 min at 25°C. Twenty microliters of a solution of 8 mg DTNB 3.25 mg and NaHCO_3 dissolved in 1.0 mL phosphate buffer pH=7.0 was added to each well. The reaction was then initiated by addition of 20 μL of 4.0 mg/mL acetylthiocholine iodide in deionized water. The hydrolysis of ATCh was monitored by measuring the formation of the yellow-colored 5-thio-2-nitrobenzoate anion at a wavelength of 412 nm for 5 min. The blank reading solution was used to justify non-enzymatic hydrolysis of substrate during the assay. The blank solution was prepared by replacing the enzyme solution with 20 μL of buffer. An identical solution without inhibitor was processed following the same protocol. The same procedure was used for the butyrylcholinesterase inhibition assay, using a 0.02 mg/mL solution of BChE and 0.5 mg/mL of butyrylthiocholine chloride substrate concentration.

Test compounds were dissolved in ethanol while the control received the same volume of the solvent. All the reactions were performed in triplicate and the initial rate was measured as the rate of change in OD/min (optical density/min) and used in subsequent calculations. IC_{50} values were determined by spectrophotometric measurement of the effect of increasing concentrations of test compounds on enzyme activity. The IC_{50} values were calculated using EZ-Fit Enzyme Kinetics Program.

In-silico methodology

Three dimensional (3D) molecular structures of twenty six adamantyl derivatives (1–26) along with tacrine as a standard were drawn by using the builder within MOE software. All constructed compounds were optimized and charges applied according to the MMFF94 protocol.

Preparation of protein targets

The X-ray crystal structures of both enzymes, human AChE (hAChE, PDB ID: 4EY5:2.30) and human BChE (hBChE, PDB ID: 2 WSL:2.0 Å) were retrieved from Protein Data Bank (PDB), and whole structures were minimized after addition of polar hydrogen atoms.

Molecular docking studies were carried out to understand the binding mode of the synthesized adamantyl derivatives 1–26 inside each of the cholinesterase enzymes using the Surflex-Dock package of Sybyl7.3^[25–27] software. Surflex is a fully automatic, flexible molecular docking algorithm that combines the scoring function from the Hammerhead docking system with a search engine that relies on a surface-based molecular similarity method as a means to rapidly generate suitable putative poses for molecular fragments.

Prior to docking, the 3D structures of inhibitors and recep-

tors were prepared according to Surflex-Dock requirements. In Surflex-Dock, automatic active site detection of prepared proteins was conducted by “protomol” and used an empirically derived scoring function based on the binding affinity. In this study, standard parameters were used to estimate the binding affinities of new adamantyl derivatives characterized by the Surflex-Dock score (total score) expressed in $-\log_{10}(K_d)$ units^[28].

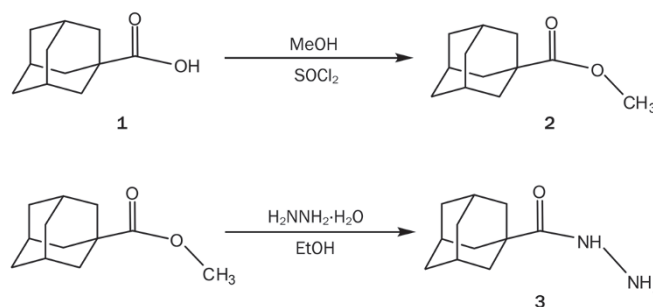
Results and discussion

Chemistry

Preparation of adamantane-1-carboxylic acid hydrazide (3)

The synthesis of adamantane-1-carboxylic acid hydrazide (3) from adamantane-1-carboxylic acid (1) began with esterification of the acid by methanol in the presence of SOCl_2 to give a white solid ester in quantitative yield (2). The ester (2) was recrystallized from ethanol and characterized by comparing the melting point to the published value. Direct esterification of (1) through acid catalysis was carried out by refluxing an ethanolic solution of the acid (1) in the presence of sulfuric acid^[29].

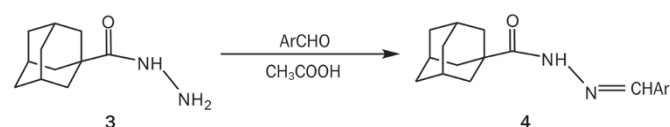
The hydrazide target compound was prepared by refluxing the ester with hydrazine hydrate in ethanol overnight. The reaction afforded the desired product (3) in 93% yield (Scheme 1). The compound was identified by comparing its mp to the published value^[30]. The $^1\text{H-NMR}$ spectra of (3) confirmed substitution of the methoxy group by hydrazine, as it showed signals from the amine hydrogens and the absence of the signal from a methyl group.



Scheme 1. Synthesis of adamantane-1-carboxylic acid hydrazide.

Preparation of N-arylidene-1-adamantylcarbohydrazides (4a–w)

We used condensation of adamantane-1-carboxylic acid hydrazide (3) with different aldehydes as the core synthetic approach for generating this series, as shown in Scheme 2.



Scheme 2. N-arylidene-1-adamantylcarbohydrazide (4a–w) synthesis.

Through this reaction, 23 different aromatic aldehydes were used to obtain the desired series of hydrazides. Of the synthesized compounds, 7 (**4a–g**) have been reported recently^[31] and the remaining 16 (**4h–w**) were synthesized for the first time. All compounds were identified using physico-chemical techniques.

¹H NMR signals from the adamantane moiety appeared as a set of three broad singlets integrating for 6, 6 and 2 protons in the range of δ 1.65–2.02 ppm. The aromatic protons appeared as expected in the range of δ 7.0–8.5 ppm, with multiplicities and coupling constants reflecting the substitution pattern of each compound. The ¹H-NMR spectra of each of the synthesized compounds showed the signal from the methyne proton (CH=N) as a singlet in the range of δ 8.29–8.65 ppm, whereas that of the amide proton resonated as a singlet (sometime broad singlet) at δ 11.2–11.9 ppm. All spectra were in full agreement with the proposed structures. Compounds **4h**, **4i**, and **4j** showed the protons of the hydroxy groups whereas the indole-containing compounds clearly showed the NH signal in the range of 10.5–13.0 ppm. The ¹³C-NMR spectral data of the hydrazones (**4a–w**) were in full agreement with the proposed structures. DEPT experiments were used to indicate the multiplicity of carbon atoms. The adamantane moiety showed four different peaks, two signals for CH₂ carbons, one for CH carbons and one for a quaternary carbon atom. Aromatic carbons appeared in the range of δ 125–165 ppm. The N=CH carbon signal appeared in the region of δ 138–148 ppm, whereas that of the amide carbon (–C=ONH–) resonated in the range of δ 170–174 ppm.

The methyl groups on the aromatic moiety resonated at the expected chemical shift of δ 20–25 ppm, whereas that of the methoxy group resonated in the range of δ 50–60 ppm. The hetero-aromatic compounds showed the anticipated chemical shifts for all of the carbon atoms compared to the known shifts reported in the literature.

High resolution mass spectra were in close agreement with the proposed formula of each compound.

Cholinesterase inhibition studies

The results (Table 1) indicated that adamantane derivatives did not exhibit inhibitory activity against AChE, except compound (**3**), which showed moderate activity with an IC₅₀ of 17.8 μ mol/L. Compounds **1**, **2**, **3**, **4d**, **4h**, **4j**, **4k**, **4l**, **4m**, **4r**, and **4s** were found to have moderate inhibitory activity against BChE, with IC₅₀ values of 29.69, 8.65, 18.05, 10.01, 75.57, 10.75, 8.80, 13.75, 12.40, 35.50, and 29.19 μ mol/L, respectively. The remaining compounds were considered to be inactive because their IC₅₀ values were more than 100 μ mol/L.

Differences in inhibitory activity within this series of compounds towards AChE and BChE can be attributed to differences in structure between these two enzymes. Nicolet *et al*^[32] indicated that, in spite of the similarities between the structures of the two enzymes, several aromatic residues lining the gorge of AChE are replaced by hydrophobic aliphatic residues in BChE. In addition, Phe²⁸⁸ and Phe²⁹⁰ in the acyl-binding pocket of AChE are replaced by Leu²⁸⁶ and Val²⁸⁸, respectively,

in BChE; these changes allow bulkier substrate moieties to bind more readily to BChE than to AChE.

It has been reported that levels of AChE decline in the brain with the progress of AD, which may limit the therapeutic value of AChE inhibitors. On the other hand, the activity of BChE increases in the brains of patients with AD^[33]. Thus, the search for selective BChE inhibitors might be considered as an innovative approach for the treatment of AD^[34]. Here, we report a new class of N-arylidene-1-adamantylcarbohydrazides that may have promising therapeutic potential for AD.

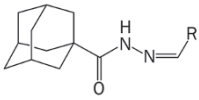
The most active compound was methyl adamantane-1-carboxylate (**2**), although its activity decreased upon conversion to the corresponding hydrazide (**3**). The activity of **4k** was almost identical to that of (**2**). The importance of the N-*p*-hydroxyphenyl moiety for the activity of adamantylcarbohydrazides was confirmed by determining the inhibitory activity of 2,4-dihydroxyphenyl (**4l**) and 2,3,4-trihydroxyphenyl-1-adamantylcarbohydrazides (**4m**). The presence of more than one hydroxyl group did not enhance inhibitory activity.

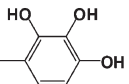
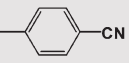

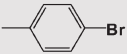

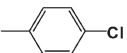

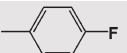
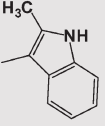
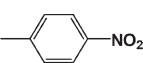
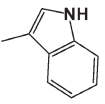

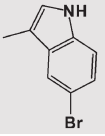
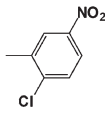
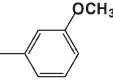



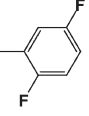
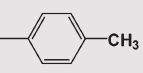
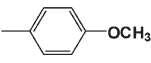
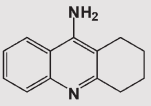
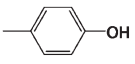
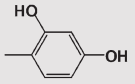
Compound **4j**, which contains a methoxy group in place of a hydroxyl group at the *p*-position of the phenyl ring, showed almost the same IC₅₀ as that of compound **4k**. This suggests that the oxygen atom at the *p*-position plays an important role in the activity of compounds **4k**, **4l**, **4m**, and **4j**. This is also evident when comparing the activity of the above mentioned four compounds with that of inactive **4h** (where phenyl is not substituted), **4u** (in which the oxygen is at the *m*-position), or **4i** (which has a methyl at the *p*-position). In the latter, the decrease in inhibitory activity might also be due to steric factors. Steric effects are indicated upon comparison of the activities of the *p*-bromo (**4a**), *p*-chloro (**4b**) and *p*-fluoro (**4c**) substituents of N-phenyl-1-adamantylcarbohydrazides, where the activity decreases as the size of the halogen increases. The difference in the reactivity between the two constitutional isomers of *p*-nitro (**4d**) and *m*-nitro (**4e**) can be attributed to both steric and electronic factors; the *p*-isomer showed moderate inhibitory activity whereas the *m*-isomer was inactive. Derivatives containing an indole ring (**4r**, **4s**, and **4t**) indicated the same trend, in which compounds **4r** and **4s** are considered as moderately active, whereas the larger **4t** is not active.

Computational studies

Molecular docking studies were performed to provide a theory for the mechanism by which our adamantyl derivatives bind with the cholinesterase enzymes. To date, several ligand-cholinesterase complexes have been reported. Docking studies applied on experimentally resolved structures of BChE (PDB code: 2 WSL) and AChE (PDB code: 4EY5) were carried out using the Surflex-Dock package contained within the Sybyl7.3 program. In the literature, cholinesterase enzymes are divided into five major domains (residue numbers mentioned here relate to BChE): 1) the peripheral active site (PAS) region (Asp68, Tyr330); 2) the catalytic active site (CAS) region (Ser196, His436, Glu325); 3) the acyl pocket region (Leu281, Trp229, Phe438); 4) the choline sub-site region (Trp80, Tyr126, Glu195); and 5) the oxyanion hole region (Gly114, Gly115,

Table 1. Summary of the *in vitro* AChE and BChE activities of compounds **1-3**, **4a-w**, and tacrine.



Compounds	Structure R-	IC ₅₀ (μmol/L) AChE	IC ₅₀ (μmol/L) BChE	Compounds	Structure R-	IC ₅₀ (μmol/L) AChE	IC ₅₀ (μmol/L) BChE
1	Scheme 1	274.41	29.69	4m		353.11	12.4
2	Scheme 1	410.9	8.65	4n		424.88	227.94
3	Scheme 1	17.8	18.05	4o		515.00	327.39
4a		561.85	201.25	4p		633.58	297.05
4b		441.31	180.93	4q		596.23	183.4
4c		570.59	147.26	4r		549.25	35.5
4d		637.28	10.01	4s		638.65	29.19
4e		765.59	236.69	4t		411.15	189.71
4f		439.18	217.96	4u		383.9	207.01
4g		218.55	165.7	4v		399.94	209.14
4h		519.72	75.57	4w		302.7	281.4
4i		624.19	135.76				
4j		607.5	10.75				
Tacrine		6.77	0.615				
4k		619.65	8.80				
4l		464.6	13.75				

Ala197). The CAS residues of both proteins make a specific contribution to the inhibitors' specificity and selectivity.

Docking results demonstrated that FDA approved drugs such as tacrine (THA) bind more efficiently inside the binding cavity of BChE than AChE, consistent with experimental results (Table 2). In the complex THA-BChE (Figure 1A), the THA moiety was bound to the CAS and cationic- π sites. Its

structure is involved in π - π bonding with the phenyl ring of Trp80 (2.94 Å) by a parallel orientation and stabilized by a hydrogen bond with a CAS residue (His436) at a distance of 2.1 Å. However, in the THA-AChE complex, two π - π interactions with THA rings at distance of 3.29 Å and 3.79 Å, respectively, were observed (Figure 1B).

On the basis of BChE experimental inhibition activities,

Table 2. Binding affinities of tacrine with new adamantyl derivatives (**1–26**) against both of the cholinesterase enzymes (hAChE and hBChE).

S No	Compound name	Surflex-Dock score (hBChE)	Surflex-Dock score (hAChE)
1	1	1.6	2.83
2	2	1.22	3.63
3	3	2.02	2.26
4	4a	0.9	2.57
5	4b	0.79	3.06
6	4c	1.19	3.21
7	4d	1.22	1.51
8	4e	0.87	2.84
9	4f	0.43	3.56
10	4g	0.85	2.76
11	4h	1.83	2.8
12	4i	1.23	1.14
13	4j	0.92	0.99
14	4k	3.46	3.53
15	4l	2.96	2.68
16	4m	2.35	2.33
17	4n	1.73	1.71
18	4o	1.51	3.3
19	4p	1.49	2.5
20	4q	0.92	3.45
21	4r	1.61	-0.2
22	4s	2.88	2.91
23	4t	2.67	3.23
24	4u	0.88	3.75
25	4v	1.02	2.94
26	4w	1.46	2.65
27	Tacrine	3.8	3.99

twenty-six new compounds of adamantyl derivatives were categorized into three different groups for understanding the results of the docking study: active compounds (0–79 $\mu\text{mol/L}$), moderately active compounds (135–250 $\mu\text{mol/L}$) and inactive compounds (>250 $\mu\text{mol/L}$). According to this classification, three different compounds were selected to explain the docking results against BChE and its results were correlated with experimental findings. Similar compounds were also used to understand the mechanism of interaction and inhibitory activities against AChE of adamantyl complexes.

Active compounds (0–79 $\mu\text{mol/L}$)

Compound **1** (29.69 $\mu\text{mol/L}$) occupied the entire enzymatic CAS, mid-gorge and PAS regions, displaying several hydrophobic interactions. The hydroxyl (OH) moiety also established a hydrogen bond (1.96 Å) with the carbonyl group of a CAS residue (His436, Figure 2). These interactions show that hydrophobic scaffolds exhibit an experimentally stronger inhibition compared to scaffolds that are hydrophilic in nature. These observations are also supported by comparison of compounds **4k**, **4l**, and **4m**, as a reduced inhibitory potency was observed as the number of hydroxyl groups on the phenyl ring were increased. These results are in agreement with experi-

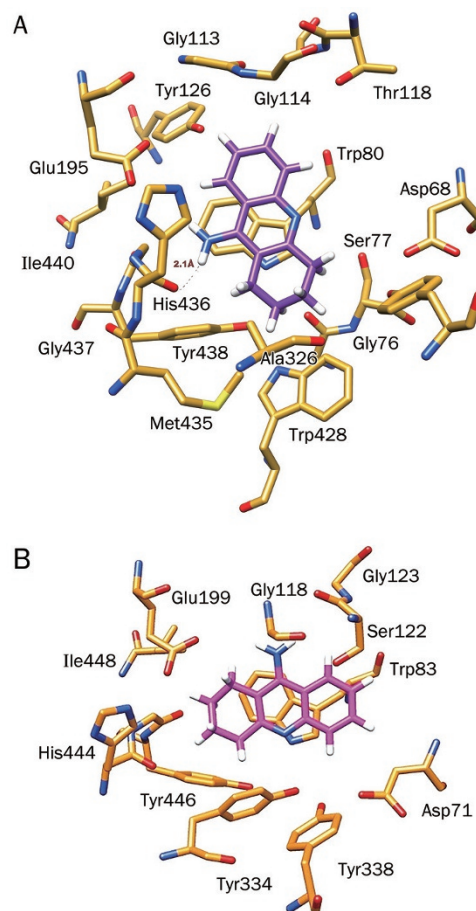


Figure 1. (A) Docked pose of the FDA approved drug tacrine as an active inhibitor inside the binding gorge of human butyrylcholinesterase (hBChE), (B) Docked pose of tacrine with human acetylcholinesterase (hAChE).

mental results. However, for AChE, the important residues of the CAS, mid-gorge and PAS regions were not closely associated with the ligands. Owing to the relatively large distance, we could not establish significant interactions aside from weak π -H interactions between compound **1** (274.4 $\mu\text{mol/L}$) and the Trp86 residue (Figure 3).

Least active compounds (135–250 $\mu\text{mol/L}$)

The molecular structure of **4t** (189.71) reveals a large hydrophobic surface that is ideal for interacting with the hydrophobic lining of the 20 Å deep gorge of BChE. Therefore, these residues can readily form hydrophobic interactions with the hydrophobic surface of **4t**, as well as interactions with the negatively charged, acidic residue Glu195 at a distance of 2.083 Å (Figure 2) that contributed to the binding affinity of **4t** to BChE. In the case of **4a**, **4b**, and **4c**, docking analysis specified that the size of the halogen had an effect on the inhibitory potency of these compounds against BChE. Eight hydrophobic active site residues of BChE were within 5 Å of **4t**, whereas only six hydrophobic residues were contained within that radius of the binding pocket in AChE. This observation may account for the relatively stronger binding of **4t** to BChE

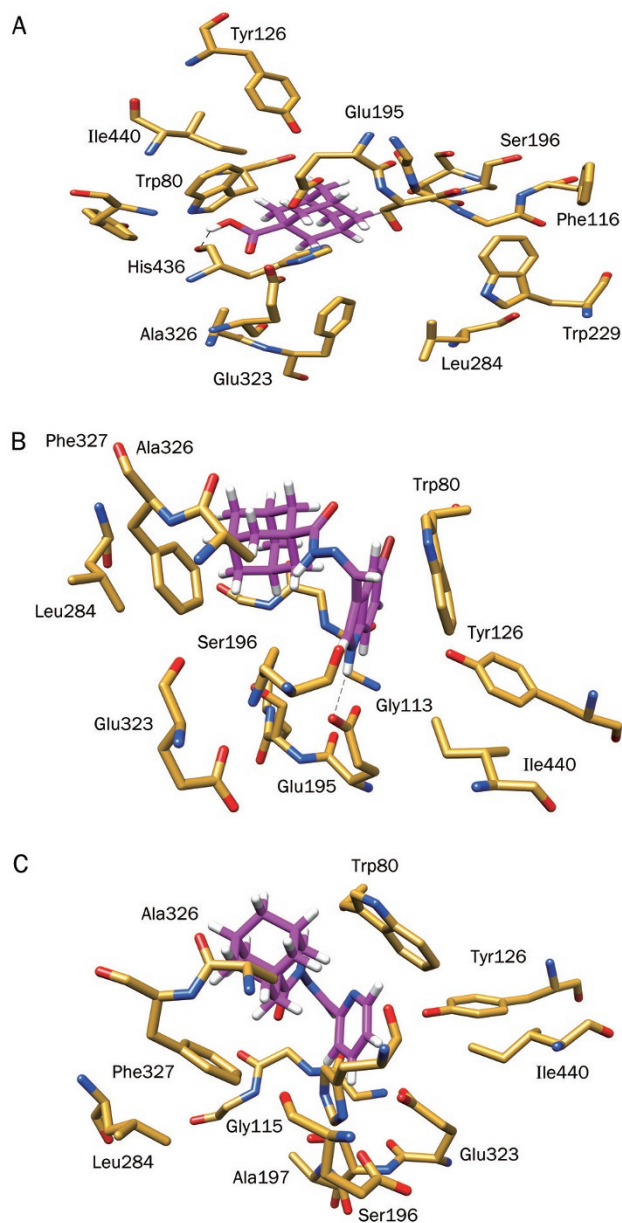


Figure 2. Putative binding mode of selected adamantyl derivatives according to their inhibition potency inside the binding gorge of human butyrylcholinesterase (hBChE): (A) active compound **1**; (B) least active compound **4t**, and (C) inactive compound **4o**.

than AChE (Figure 3). However, **4t** also has the potential to form electrostatic interactions with active site residues of both enzymes, therefore, the inhibitory activity of **4t** for each protein may differ primarily by hydrophobic interactions.

Inactive (>250 $\mu\text{mol/L}$)

Compound **4o** (327 $\mu\text{mol/L}$) showed no-inhibitory potency towards BChE and has no H-bond interaction or other interactions to stabilize the complex system (Figure 2). Similarly, this compound was also inactive against AChE (Figure 3). Comparison of **4o** with **4t** reveals that the orientation of the ada-

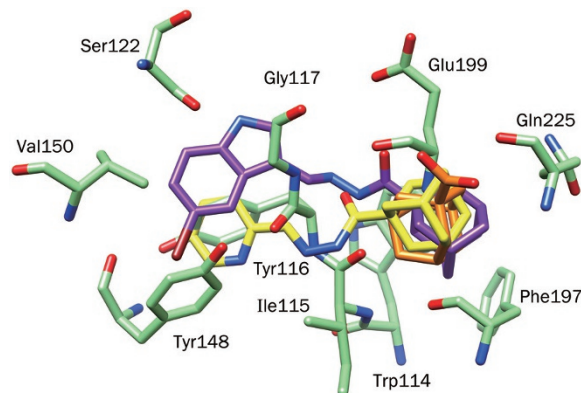


Figure 3. Representation of the top-scored docked poses of selected adamantyl derivatives to understand the binding pattern within the human acetylcholinesterase active site for compound **1** (orange), least active **4t** (magenta) and inactive compound **4o** (yellow).

mantyl moiety of **4o** is relatively far away from the interactive hydrophobic residues because of the pyridine ring. This factor may be responsible for the lack of potency observed against both the cholinesterase enzymes.

Conclusion

Condensation of adamantane-1-carboxylic acid hydrazide with the corresponding aldehydes produced a series of N-arylidene-1-adamantylcarbohydrazides in good yield. Screening the inhibitory activities of the compounds against each cholinesterase enzyme showed that all compounds were inactive towards AChE, but that several showed moderate activity towards BChE. Compound (**3**) shows an almost equal inhibitory activity towards both enzymes. The relatively low enzyme inhibition activities of the compounds may be due to steric effects and the large size of hydrazones. Additionally, theoretically estimated binding affinities are close to the range of experimental values. According to the binding modes, the hydrophobic interactions between adamantyl derivatives and their surrounding residues of the active site play predominant roles, while hydrophilic interactions were also found. When different adamantyl derivatives (**1-26**) were docked inside each cholinesterase enzyme (BChE and AChE), adamantyl compounds were selectively observed to inhibit BChE over AChE, possibly due to the larger active site of BChE.

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Author contribution

Amal AL-ABOUDI has designed the research, wrote the paper and performed the *in vitro* enzyme inhibition studies; Raed A AL-QAWASMEH has designed the synthesis, supervised it and analyzed the spectral data; Alaa SHAHWAN has per-

formed the synthesis; Uzma MAHMOOD has performed docking studies; Asaad KHALID has supervised the *in vitro* enzyme inhibition studies, and analyzed the results; Zaheer UL-HAQ has supervised the *in silico* studies and analyzed the results.

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