

Original Article

Isoindolone derivative QSN-10c induces leukemic cell apoptosis and suppresses angiogenesis via PI3K/AKT signaling pathway inhibition

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Aim: 2-(4,6-Dimethoxy-1,3-dioxoisindolin-2-yl) ethyl 2-chloroacetate (QSN-10c) is one of isoindolone derivatives with antiproliferative activity against human umbilical vein endothelial cells (HUVECs). The aim of this study was to investigate its antitumor activity *in vitro* and anti-angiogenic effects *in vitro* and *in vivo*.

Methods: K562 leukemic cells and HUVECs were used for *in vitro* studies. Cell viability was examined using MTT assay. Cell apoptosis and mitochondrial transmembrane potential ($\Delta\psi_m$) were detected with flow cytometry. Tube formation and migration of HUVECs were studied using two-dimensional Matrigel assay and wound-healing migration assay, respectively. VEGF levels were analyzed with RT-PCR and Western blotting. A zebrafish embryo model was used for *in vivo* anti-angiogenic studies. The molecular mechanisms for apoptosis in K562 cells and antiangiogenesis were measured with Western blotting.

Results: In antitumor activity studies, QSN-10c suppressed the viability of K562 cells and induced apoptosis in dose- and time-dependent manners. Furthermore, QSN-10c dose-dependently decreased the $\Delta\psi_m$ in K562 cells, increased the release of cytochrome c and the level of Bax, and decreased the level of Bcl-2, suggesting that QSN-10c-induced apoptosis of K562 cells was mediated via the mitochondrial apoptotic pathway. In anti-angiogenic activity studies, QSN-10c suppressed the viability of HUVECs and induced apoptosis in dose dependent manners. QSN-10c treatment did not alter the $\Delta\psi_m$ in HUVECs, but dose-dependently inhibited the expression of VEGF, inhibited the tube formation and cell migration *in vitro*, and significantly suppressed the number of ISVs in zebrafish embryos *in vivo*. Furthermore, QSN-10c dose-dependently suppressed the phosphorylation of AKT and GSK3 β in both HUVECs and K562 cells.

Conclusion: QSN-10c is a novel antitumor compound that exerts both antitumor and anti-angiogenic effects via inhibiting the PI3K/AKT/GSK3 β signaling pathway.

Keywords: isoindolone; anticancer drug; leukemia; zebrafish; angiogenesis; apoptosis; AKT; GSK3 β

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Introduction

Angiogenesis, the formation of new blood vessels from pre-existing vasculature, occurs under a variety of physiological and pathological conditions. Accumulating evidence indicates that angiogenesis is especially critical for the growth and progression of solid tumors. The growth of a tumor mass beyond 2 to 3 mm³ is often preceded by increased formation of new blood vessels essential for the delivery of nutrients and oxygen as well as the removal of metabolic waste from the tumor microenvironment^[1,2]. Moreover, many recent stud-

ies have demonstrated the important role of angiogenesis in hematologic neoplasms^[3-5]. Therefore, anti-angiogenic and angio-prevention therapies represent promising approaches to control tumor growth and metastasis^[6,7]. Neoangiogenesis is a highly ordered, multistep molecular process tightly regulated in endothelial cells^[8,9] that involves the proliferation of endothelial cells (ECs), motility, cell realignment, vessel formation, and the production of a new basement membrane^[1,10,11]. Of the numerous growth factors and cytokines that have been shown to possess angiogenic effects, vascular endothelial growth factor (VEGF) is an attractive therapeutic target for the development of novel anticancer agents^[12]. VEGF is produced in tumor and vascular cells, thus stimulating angiogenesis through an autocrine fashion^[13]. The inhibition of VEGF expression in endothelial cells contributes to reduced

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angiogenesis in HUVECs. In endothelial cells, VEGF has been shown to exert mitogenic activities and vasopermeability responses primarily through VEGFR-2^[14], thereby promoting endothelial cells proliferation and migration via increased phosphorylation of extracellular signal-regulated kinase 1/2 (ERK1/2) (proliferation), phosphorylated-serine/threonine protein kinase family protein kinase B (AKT) (migration and survival), and c-Jun N-terminal kinase (JNK) (vascular permeability)^[15–18].

In our search for tumor inhibitors from natural sources, we identified 4,6-dihydroxyisoindol-1,3-dione (Figure 1) as a new compound isolated from *Lasiosphaera fenlzii* Reich. Our previous studies demonstrated that this compound has enhanced anti-angiogenic activity compared with thalidomide via inhibition of VEGF secretion. However, the compound exhibits only a slight inhibitory effect on tumor cell proliferation^[19]. To find a compound with antitumor and anti-angiogenic activities *in vitro*, a series of isoindolone derivatives were subsequently synthesized, and their cytotoxicities were preliminarily evaluated in A549 and HUVEC cells. A new compound, 2-(4,6-dimethoxy-1,3-dioxoisoindolin-2-yl) ethyl 2-chloroacetate (QSN-10c, Figure 1), demonstrated moderate antiproliferative activities in HUVECs^[20]. The purpose of the present study is to examine the effects of QSN-10c on angiogenesis *in vitro* (VEGF expression, tube formation, and HUVEC migration) and *in vivo* (zebra embryo model) as well as the possible mechanisms involved [phosphorylation of proteins involved in the AKT signaling pathway and the mitogen-activated protein kinase pathway (MAPK)]. In addition, QSN-10c also demonstrated enhanced cytotoxicity in K562 leukemic cells in our antitumor screening. Consequently, the mechanism for apoptotic induction in K562 cells is also reported here.

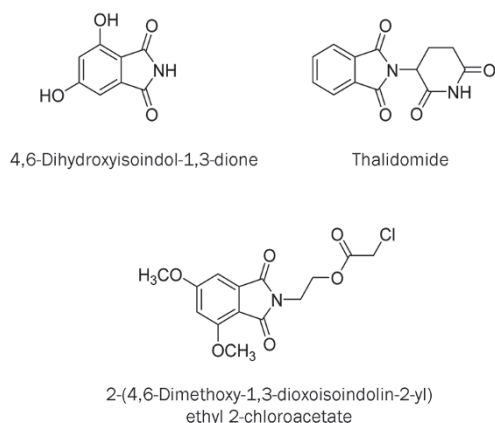


Figure 1. The chemical structures of 4,6-dihydroxyisoindol-1,3-dione, thalidomide and QSN-10c.

Materials and methods

Reagents

QSN-10c was prepared by the Key Laboratory of the Ministry of Education of Shenyang Pharmaceutical University with

greater than 98% purity. Thalidomide was a gift from the Changzhou Pharmaceutical Co, Ltd (China). Antibodies for anti-phospho-AKT (Thr³⁰⁸) and anti-AKT were from Bioworld Technology Inc (Minneapolis, USA). Antibodies for anti-phospho-GSK3 β (Ser⁹), phospho-p44/42 MAPK (ERK1/2) (Thr202/Tyr204), phospho-SAPK/JNK (Thr183/Tyr185), anti-GSK3 β , anti-p44/42 MAPK (ERK1/2) (Thr202/Tyr204), and anti-SAPK/JNK were from Cell Signaling Technology Inc (USA). Antibodies for cytochrome *c* (A-8), Bcl-2 (C21), Bax (P-19), VEGF (147), and anti-mouse β -actin (C4) were from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The secondary antibodies were purchased from ZSGB-BIO (Beijing, China). The enhanced chemiluminescent (ECL) plus reagent kit was obtained from the Beyotime Institute of Biotechnology (Haimen, China).

Cell proliferation assay

HUVEC and K562 cells dispersed evenly in medium (containing 10% FBS) were seeded and incubated in a 96-well plate at a density of 5×10^3 cells/well for 24 and 1 h, respectively. The cells were then treated with various concentrations of QSN-10c, thalidomide, and 4,6-dihydroxyisoindol-1,3-dione for 24 h with three replicates for each treatment. Methylthiazolyldiphenyl-tetrazolium bromide (MTT) was added to each well for 4 h. After discarding the supernatant, the insoluble formazane product was dissolved in 150 μ L DMSO. The optical density was measured at 490 nm using an Elx800 microplate reader (Bio-Tek, USA). The percentage of viable cells was calculated using the following equation: $A_s/A_c \times 100\%$, where A_c is the absorbance of the negative control and A_s is the sample absorbance.

K562 cells were seeded in 6-well plates at a density of 3×10^5 cells per well for 1 h and then treated with DMSO (0.1%) and 100 μ mol/L QSN-10c for 6, 12, 24, and 48 h. To count the cells, the viable cells were distinguished using trypan blue dye. The number of viable cells was counted. The percentage of viable cells was calculated using the following equation: viable cells (sample)/viable cells (control) $\times 100\%$. The assay was independently repeated thrice.

Apoptosis detection by flow cytometry

The sub-diploid (sub-G₁) population was analyzed by flow cytometry using propidium iodide (PI, sigma) staining as previously described^[21]. Briefly, HUVEC and K562 cells were treated with 100 μ mol/L QSN-10c for 0, 6, 12, 24, and 48 h. The cells were centrifuged at 1500 rounds per minute for 10 min at 4°C, resuspended in PBS, fixed via the drop-wise addition of ice-cold ethanol (100%) to a final concentration of 70% and incubated for 30 min on ice. The fixed cells were pelleted and treated with 500 μ L propidium iodide working solution (100 μ g/mL RNase A and 100 μ g/mL propidium iodide in PBS) for 30 min at room temperature protected from light. DNA content was assessed using a FACScan flow cytometer (Becton Dickinson, Heidelberg, Germany). Annexin V staining was also used to detect apoptotic cells according to the FITC Annexin V Apoptosis Detection Kit (Becton, Dickinson

Company) instructions. The samples were analyzed by flow cytometry.

Measurement of mitochondrial transmembrane potential in flow cytometry

Mitochondrial membrane potential ($\Delta\psi_m$) was measured using Rhodamine 123 (Rho 123) staining (Sigma). Briefly, treated cells were collected and washed twice with cold PBS. Finally, cells were stained with Rho123 (10 $\mu\text{g}/\text{mL}$) for 30 min and measured using a FACScan flow cytometer.

Endothelial cell capillary-like tube formation assay

Matrigel™ Basement Membrane Matrix (growth factor reduced; BD Biosciences, San Jose, CA, USA) was thawed at 4°C, pipetted into prechilled 96-well plates (50 μL Matrigel/well), and incubated at 37°C for 30 min. A total of 1.6×10^4 HUVECs dispersed evenly in medium containing 1% FBS were placed onto the Matrigel layer in each well. The cells were treated with DMSO (0.1%) or various concentrations of QSN-10c (25, 50, 100, and 200 $\mu\text{mol}/\text{L}$). After 8 h of incubation, the network-like structures of endothelial cells were examined using an inverted microscope (Olympus, Tokyo) at 100× magnification. The tube-like structures were defined as endothelial cord formations connected at both ends^[22]. The average number of branching points in five random fields per well was quantified using Image Pro-Plus 6.0 software. Cells treated with DMSO (0.1%) only served as the vehicle control. The inhibition percentage was expressed as the percentage of the vehicle control (100%). The assay was independently repeated thrice.

HUVEC migration in a wound-healing assay

An *in vitro* wound-healing assay was performed to measure the unidirectional migration of HUVECs. HUVECs were seeded at 5×10^5 cells per well into a 6-well plate and allowed to grow to full confluence. After starvation with 1640 medium containing 1% FBS for 12 h to inactivate cell proliferation, the cells were wounded with a pipette tip, and washed with PBS. Fresh medium was added to the wells with or without DMSO (0.1%) or various concentrations of QSN-10c (50, 100, and 200 $\mu\text{mol}/\text{L}$) or thalidomide (100, 200 $\mu\text{mol}/\text{L}$), and the cells were incubated for 12 h. Images were obtained at the time of the wounding and at 12-h intervals thereafter using a phase-contrast microscope (Olympus, Tokyo; 100× magnification). Cell migration was quantified according to methods reported in the literature^[23]. The lesion area in each field of view was measured using the data from time 0 (T_0 wound area), and the wound area was then converted to the mean % recovery (%R) using the following equation:

$$\%R = [1 - (\text{wound area at } T_t / \text{wound area at } T_0)] \times 100$$

where T_0 is the wounded area 0 h and T_t is the wounded area 12 h post-injury. All experiments were performed in triplicate, and each experiment was repeated at least thrice.

The detection of VEGF mRNA levels using RT-PCR

VEGF mRNA levels were detected as previously reported^[24].

HUVECs in logarithmic growth phase were seeded into 25-mL culture bottles at a density of $2 \times 10^5/\text{mL}$. The cells were divided into control and experimental groups containing 50, 100, and 200 $\mu\text{mol}/\text{L}$ QSN-10c. Total cellular RNA was isolated by TRIzol reagent extraction (TaKaRa Biotech, China) at the indicated times. Total RNA (5 μg) was reverse transcribed into cDNA using the TIANScript RT reagent kit (Tiagen Biotech Co, Ltd, Beijing). The following primers were designed and synthesized by Takara Bio: VEGF-A sense, 5'-TTG CCT TGC TGC TCT TCC TC-3' and VEGF-A antisense, 5'-TGC ATG GTG ATG TTG GAC TC-3' (280 bp product); GAPDH sense, 5'-GCA CCG TCA AGG CTG AGA AC-3' and GAPDH antisense, 5'-ATG GTG GTG AAG ACG CCA GT-3' (142 bp product). The following reaction conditions were used: 95°C for 5 min; 30 cycles of 94°C for 30 s, 63°C for 30 s, and 72°C for 30 s; extension at 72°C for 7 min. For the PCR product analysis, VEGF and GAPDH PCR products from the same sample were observed using 2% agarose gel electrophoresis. The images were obtained using a gel imaging system, and the target gene expression levels were measured via semi-quantitative analysis.

The relative mRNA content = accumulative photon value of the target gene band / accumulative photon value of the internal standard GAPDH band.

Western blotting

Adherent cells were washed thrice with ice-cold phosphate buffered saline (PBS) before lysed in RIPA buffer containing the following protease and phosphatase inhibitors: aprotinin (10 mg/mL), leupeptin (10 mg/mL), phenylmethylsulfonyl fluoride (1.72 mmol/L), NaF (100 mmol/L), and NaVO_3 (500 mmol/L). K562 cells were collected and then lysed as described above. Equal amounts of protein were separated by 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis and transferred onto PVDF membranes. The blots were incubated with primary antibodies followed by secondary antibody conjugated to horseradish peroxidase; antibody binding was detected with BeyoECL Plus.

Anti-angiogenic activity in the zebrafish embryo model

The *in vivo* anti-angiogenic activity of QSN-10c was further investigated in the zebrafish embryo according to standard operation practices^[25]. Briefly, zebrafish embryos were generated by natural pair-wise mating and raised at 28.5°C in embryo water (5 mmol/L NaCl, 0.17 mmol/L KCl, 0.4 mmol/L CaCl_2 , 0.16 mmol/L MgSO_4). QSN-10c was diluted in DMSO as needed and then transferred to the embryo water. Healthy, hatched zebrafish embryos were selected at 24 hpf (hours post fertilization), and treated with DMSO (0.1%) or varying concentrations of QSN-10c (5, 10, and 20 $\mu\text{g}/\text{mL}$). The embryos were then incubated in 24-well plates (10–15 embryos/well) at 28.5°C from 24 to 48 hpf. At 48 hpf, the zebrafish were removed from the 24-well plates, and viability and vessel morphological changes were assessed. The zebrafish section just below the yolk sac was selected for the assessment of complete intersegmental vessels (ISVs) by manual

counting. ISVs with blood flow were counted as effective; therefore, images could not be obtained. Embryos treated with DMSO (0.1%) served as the vehicle control. The assay was independently repeated thrice. The percentage of inhibition ratio = $[1 - (ISV_{S(\text{sample})} / ISV_{S(\text{control})})] \times 100\%$.

Statistical analysis

The results were expressed as the mean \pm SD. Statistical significance was compared between the QSN-10c-treated and control groups using Student's *t*-test. Results with $P < 0.05$ were considered significantly different from the control. The significance of the difference between two groups was assessed using Student's *t*-test.

Results

QSN-10c decreases the viability of HUVEC and K562 cells

To elucidate the anti-angiogenic effects of QSN-10c *in vitro*, the viability of human umbilical vein endothelial cell (HUVEC) treated with QSN-10c for 24 h was evaluated. In addition, the effect of QSN-10c on K562 cell viability was also examined. Similar to our previous studies, QSN-10c induced minimal reduction in the growth rate of HUVEC cells with an IC_{50} value greater than 150 $\mu\text{mol/L}$ (Figure 2A). QSN-10c inhibited K562 cell viability with an IC_{50} value of $114.0 \pm 2.3 \mu\text{mol/L}$; this effect is greater than that observed for thalidomide and 4,6-dihydroxyisoindol-1,3-dione (Figure 2B). The decrease in cell viability upon treatment with 100 $\mu\text{mol/L}$ QSN-10c was time-dependent in K562 cells (Figure 2C).

QSN-10c induces apoptosis in a time- and concentration-dependent manner in K562 cells

The sub-diploid (sub- G_1) population after PI staining was quantified to determine the extent of apoptosis induced by QSN-10c. A time-dependent increase in sub- G_1 cells was visible after treatment with 100 $\mu\text{mol/L}$ QSN-10c (Figure 3A, 3B, 6, 12, 24, and 48 h after treatment resulted in $3.47\% \pm 0.11\%$, $4.31\% \pm 0.46\%$, $5.38\% \pm 0.51\%$, $11.27\% \pm 0.73\%$ sub- G_1 cells, respectively, *vs* $1.75\% \pm 0.17\%$, $2.10\% \pm 0.28\%$, $2.16\% \pm 0.29\%$, $2.25\% \pm 0.21\%$ in the respective control groups). However, no significant apoptotic induction was observed in HUVEC cells treated with the same QSN-10c concentration (data not shown).

To further characterize QSN-10c-induced apoptosis in K562 cells, annexin V staining was performed. The results demonstrate an approximate 3- to 11-fold increase in the apoptotic fraction of the groups treated with 50–150 $\mu\text{mol/L}$ QSN-10c compared with the control (Figure 3C, 3D). Whereas, 100–200 $\mu\text{mol/L}$ QSN-10c displayed minimal induction of apoptosis in HUVEC cells (Figure 3E).

QSN-10c-induced apoptosis is accompanied by mitochondrial changes and mitochondria-related protein alterations

Many anti-neoplastic drugs induce apoptosis in cancer cells via the mitochondrial apoptotic pathway^[26–29]. A hallmark of death induction via this pathway is a rapid and early loss of the mitochondrial membrane potential ($\Delta\psi_m$). Thus, we ana-

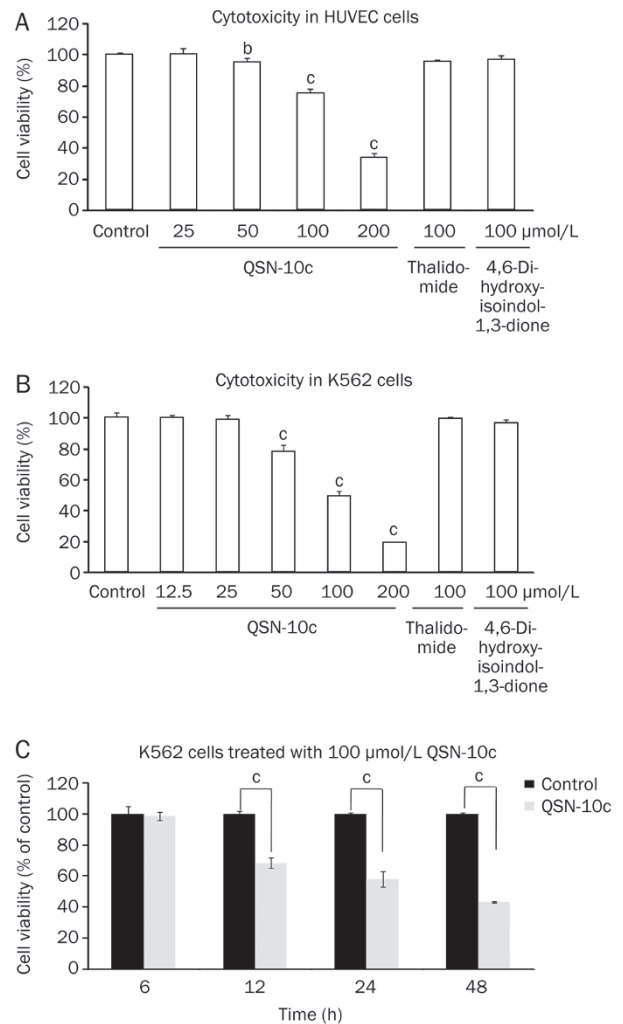


Figure 2. Effects of QSN-10c on viability of HUVECs and K562 cells. (A) Antiproliferative activity in HUVECs after 24 h exposure. (B) Cytotoxicity in K562 cells after 24 h exposure. QSN-10c inhibited the viability of K562 cells in a dose-dependent manner. (C) K562 cells were cultured in 6-well plates for different time periods in the presence of 100 $\mu\text{mol/L}$ QSN-10c. The viability was decreased in a time-dependent manner. Data were expressed as mean \pm SD. $n=3$. ^b $P < 0.05$, ^c $P < 0.01$ compared with control. Thalidomide was given as positive control.

lyzed the integrity of mitochondrial function after treatment with QSN-10c. QSN-10c treatment resulted in a significant concentration-dependent breakdown of the $\Delta\psi_m$ (Figure 4A, 48 h after treatment with 50, 100, or 150 $\mu\text{mol/L}$ QSN-10c). HUVEC cells treated with QSN-10c showed no change in $\Delta\psi_m$ (Figure 4B).

Given that changes in the mitochondrial membrane potential are typically associated with increased permeability of the outer mitochondrial membrane that promotes efflux of apoptogenic proteins to the cytosol^[30], we measured the release of cytochrome *c* from the mitochondria to the cytosol by Western blot. As expected, the expression of cytoplasmic cytochrome *c* increased in a dose-dependent manner (Figure 4C). Changes in mitochondrial membrane potential can result from the

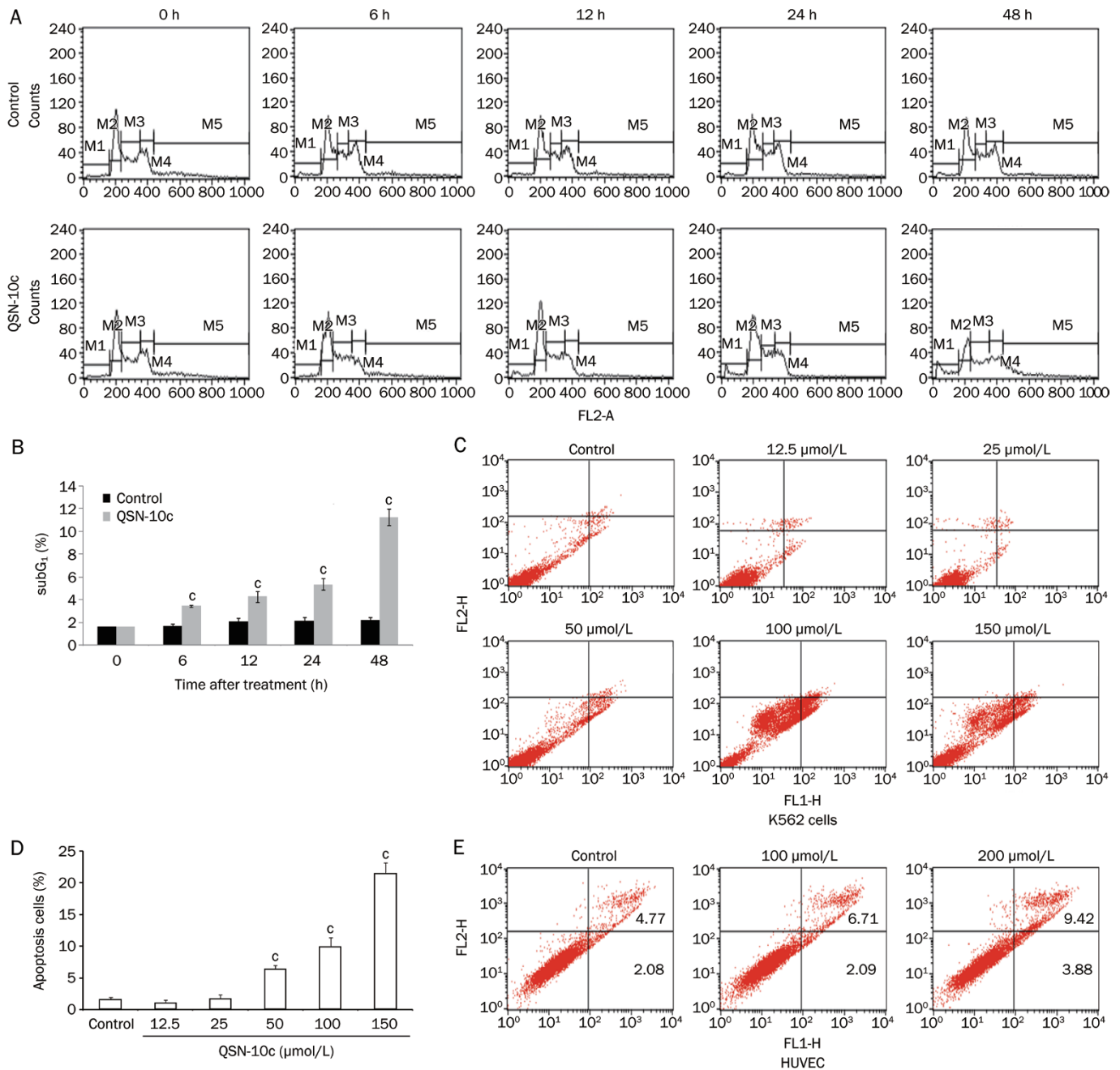


Figure 3. QSN-10c increased apoptosis in K562 cells and HUVECs. PI staining (A–B) and FITC-conjugated Annexin V/PI staining (C–D) were conducted to evaluate the cell apoptosis induced by QSN-10c. (A) Representative patterns of flowcytometric distribution of K562 cells treated with QSN-10c for different time. M1 marks subG₁ apoptotic cells. (B) Quantitative comparison of the numbers of subG₁ phase in different groups. (C) Representative patterns of flowcytometric distribution of K562 cells. Dot plot was divided into a quadrant: upper left, necrotic cells; upper right, late apoptotic cells; lower left, living cells; and lower right, early apoptotic cells. (D) Graphs demonstrate the sum of cells in early and late apoptosis of K562 cells. (E) Representative patterns of flowcytometric distribution of HUVEC. Cells receiving only DMSO (0.1%) served as a vehicle control. Data were expressed as mean±SD. *n*=3. [°]*P*<0.01 vs control.

action of pro- and/or anti-apoptotic members of the Bcl-2 family^[31]. Thus, we examined the effects of QSN-10c on Bcl-2 and Bax protein expression. The Western blot revealed that QSN-10c reduces the expression of the anti-apoptotic protein Bcl-2 and increases the levels of the pro-apoptotic protein Bax in a dose-dependent manner (Figure 4C).

The effects of QSN-10c on tube formation in HUVECs

Although angiogenesis is a complex procedure involving various cell types, endothelial cell tube formation is a key step of angiogenesis^[32]. To further assess the effects of QSN-10c on endothelial cell tube formation, we used a two-dimensional Matrigel assay to examine the effect of QSN-10c on HUVEC

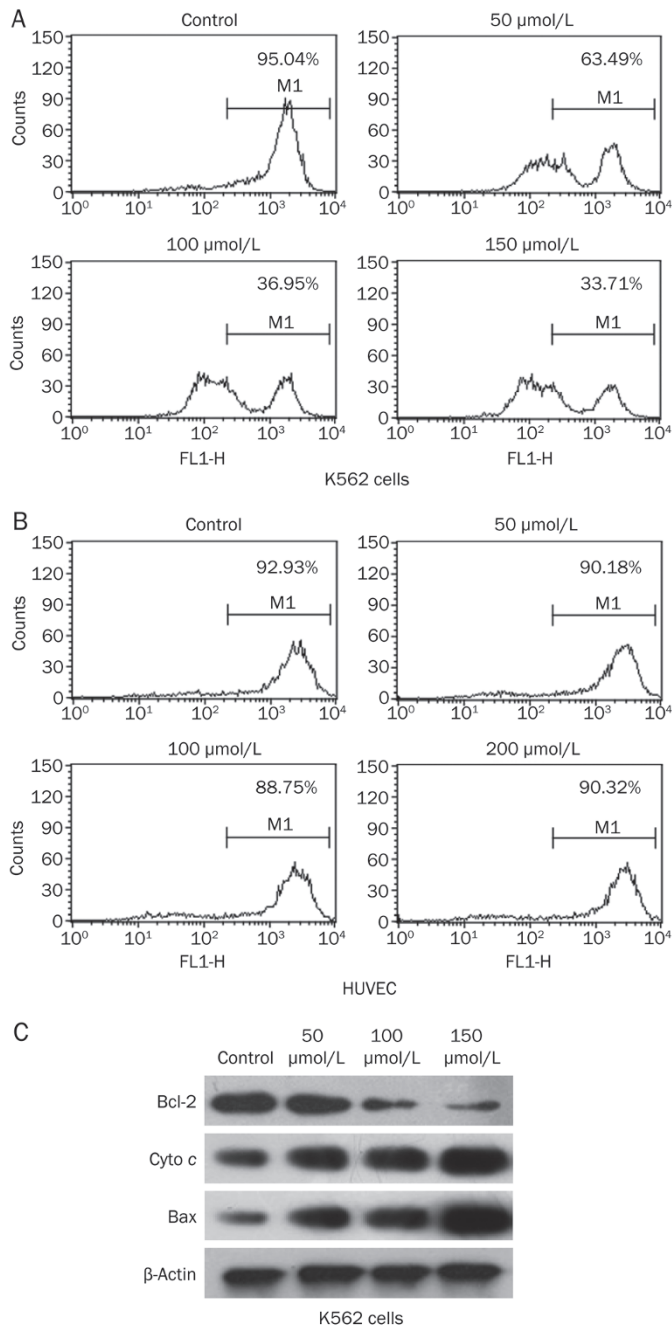


Figure 4. Effects of QSN-10c on mitochondrial membrane potential and expression of mitochondria-related apoptotic proteins. (A–B) The mitochondrial membrane potential of K562 cells treated with QSN-10c was decreased in a dose-dependent manner. (C) Treatment of cells with various concentrations of QSN-10c for 48 h resulted in dose-dependent increase in the release of cytochrome c, and in the expression of Bax. The level of Bcl-2 was decreased in a dose-dependent manner.

tube formation. When HUVECs were seeded on the Matrigel, robust tubular-like structures were formed. Capillary tube formation on Matrigel was inhibited by approximately 7.4% after treatment with 25 μmol/L QSN-10c for 8 h ($P>0.05$). Tube formation was inhibited by approximately 50% with 50 μmol/L

QSN-10c, and tube formation was almost completely inhibited upon treatment with 200 μmol/L. The QSN-10c-mediated effects were more robust than the results of treatment with 4,6-dihydroxyisoindol-1,3-dione and thalidomide (Figure 5A–5G, $P<0.05$, $P<0.01$).

The effects of QSN-10c on HUVEC migration

As cell migration is also essential for angiogenesis in endothelial cells^[33], we investigated the inhibitory effects of QSN-10c on the chemotactic motility of endothelial cells using a wound-healing migration assay. We discovered that QSN-10c inhibits HUVEC migration in a dose-dependent manner, with significant inhibition at 100–200 μmol/L (Figure 6A–6G, $P<0.01$, $P<0.001$). These findings indicate that QSN-10c may inhibit angiogenesis *in vitro* by preventing endothelial cell migration and tubular structure formation.

VEGF levels are decreased in HUVEC cells

To investigate whether QSN-10c modulates the VEGF-mediated autocrine effects on endothelial cells, RT-PCR and Western blot were performed to examine VEGF expression in HUVECs. The results indicated that 24 h of treatment with 50–100 μmol/L QSN-10c clearly decreased VEGF mRNA levels (Figure 7A, 7B, $P<0.05$). We further determined the effect of QSN-10c on VEGF expression by Western blot. QSN-10c decreased VEGF protein in a dose-dependent manner compared with the control (Figure 7C). These results suggest that QSN-10c-mediated VEGF inhibition potentially contributes to reduced angiogenesis by HUVECs.

The PI3K/AKT/GSK3β pathway is involved in QSN-10c-mediated antiangiogenesis and K562 cell apoptosis

To investigate the molecular mechanism of QSN-10c-mediated antiangiogenesis, we examined the effect of QSN-10c on the activity of AKT and GSK3β, important components of the PI3K/AKT signaling pathway that plays a crucial role in HUVEC migration^[34]. Our data indicated that AKT and GSK3β phosphorylation were significantly reduced by QSN-10c in a dose-dependent manner in HUVEC cells. However, QSN-10c did not affect the phosphorylation of ERK1/2 and JNK, signal proteins involved in the MAPK pathway (Figure 8A). To investigate the role of the PI3K/AKT signaling pathway in QSN-10c-mediated apoptosis in K562 cells, we also examined the expression of AKT and GSK3β in K562 cells treated with various concentrations of QSN-10c. The result suggest that QSN-10c also inhibits the expression of p-AKT and p-GSK3β in a dose-dependent manner (Figure 8B), indicating that QSN-10c induces K562 cell apoptosis at least in part via inhibition of the PI3K/AKT pathway.

Anti-angiogenic activity in zebrafish embryo model

Based on QSN-10c activities *in vitro*, we further evaluated the anti-angiogenic property of QSN-10c in a zebrafish embryo model. Zebrafish embryos were treated with DMSO (0.1%) or various concentrations of QSN-10c (5, 10, and 20 μg/mL) from the shield stage (24 hpf) to 48 hpf. Significant reductions

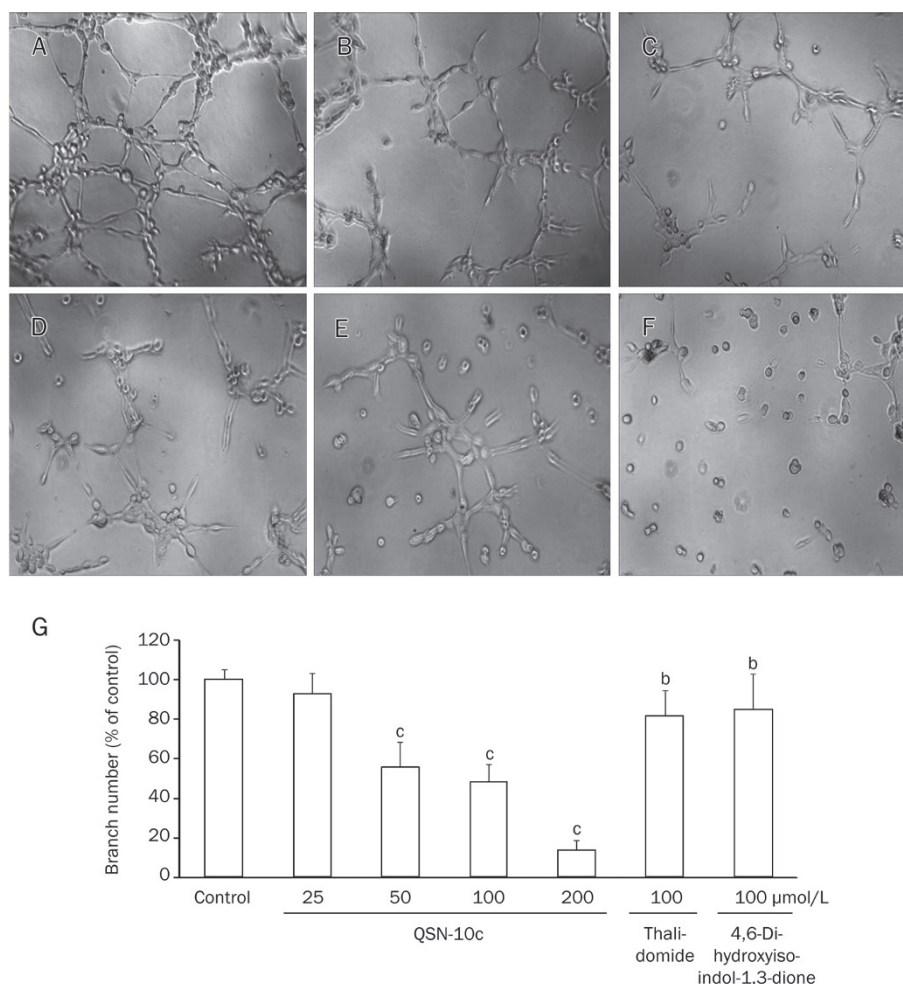


Figure 5. QSN-10c inhibits capillary structure formation of endothelial cells on Matrigel. (A) HUVECs cultured on Matrigel were treated with only DMSO (0.1%). (B, C) HUVECs cultured on Matrigel were treated with 100 μmol/L of thalidomide and 4,6-dihydroxyisoindol-1,3-dione, respectively. (D–F) HUVECs cultured on Matrigel were treated with various concentrations of QSN-10c (50, 100, and 200 μmol/L). (G) Quantitative comparison of the numbers of branching points in different groups. Cells receiving only DMSO (0.1%) served as a vehicle control, thalidomide as a positive control. Data are expressed as percentages of the vehicle control (100%) in mean±SD from three independent experiments. ^b*P*<0.05, ^c*P*<0.01 compared with control.

in the number of complete ISVs were observed in the 10–20 μg/mL QSN-10c-treated groups compared with the vehicle control group (Figure 9, *P*<0.05).

Discussion

Angiogenesis is now recognized as playing a crucial role in the initiation and progression of many diseases, such as cancer and RA, and anti-angiogenic interventions are becoming a promising approach for the treatment of these diseases. Many reported agents possess anti-angiogenic activity via the inhibition of HUVEC proliferation, migration, and tube formation *in vitro*. Moreover, bevacizumab (Avastin, anti-VEGF monoclonal antibody) and vatalanib (VEGFR inhibitor) are approved for the treatment of various malignant tumors by targeting angiogenic pathways. Thalidomide [α -(*N*-phthalimido)-glutaramide] is an immunomodulatory and anti-angiogenic drug. Recent research has shown that the drug has antitumoral properties, and promising results from thalidomide treatment

in patients with myeloma, myelodysplastic syndrome, and variety of solid tumors has been reported^[35]. However, thalidomide has been shown to require bio-activation to exert its anti-angiogenic effect in isolated blood vessels and endothelial cells^[36, 37]. Moreover, thalidomide displayed varying anti-angiogenic activities in endothelial cells (ECs) in the literature^[38, 39]. Thus, the specific mechanism of antitumoral action for thalidomide remains unclear. In this study, thalidomide was used as positive control and displayed minimal anti-angiogenic activities, similar to previous findings^[39].

The preliminary comparison between 4,6-dihydroxyisoindol-1,3-dione and thalidomide has been previously performed. Because 4,6-dihydroxyisoindol-1,3-dione exhibits specific anti-angiogenic activities, a series of derivatives were subsequently synthesized, and then their cytotoxicity and anti-angiogenic activity was evaluated in tumor cells to identify candidates with enhanced properties. Among the derivatives, QSN-10c displayed moderate cytotoxicity in A549 and HUVEC

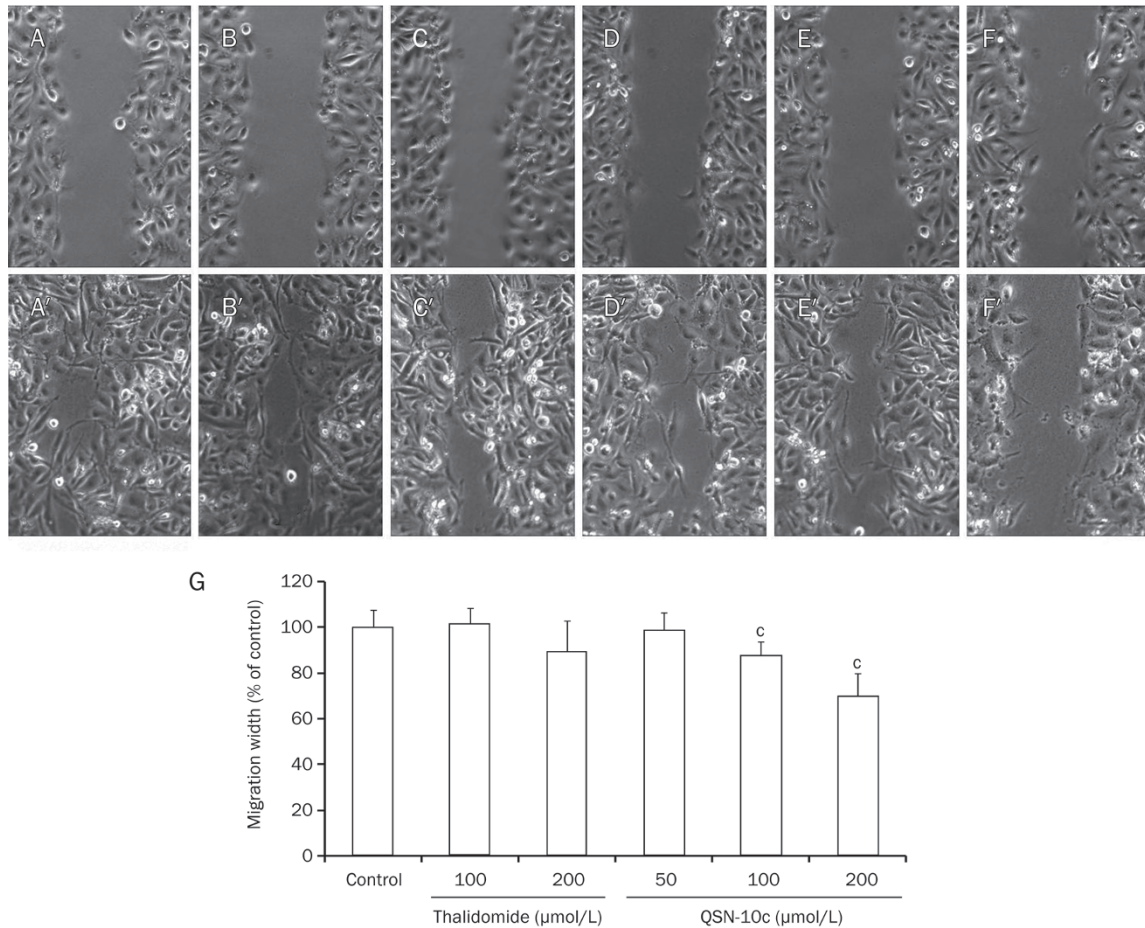


Figure 6. QSN-10c inhibits chemotactic motility of endothelial cells. (A/A') Migration assay of HUVECs treated with only DMSO (0.1%) (0 h/12 h). (B, C/B', C') Migration assay of HUVECs treated with different concentrations (100–200 μmol/L) of thalidomide (0 h/12 h). (D–F/D'–F') Migration assay of HUVECs treated with different concentrations (50–200 μmol/L) of QSN-10c (0 h/12 h). (G) Quantitative comparison of the average migrated width in different groups. Cells receiving only DMSO (0.1%) served as a vehicle control, receiving thalidomide as positive control. Data are expressed as percentages of the vehicle control (100%) in mean±SD from three independent experiments. ^cP<0.01 compared with control.

cells in our previous study. To further elucidate the effects of QSN-10c on angiogenesis and hematologic neoplasms, we employed *in vitro* HUVEC models (migration, tube formation, and VEGF expression) and an *in vivo* zebrafish embryo model. In addition, the cytotoxicity of QSN-10c was investigated in K562 cells. Under the conditions used in this study, QSN-10c exhibited better antiproliferative activity than thalidomide and 4,6-dihydroxyisoindol-1,3-dione in K562 cells. Additional studies suggested that 100 μmol/L QSN-10c induces apoptosis via the mitochondrial apoptotic pathway. In endothelial cells, 50–200 μmol/L QSN-10c also exhibited enhanced inhibition of HUVEC migration and tube formation as well as VEGF expression. Furthermore, we showed that QSN-10c inhibits angiogenesis in an *in vivo* zebrafish model.

With regard to molecular targeting mechanisms, VEGF-mediated biological effects initiate the binding of VEGF to VEGFR-2, thereby resulting in the autophosphorylation of tyrosine residues in the receptors. In fact, the Tg (VEGFR2: GFP) transgenic zebrafish model that expresses fluorescent tags in endothelial cells was first employed to evaluate the

anti-angiogenic activity of QSN-10c. In this transgenic line, QSN-10c showed no inhibitory activity (data not shown), indicating that QSN-10c possibly has no effect on VEGFR2 protein. Consequently, a non-transgenic zebrafish embryo model was used for further studies, and the primary downstream pathways were investigated by Western blot to elucidate in part the anti-angiogenic mechanism of QSN-10c. Our preliminary studies indicate that QSN-10c inhibits the activities of AKT (protein kinase B) and GSK3β, crucial proteins in the PI3K/AKT signaling pathway. PI3K is activated in response to diverse mitogenic signals and catalyzes the formation of the secondary lipid messenger phosphatidylinositol-3,4,5-triphosphate. AKT binds to phosphatidylinositol-3,4,5-triphosphate via pleckstrin homology domains and is recruited to the plasma membrane, where AKT is phosphorylated at Thr³⁰⁸ by phosphoinositide dependent kinase-1 (PDK1) and subsequently activated. Also in K562 cells, the decrease of AKT and GSK3β was found. Interestingly, EC migration is AKT-phosphatidylinositol-3-kinase (PI3K) dependent^[40–42], and the PI3K/AKT pathway transduces anti-apoptotic signals

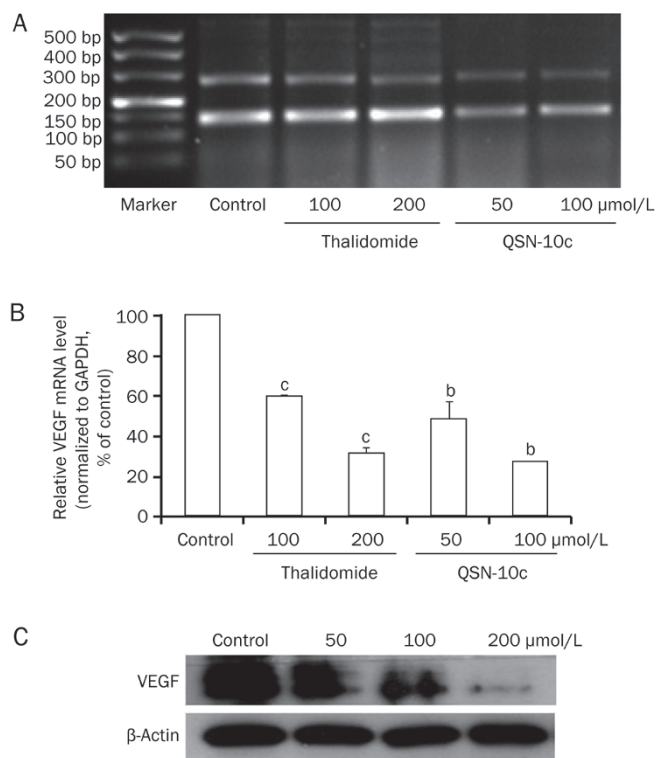


Figure 7. Effects of QSN-10c on VEGF expression in HUVECs. (A) Electrophoresis bands expressed with VEGF and internal standard GAPDH mRNA in various groups treated with different concentrations of thalidomide and QSN-10c. The upper bands were corresponded to VEGF, and the lower to GAPDH. (B) Relative VEGF mRNA levels were quantified by integral optical density analysis of the bands from (A) and then normalization to GAPDH in HUVECs. Cells received thalidomide as positive control. Data are expressed as percentages of the vehicle control (100%) in mean±SD from three independent experiments. ^b $P < 0.05$, ^c $P < 0.01$ compared with control. (C) HUVECs were treated with QSN-10c for 24 h. Cell extract was prepared and subjected to Western blot using anti-VEGF antibody. β -Actin served as a loading control. QSN-10c decreased VEGF protein expression in HUVECs in a dose dependent manner as detected by Western blot.

in tumor cells^[43, 44]. AKT inhibition has been reported to follow the induction of the downstream mitochondrial apoptotic pathway by alterations in the Bcl-2/Bax ratio^[45, 46]. According to increasing evidence regarding the importance of angiogenesis in hematologic neoplasms, the antiproliferative and anti-angiogenic activity of QSN-10c in hematologic K562 cells may support further systematic studies.

VEGF is a potent pro-angiogenic factor that stimulates endothelial cells. Kim KH *et al* found that galbanic acid isolated from *Ferula assafoetida* could inhibit the proliferation of HUVECs stimulated by VEGF better than HUVECs under non-proliferative conditions (*ie*, without additional angiogenic growth factors)^[47]. Given the similarity between QSN-10c and thalidomide as well as the unclear molecular mechanisms of QSN-10c, the effect of QSN-10c on HUVEC proliferation, migration, and capillary structure formation was evaluated without VEGF stimulation. Based on the data from our study, 50–200 μ mol/L QSN-10c demonstrated specific anti-angio-

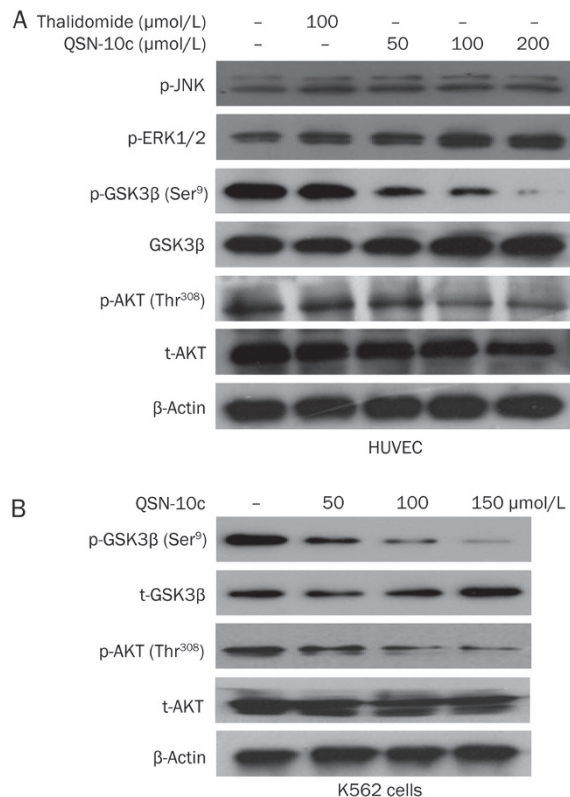


Figure 8. Effects of QSN-10c on phosphorylation of proteins involved in PI3K/AKT and MAPK signaling pathway. (A) HUVEC cells were treated with desired concentrations of QSN-10c for 24 h, cell lysates subjected to Western blot analysis with phospho-Thr³⁰⁸ AKT and total AKT antibodies respectively. Phospho-GSK3 β , JNK, and ERK1/2 were also analyzed. (B) K562 cells were treated with desired concentrations of QSN-10c for 48 h, cell lysates subjected to western blot analysis with phospho-Thr³⁰⁸ AKT, total AKT, phospho-Ser9 GSK3 β and total GSK3 β antibodies, respectively. β -Actin reprobing is showing equal loading of the total protein samples.

genic activities in endothelial cells as well as inhibition of the AKT signaling pathway. We hypothesize that QSN-10c might display enhanced activity under mitogen-activated conditions (*ie*, VEGF stimulation); however, this hypothesis requires further testing.

In conclusion, our findings indicate that QSN-10c inhibits angiogenesis *in vitro*, and this effect is mediated by inhibition of endothelial cell tube formation and migration. The inhibition of these endothelial cell activities is mediated in part through inhibition of the AKT pathway, which plays multiple roles in the regulation of neovascularization. Furthermore, QSN-10c also directly induces apoptosis in K562 leukemia cells via the mitochondrial apoptotic pathway. On the basis of these findings, QSN-10c could serve as an effective candidate for further study. In addition, more derivatives with enhanced efficacy can be designed.

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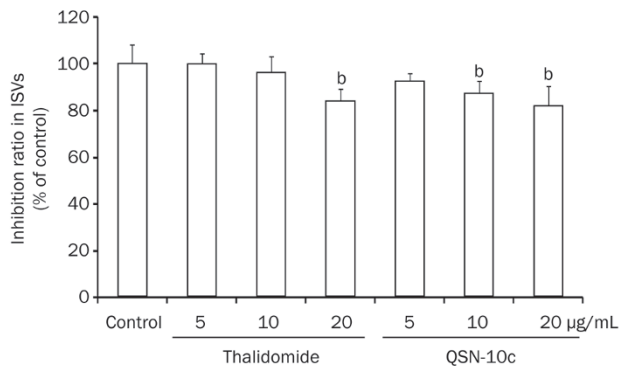


Figure 9. QSN-10c inhibits blood vessel formation in ISVs of zebrafish embryos. Quantitative comparison of blood vessel formation in the vehicle control group and QSN-10c-treated groups. Data are expressed as mean±SD from three independent experiments. ^b $P < 0.05$ compared with vehicle control.

viding the zebrafish embryo research platform.

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