HapMap-based study of the 17q21 ERBB2 amplicon in susceptibility to breast cancer

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ERBB2 is frequently amplified in breast tumours as part of a wide region of amplification on chromosome 17q21. This amplicon contains many candidate genes for breast cancer susceptibility. We used a genetic association study design to determine if common genetic variation (frequency ≥5%) in a 400-kb region surrounding ERBB2 and containing the PPARBP, CRK7, NEUROD2, PPP1R1B, STARD3, TCAP, PNMT, CAB2, ERBB2, C17ORF37, GRB7 and ZNFN1A3 genes, was associated with breast cancer risk. Sixteen tagging single-nucleotide polymorphisms (tSNPs) selected within blocks of linkage disequilibrium from the HapMap database, one HapMap singleton SNP, and six additional SNPs randomly selected from dbSNP were genotyped using Taqman in a large study set of British women (2275 cases, 2280 controls). We observed no association between any of the genotypes or associated haplotypes and disease risk. In order to simulate unidentified SNPs, we performed the leave-one-out cross-validation procedure on the HapMap data; over 90% of the common genetic variation was well represented by tagging polymorphisms. We are therefore likely to have tagged any common variants present in our population. In summary, we found no association between common genetic variation in the 17q21 ERBB2 amplicon and breast cancer risk in British women.

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Only a small proportion of the excess familial risk associated with breast cancer is accounted for by known highly penetrant genes, *BRCA1* and *BRCA2*. The remainder is probably due to a combination of weakly predisposing alleles including both common and rare variants (Pharoah *et al*, 2002; Dite *et al*, 2003). We have previously shown that common alleles in *ERBB2* are not involved in breast cancer susceptibility (Benusiglio *et al*, 2005). However, *ERBB2* is part of a wider region of chromosome 17q21 frequently amplified in breast cancer. This amplicon encompasses many genes and it is conceivable that, as suggested by the variability of response to anti-*ERBB2* therapy in patients with *ERBB2* amplification, more than one gene in the amplicon could contribute to breast cancer susceptibility, development and progression (Willis *et al*, 2003).

Kauraniemi et al (2001, 2003) first carried out a systematic survey of copy number and expression patterns in all genes within the 17q21 locus on breast cancer cell lines and tumour samples; they identified a 200-kilobase (kb) minimal common region of

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amplification around *ERBB2* containing the following genes: neurogenic differentiation 2 (*NEUROD2*), protein phosphatase 1 regulatory subunit 1B (*PPP1R1B*), START domain containing 3 (*STARD3*), titin-cap (*TCAP*), phenylethanolamine-*N*-methyltransferase (*PNMT*), *CAB2* (Per1-Like Domain Containing 1 [*PERLD1*]), *C17ORF37*, growth factor receptor-bound protein 7 (*GRB7*) and zinc-finger protein subfamily 1A 3 (*ZNFN1A3*).

Gene expression levels in cancer cells together with knowledge about protein function can help in assessing which genes are involved in oncogenesis (Futreal et al, 2004). For example, STARD3, PNMT, CAB2, C17ORF37 and GRB7 show a significant correlation between amplification status and expression level (Kauraniemi et al, 2001, 2003; Willis et al, 2003; Orsetti et al, 2004), and STARD3, PNMT, CAB2, GRB7 and ZNFN1A3 could all be biologically relevant to breast cancer. STARD3 mediates intracellular trafficking of cholesterol and can augment steroid hormone synthesis (Strauss, III et al, 2003). Overexpression of PNMT results in suppression of circulating leptin levels - a potent regulator of body weight - in transgenic mice (Bottner et al, 2000; Harvey and Ashford, 2003). CAB2 is a human homologue of the yeast COS16 gene required for the repair of DNA double-strand breaks (Nezu et al, 2002) and GRB7 regulates cell migration through its involvement in cell signalling pathways (Han et al, 2001). Finally, ZNFN1A3 appears to function as a tumour suppressor since its

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downregulation in the mouse leads to leukaemias and lymphomas (Rebollo and Schmitt, 2003).

Two neighbouring genes, peroxisome proliferator-activated receptor binding protein (PPARBP) and CDC2-related protein kinase 7 (CRK7), located about 50 kb upstream from NEUROD2 the first gene on the minimum region of amplification - are often, although less consistently, co-amplified with ERBB2 (Kauraniemi et al, 2001, 2003). Both are potentially implicated in cancer biology. PPARBP, by its ability to function as an oestrogen receptor coactivator, might play a role in mammary epithelial differentiation (Zhu et al, 1999) while CRK7 could link transcription with the splicing machinery (Ko et al, 2001).

The case - control study design is well suited to the identification of small-effect genes that are likely to underlie common, complex diseases such as breast cancer (Risch, 2000). Two approaches have been proposed. The direct, hypothesis-driven approach is to investigate single nucleotide polymorphisms (SNPs), which are thought to have functional effects and thus influence directly the traits under study (Tabor et al, 2002). The indirect, tagging approach is to select a set of empirical tagging SNPs (tSNPs) that best capture the common genetic variation within the gene. They serve as markers to detect associations between a particular region and diseases, whether or not the tSNPs themselves have a functional effect (Gabriel et al, 2002). It is not necessary to genotype all polymorphisms because the alleles of SNPs that are physically close to each other tend to be correlated with each other: they are in linkage disequilibrium (LD) (Pharoah et al, 2004). The HapMap online database (http://www.hapmap.org) allows the tagging approach to be applied readily to many genes or regions (Gibbs et al, 2003). By March 2006, the database held the genotypes of individuals with European, African-American, and Asian ancestry for nearly four million SNPs.

We aimed to determine whether common genetic variation (frequency >5%) in the ERBB2 amplicon is involved in breast cancer susceptibility. We used HapMap data to identify tSNPs for genotyping in a large breast cancer case-control study of white British women. Data for five of the SNPs described in this report have been previously published (Benusiglio et al, 2005), but are also included here for completeness.

METHODS

Patients and controls

Cases were drawn from SEARCH, an ongoing population-based study in which cases are ascertained through the East Anglian Cancer Registry. All patients diagnosed with invasive breast cancer below age 55 years since 1991 and still alive in 1996 (prevalent cases; median age 48 years), together with all those diagnosed < 70 years between 1996 and the present (incident cases; median age 52 years), were eligible to take part. All study participants completed an epidemiological questionnaire and provided a blood sample for DNA analysis. Sixty-seven percent of eligible breast cancer patients returned a questionnaire and 64% provided a blood sample. Controls were randomly selected from the Norfolk component of EPIC (European Prospective Investigation of Cancer). EPIC is a prospective study of diet and cancer being carried out in nine European countries. The EPIC-Norfolk cohort comprises 25 000 individuals resident in Norfolk, East Anglia, the same region from which the cases have been recruited. Controls are not individually matched to cases, but are broadly similar in age, being aged 42-81 years. The ethnic background of both cases and controls as reported on the questionnaires is similar, with >98% being white. All participants have given written consent and the study is approved by the Eastern Region Multicentre Research Ethics Committee.

A total of 4474 cases, of whom 27% were prevalent cases, and 4560 controls, were available for analysis. The samples have been split into two sets in order to conserve DNA and reduce genotyping costs. The first set (2275 cases, 2280 controls) is genotyped for all SNPs. Any SNP that shows association in set 1 at the P < 0.1 level, can then be tested in the second set (2199 cases, 2280 controls). This staged approach substantially reduces genotyping costs without significantly affecting statistical power (see below). Cases with high yields of genomic DNA were selected for set 1 from the first 3500 recruited, with set 2 comprising the remainder of these plus the next 974 incident cases recruited. As the prevalent cases were the first recruited, the proportion of prevalent cases was somewhat higher in set 1 than set 2 (33 vs 20%). Median age at diagnosis was similar in both sets (51- and 52year-old, respectively). Median time from diagnosis to blood draw was slightly longer for set 2 (15 months) than for set 1 (9 months). There were no significant differences in the morphology, histopathological grade or clinical stage of the cases by set or by prevalent/incident status.

SNP identification and selection

The amplicon of interest is a 400-kb region, with PPARBP in position 5' and ZNFN1A3 in position 3' (Figure 1). A 50-kb segment located between CRK7 and NEUROD2 was excluded as it contained no known gene, splitting the amplicon into a 150-kb region including PPARBP and CRK7 (region A) and a 200-kb region including NEUROD2, PPP1R1B, STARD3, TCAP, PNMT, CAB2, ERBB2, C17ORF37, GRB7 and ZNFN1A3 (region B).

We used data on common SNPs from HapMap (European samples, public release #15) to identify tSNPs. After exclusion of one singleton SNP that was poorly linked with any other SNP (D' <0.3), blocks of LD were defined on the basis of limited haplotype diversity (common haplotypes must account for at least 90% of all haplotypes) (Cardon and Abecasis, 2003). tSNPs were selected within blocks using the tagSNPs program so that unmeasured

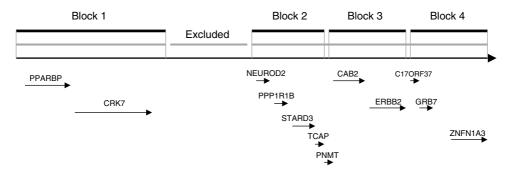


Figure I The 400-kb ERBB2 amplicon. It is split in two (regions A and B) by a 50-kb segment containing no known gene. Region A consists of one LD block while region **B** consists of three blocks, blocks 2, 3 and 4.

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SNPs were tagged with a minimum r_s^2 of 0.8 (Stram, 2004). The r_s^2 coefficient measures LD between unmeasured SNPs and haplotypes defined by the selected tSNPs. The five *ERBB2* SNPs previously genotyped (ERBB2-01 to -05) were forced as tSNPs into the selection algorithm and SNPs surrounded by repetitive DNA sequences were excluded (they could not be selected as tSNPs). The average SNP density of the HapMap SNPs was 1 SNP/5 kb. In order to increase the SNP density and improve the tagging properties of our SNP set, we also selected at the beginning of the study additional random, validated polymorphisms from the dbSNP database (http://www.ncbi.nlm.nih.gov/projects/SNP).

Genotyping

Genotyping was carried out using Taqman (Applied Biosystem, Warrington, UK). Primers and probes were supplied directly by Applied Biosystems, except those for ERBB2-04 and -05 that were designed 'in-house' with the Primer Express Oligo Design Software v2.0 (Applied Biosystems). Sequences are available on request. Reactions were carried out at 54 or 60°C in 384-well plates with cases and controls plated together. Each plate included two negative controls with no DNA and 12 samples duplicated on a separate quality control plate. Plates were read on the ABI Prism 7900 using the Sequence Detection Software (Applied Biosystems). Complete concordance between samples and their duplicates, excluding undetermined genotypes, was required for the assay to be validated. Failed genotypes were not repeated.

Statistical methods

For each SNP, deviation of genotype frequencies in controls from Hardy–Weinberg (HW) equilibrium was assessed by a standard χ^2 test (1 degree of freedom (df)). Genotype frequencies in cases and controls were compared by a χ^2 test for heterogeneity (2 df). Genotype-specific risks were estimated as odds ratios (OR) using standard cross-product ratio and confidence intervals were

calculated using the variance of the log (OR), which was estimated by the standard Taylor expansion. A comparison of haplotype frequencies between cases and controls was carried out using the haplo.score routine implemented in S-plus (Schaid *et al*, 2002). Haplotypes with an estimated frequency of <5% were pooled. Haplo.score uses a likelihood that depends on estimated haplotype frequencies to test the statistical association between haplotypes and phenotype. It is based on score statistics, which provide both global tests and haplotype-specific tests.

RESULTS

The HapMap release #15 included genotypes for 66 common SNPs (26 in region A and 40 in region B). Region A consisted of only one LD block, block 1 with 26 SNPs, whereas region B consisted of three blocks, block 2 with 14 SNPs, block 3 with 11 SNPs and block 4 with 15 SNPs. One singleton SNP, ERBB2-04 (I655V), was excluded from block 3, leaving 10 SNPs in block 3. Block 1 included *PPARBP* and *CRK7*, block 2 included *NEUROD2*, *PPP1R1B*, *STARD3* and *TCAP*, block 3 included *CAB2* and *ERBB2* and block 4 included *C17ORF37*, *GRB7* and *ZNFN1A3* (Figure 1).

Sixteen tSNPs were selected for genotyping: four in block 1, three in block 2, five in block 3 and four in block 4 (Table 1). ERBB2-04 - the HapMap singleton SNP – and six additional SNPs randomly selected from dbSNP (one in block 2, four in block 3 and one in block 4) were also genotyped. None of the genotype distributions in controls differed significantly from those expected under HW equilibrium. There was no evidence that any of the SNPs is associated with breast cancer, and none of the SNPs exceeded the significance threshold for genotyping in the second set of cases and controls (Table 2). The genotype-specific ORs were all close to unity with confidence intervals including one (Figure 2). The tSNPs generated five common haplotypes in block 1, three in block 2, four in block 3 and four in block 4 (Table 3). The global test of association was not significant for any of the four blocks (P=0.16, 0.58, 0.48) and (P=0.16

Table I SNPs selected for genotyping, the database they were selected from, the LD block to which they belong, their location within genes and their frequencies in databases

SNP	rs number	Database	Block	Location	Base change	MAF
PPARBP-01	rs6503513	НарМар	ĺ	START -1240	a>g	0.16
PPARBP-02	rs11655550	НарМар		STOP +2808	t>c	0.24
CRK7-01	rs2303315	НарМар		IVS10 +111	t>a	0.13
CRK7-02	rs4404103	НарМар		IVS13 +88	a>g	0.12
NEUROD2-01	rs12453682	НарМар	2	STOP +5832	t>c	0.28
PPPIRIB-01	rs1874228	НарМар		START -8372	g>a	0.24
PPPIRIB-02	rs879606	НарМар		START -1797	g>a	0.13
STARD3-01	rs3817160	dbSNP		IVSI +331	c>g	0.45
CAB2-01	rs2952151	НарМар	3	START -560	c>t	0.28
CAB2-02	rs907087	dbŚNP		START -269	a>g	0.46
CAB2-03	rs1565920	dbSNP		IVS5 +681	a>g	0.38
CAB2-04	rs907089	dbSNP		IVS5 +2668	a>g	0.39
CAB2-05	rs1476278	НарМар		IVS5 +5311	a>g	0.29
ERBB2-01	rs4252596	dbSNP		START -657	c>a	0.07
ERBB2-02	rs2952155	НарМар		IVSI +5154	c>t	0.2
ERBB2-03	rs1810132	НарМар		IVS4 +300	t>c	0.28
ERBB2-04	rs1801200	НарМар		EX17 (1655 V)	a>g	0.33
ERBB2-05	rs1058808	НарМар		EX27 (A1170P)	g>c	0.29
C17ORF37-01	rs4252665	НарМар	4	START -26	c>t	0.06
GRB7-01	rs8192704	НарМар		IVS2 +13	g>a	0.16
GRB7-02	rs11078921	dbSNP		STOP +5330	c>a	0.26
ZNFN1A3-01	rs907091	НарМар		START -361	c>t	0.47
ZNFN I A3-02	rs10445308	НарМар		IVS2 +4027	c>t	0.48

Sixteen SNPs were HapMap tSNPs, one was a HapMap singleton (ERBB2-04) and six were randomly selected from the dbSNP database.



Table 2 Genotype frequencies, minor allele frequencies (MAF) and P-values for 23 SNPs genotyped in 2275 women with breast cancer and 2280 controls

PPARBP-01 Cases 2190 0.20 1422 665 103 0.85 Controls 2774 1481 694 99 0.85 Controls 2774 1481 694 99 0.85 Cases 2181 0.19 1421 675 85 0.47 Cases 2186 0.14 1688 499 29 0.36 Cases 2186 0.14 1688 499 29 0.36 Cases 2186 0.14 1688 551 37 0.07 Cases 2186 0.1685 551 37 0.07 Cases 2180 0.10 1797 379 17 0.79 Controls 2275 1859 394 22 0.36 Cases 2180 0.31 1020 941 219 0.4 Cases 2180 0.31 1020 0.4 Cases 2180 0.5 Cases 2184 0.5 Cases 2185 0.5 Cases 218	SNP	Series	Number genotyped	MAF	Common homozygote	Heterozygote	Rare homozygote	P-value ^a
PRABRPQ2 Cases Controls 2181 bit Controls 0.19 bit 1516 bit 665 bit 92 bit 1516 bit 665 bit 92 bit 1516 bit 665 bit 37 bit 1516 bit 1516 bit 1518 bit 151	PPARBP-01	Cases	2190	0.20	1422	665	103	0.85
Controls Cares 2186 0.14 1658 499 29 0.36		Controls	2274		1481	694	99	
CRK7-01 Cases 2186 0.14 1658 499 29 0.36 Controls 2273 1685 551 37 7 77 0.79 179 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 177 0.79 1879 179 179 1879 179 179 179 179 179 179 179 179 179 1	PPARBP-02	Cases	2181	0.19	1421	675	85	0.47
Controls		Controls				665		
CRK7-02 Cases	CRK7-01	Cases		0.14				0.36
NEUROD Cases 2180		Controls				551		
NEUROD2-01 Cases 2180 0.31 1020 9.41 219 0.4 Controls 2271 1090 937 244 979 1150 271 1090 937 244 979 1150 2271 1090 937 244 979 1150 2245 1150 860 175 979 1150 860 175 979 1150 860 175 979 1150 860 175 979 1150 860 175 979 1150 860 175 979 1150 860 175 979 1150 860 175 979 1150 1750 1750 1750 1750 1750 1750 1750	CRK7-02	Cases		0.10				0.79
Controls		Controls	2275		1859	394	22	
PPPIRIB-01 Cases 2 50 0.27 1 52 833 165 0.95 PPPIRIB-02 Cases 2085 0.17 1435 585 65 0.94 STARD3-01 Cases 2 158 0.22 1303 736 119 0.96 Cases 2 158 0.22 1303 736 119 0.96 CAB2-01 Cases 2 184 0.32 1042 233 2 19 0.71 CAB2-01 Cases 1 192 0.31 930 801 192 0.84 CAB2-02 Cases 1 923 0.31 930 801 192 0.94 CAB2-03 Cases 1 923 0.33 995 953 231 0.85 CAB2-04 Cases 2 179 0.33 995 953 231 0.85 CAB2-05 Cases 2 108 0.34 961 969 255 0.78 CAB2-04 Cases <	NEUROD2-01	Cases		0.31		941	219	0.4
Controls		Controls	2271		1090	937	244	
PPFIRIB-02 Cases 2085 0.17 1435 585 65 0.94 STARD3-01 Cases 2158 0.22 1303 736 119 0.96 CAB2-01 Cases 2184 0.32 1042 2933 219 0.71 CAB2-01 Cases 1184 0.32 1042 2933 219 0.71 CAB2-02 Cases 1193 0.31 930 801 192 0.84 CAB2-03 Cases 2179 0.33 995 953 231 0.85 CAB2-04 Cases 2189 0.34 961 969 255 0.78 CAB2-05 Cases 2185 0.34 961 969 255 0.78 CAB2-05 Cases 2185 0.34 961 969 255 0.78 ERB2-01 Cases 2038 0.35 901 890 247 0.88 ERB2-05 Cases 2038<	PPPIRIB-01	Cases	2150	0.27	1152	833	165	0.95
STARD3-01 Cases 2198 1520 607 71 STARD3-01 Cases 2158 0.22 1303 736 119 0.96 CAB2-01 Cases 2184 0.32 1042 923 219 0.71 CAB2-02 Cases 11923 0.31 930 801 192 0.84 CAB2-02 Cases 11923 0.31 930 801 192 0.84 CAB2-03 Cases 11923 0.31 930 801 192 0.84 CAB2-04 Cases 11923 0.31 930 801 192 0.84 CAB2-05 Cases 2179 0.33 995 953 231 0.85 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-05 Cases 208 0.35 901 890 247 0.88 ERB2-05 Cases 2038 0.35 901 <td></td> <td>Controls</td> <td>2245</td> <td></td> <td>1210</td> <td>860</td> <td>175</td> <td></td>		Controls	2245		1210	860	175	
STARD3-01 Cases Controls 2158 0.22 1303 736 119 0.96 CAB2-01 Cases 2184 0.32 1042 923 219 0.71 CAB2-02 Cases 1923 0.31 930 801 192 0.84 CAB2-02 Cases 1923 0.31 930 801 192 0.84 CAB2-03 Cases 2179 0.33 995 953 231 0.85 CAB2-03 Cases 2185 0.34 961 969 255 0.78 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-05 Cases 2038 0.35 901 890 247 0.58 CAB2-06 Cases 2038 0.35 901 890 247 0.58 CAB2-07 Cases 2038 0.35 901 890 247 0.58 ERBB2-01 Cases 2	PPPIRIB-02	Cases	2085	0.17	1435	585	65	0.94
CAB2-01 Controls 2266 1377 767 122 CAB2-01 Cases 2184 0.32 1042 923 219 0.71 CAB2-02 Cases 1923 0.31 930 801 192 0.84 CAB2-03 Cases 2179 0.33 995 953 231 0.85 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-05 Cases 2038 0.35 901 890 247 0.58 ERBB2-01 Cases 2023 0.13 1548 433 42 0.14 ERBB2-02 Cases 2020 0.26 1162 738 140 0.45 ERBB2-03 Cases 2050 0.32 969 <td></td> <td>Controls</td> <td>2198</td> <td></td> <td>1520</td> <td>607</td> <td>71</td> <td></td>		Controls	2198		1520	607	71	
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CAB2-02 Cases 1923 0.31 930 801 192 0.84 CAB2-02 Cases 1923 0.31 930 801 192 0.84 CAB2-03 Cases 2179 0.33 995 953 231 0.85 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-04 Cases 2185 0.34 961 969 255 0.78 CAB2-05 Cases 2038 0.35 901 890 247 0.58 ERBB2-01 Cases 2023 0.13 1548 433 42 0.14 ERBB2-01 Cases 2023 0.13 1548 433 42 0.14 ERBB2-02 Cases 2040 0.26 1162 738 140 0.45 ERBB2-03 Cases 2050		Controls	2266		1377	767	122	
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CAB2-03 Cases 2179 0.33 995 953 231 0.85 CAB2-04 Cases 2179 0.33 995 953 231 0.85 CAB2-04 Cases 2185 0.34 1024 1014 236 CAB2-05 Cases 2185 0.34 961 969 255 0.78 CAB2-05 Cases 2038 0.35 901 890 247 0.82 CAB2-05 Cases 2038 0.35 901 890 247 0.88 CAB2-05 Cases 2023 0.13 1548 433 42 0.14 ERBB2-01 Cases 2023 0.13 1548 433 42 0.14 ERBB2-02 Cases 2040 0.26 1162 738 140 0.45 ERBB2-03 Cases 2050 0.32 969 861 220 0.69 ERBB2-04 Cases 1999 0.25 <td></td> <td>Controls</td> <td>2276</td> <td></td> <td>1064</td> <td>969</td> <td>243</td> <td></td>		Controls	2276		1064	969	243	
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	ZNFN1A3-07			0.48				0.99

^aTest for heterogeneity of genotype frequencies between cases and controls (2 df).

nor were there any differences between cases and controls for the individual haplotype frequencies.

DISCUSSION

We used a comprehensive SNP tagging approach using the publicly available HapMap data, complemented by the random selection of SNPs from dbSNP, to determine if common variation in the *ERBB2* amplicon is involved in breast cancer susceptibility. A total of 23 SNPs in 4 LD blocks were genotyped; we saw no association between any of the genotypes or associated haplotypes and risk of breast cancer.

We could have failed to observe a true association because of inadequate tagging, insufficient statistical power, or the effect of confounders. The selection of tagging SNPs is most reliable where the region of interest has been resequenced in a sample of individuals sufficiently large to identify all common variants. Such data were not available for this region, and it is possible that we have not adequately tagged an unidentified, disease-predisposing SNP. We estimated how well tSNPs would tag such unknown SNPs by performing a leave-one-out cross validation procedure on the HapMap data used for tSNP selection. Each of the 65 known SNPs were dropped in turn and tSNPs selected from the remaining SNPs within the block, thus simulating unidentified polymorphisms (Ahmadi et al, 2005). The ability of the tSNPs to tag the dropped SNP was then evaluated by calculating r_s^2 . The average r_s^2 for all dropped SNPs was 0.91 and 59 out of the 65 dropped SNPs (91%) were tagged with an $r_s^2 > 0.75$. This suggests that 91% of the unknown variation was well tagged. Furthermore, it has been shown that Phase 2 HapMap data provides a robust alternative to complete re-sequencing data with minimal loss of power (De Bakker et al, 2005). After we had completed the genotyping for this study, data for phase 2 of the HapMap project were released. For

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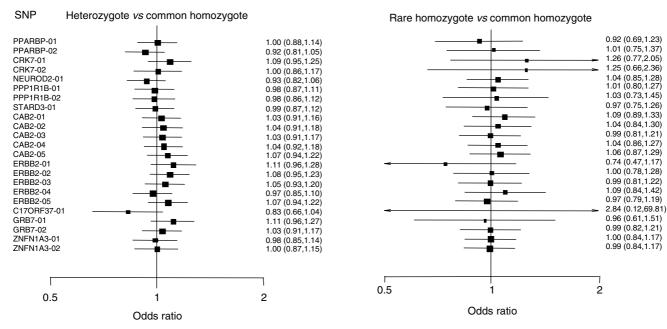


Figure 2 Genotype-specific risks for 23 SNPs genotyped in 2275 women with breast cancer and 2280 controls.

Table 3 Haplotype frequencies within LD blocks. ERBB2-04, a HapMap singleton SNP, was excluded from block 3 for haplotype analyses as it is poorly correlated with any other SNPs in the block

Block	Global test of association	Haplotype	Frequency in controls (%)	P-value
I	0.16	-a-t-t-a-	41	0.74
		-a-c-t-a-	19	0.43
		-g-t-t-a-	16	0.24
		-a-t-a-a-	13	0.13
		-a-t-t-g-	7	0.48
2	0.58	-t-g-g-c-	68	0.8
		-c-a-a-g-	17	0.96
		-c-a-g-c-	7	0.41
3	0.48	-c-a-a-a-a-c-c-t-g-	51	0.34
		-t-g-g-g-g-c-t-c-c-	24	0.24
		-c-a-a-a-a-a-c-t-g-	13	0.31
		-t-g-g-g-g-c-c-c-c-	6	0.57
4	0.44	-c-g-c-t-c-	37	0.55
		-c-g-a-c-t-	28	0.75
		-c-g-c-c-t-	13	0.27
		-c-a-c-t-c-	9	0.19

region A, there were now data for 35 SNPs – SNP density 1 SNP/ 4.2 kb. We used the programme TAGGER to test the performance of our selected tSNPs on HapMap phase 2 (Paul de Bakker, http://www.broad.mit.edu/mpg/tagger/). Mean pairwise r^2 ($r_{\rm p}^2$) was 0.92 with 31 HapMap SNPs being tagged with $r_{\rm p}^2 > 0.8$ and a minimum $r_{\rm p}^2$ of 0.21. For region B there were data on 57 SNPs (1 SNP/3.3 kb). Mean $r_{\rm p}^2$ was 0.91 with 53 HapMap SNPs being tagged with $r_{\rm p}^2 > 0.8$ and a minimum $r_{\rm p}^2$ of 0.12. These are conservative estimates since additional SNPs with undetermined tagging properties were randomly selected from dbSNP. Thus, the majority of common genetic variation is likely to have been captured, but we cannot exclude the possibility that an important common variant was missed.

PNMT is thought to regulate leptin levels and could be relevant to breast cancer biology via its effect on body weight (Harvey and Ashford, 2003; Stattin et al, 2004). None of the HapMap SNPs were in PNMT, raising questions regarding appropriate tagging of the gene. Ahmadi et al (2005) studied tagging patterns across 55 genes, including PNMT, that control the absorption, distribution, metabolism and excretion of drugs; they were able to determine that variation within PNMT was adequately tagged by rs1053651 in TCAP and rs903502 in CAB2, two HapMap SNPs that were well tagged by our set of tSNPs.

The statistical power of the study depends on the risk allele frequency, the risks conferred and the genetic mode of action (dominant, recessive, codominant). The staged approach substantially reduces genotyping costs without significantly affecting statistical power. For example, assuming that the causative SNP is tagged with $r_p^2 = 0.8$, a type I error rate of 0.0001 and genotyping success rate of 0.95, the staged study has 86% power to detect a dominant allele with a minor allele frequency (MAF) of 0.05 that confers a relative risk of 1.5 or 87% power to detect a dominant allele with a MAF of 0.25 that confers a relative risk of 1.3. Power to detect recessive alleles is less - 53% for an allele with a MAF of 0.25 and a relative risk of 1.5 and 71% for an allele with a MAF of 0.5 and a relative risk of 1.3. Such high power is illustrated by the narrow confidence intervals observed for odds ratios associated with genotypes: heterozygote odds ratios higher than 1.28 were excluded for all SNPs. Based on the upper confidence limits for all the risk estimates we can exclude SNPs that explain more than 0.8% of the excess familial risk of breast cancer. The possibility of variants with smaller effects on risk cannot be excluded. Similarly the possibility that there are rare variants with modest, or even large effect cannot be excluded.

Confounders are factors that are associated with both genotype and phenotype. They may bias results towards false positives and false negatives. In the context of breast cancer genetic association studies, it is difficult to envisage a true confounder. Most factors that are likely to be associated with both genotype and breast cancer will be intermediate factors, not confounders. The cases and controls used for these analyses were not matched for age (though broadly similar). However, there was no association of genotype with age in controls for any of the SNPs



studied and age-adjusted odds ratios were similar to the unadjusted values.

Some authors have advocated the use of histopathologic or demographic data that subclassify individuals in order to identify homogeneous subsets for analysis (Rebbeck *et al*, 2004). In the absence of any main effect or strong biological rationale, we have not carried out subgroup analyses. The number of possible *post hoc*, subgroup analyses is large and there is a strong possibility that one or more tests will be statistically significant simply by chance (Colhoun *et al*, 2003; Pharoah *et al*, 2005). Much larger sample sizes would be required to obtain reliable results. Nor did we test for SNP-SNP or SNP-environment interactions as the number of such interactions is very large – there are over 2000 possible two- and three-way interactions between the 23 genotyped common SNPs – and a clear strategy on how to best approach interactions has yet to be defined (Ritchie *et al*, 2003; Hunter, 2005).

We found no evidence that common genetic variation in the *ERBB2* amplicon is associated with an altered risk of breast cancer. The strategy of selecting candidate genes from regions that are often amplified was not successful here, but we have only evaluated a small proportion of the genome that is commonly somatically altered in breast cancer. The search for common susceptibility alleles for breast and the other common cancers using a candidate

gene approach has been notable for its lack of success. However, there are many, possibly thousands, of candidate genes of which only tens or hundreds have been comprehensively assessed for susceptibility. Advances in our understanding of human genomic architecture with rapid developments in high-throughput genotyping technology have made empirical, genome-wide association (GWA) studies feasible. The results of the first GWA scans in breast and colorectal cancer are expected in the near future. These should provide evidence whether or not common susceptibility variants exist. They may also provide an indication whether candidate gene studies remain a valid approach and, if so, what the likely candidates are.

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