

# Angiotensin II type I receptor expression in ovarian cancer and its correlation with tumour angiogenesis and patient survival

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Angiotensin II, a main effector peptide in the renin–angiotensin system, acts as a growth-promoting and angiogenic factor via type I angiotensin II receptors (AT<sub>1</sub>R). We have recently demonstrated that angiotensin II enhanced tumour cell invasion and vascular endothelial growth factor (VEGF) secretion via AT<sub>1</sub>R in ovarian cancer cell lines *in vitro*. The aim of the present study was to determine whether AT<sub>1</sub>R expression in ovarian cancer is correlated with clinicopathological parameters, angiogenic factors and patient survival. Immunohistochemical staining for AT<sub>1</sub>R, VEGF, CD34 and proliferating cell nuclear antigen (PCNA) were analysed in ovarian cancer tissues ( $n=67$ ). Intratumour microvessel density (MVD) was analysed by counting the CD34-positive endothelial cells. Type I angiotensin II receptors were expressed in 85% of the cases examined, of which 55% were strongly positive. Type I angiotensin II receptors expression was positively correlated with VEGF expression intensity and MVD, but not with histological subtype, grade, FIGO stage or PCNA labelling index. In patients who had positive staining for AT<sub>1</sub>R, the overall survival and progression-free survival were significantly poor ( $P=0.041$  and 0.017, respectively) as compared to those in patients who had negative staining for AT<sub>1</sub>R, although VEGF, but not AT<sub>1</sub>R, was an independent prognostic factor on multivariate analysis. These results demonstrated that AT<sub>1</sub>R correlated with tumour angiogenesis and poor patient outcome in ovarian cancer, suggesting its clinical potential for a novel molecular target in strategies for ovarian cancer treatment.

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Ovarian cancer is the leading cause of death from female genital malignancies despite significant advances in diagnosis and treatment (McGuire and Markman, 2003). Several clinicopathological parameters have been reported to be of prognostic significance in ovarian cancer, including disease stage, histological grade, histological subtype, residual tumour volume, presence of cytologically malignant ascites and response to chemotherapy (Omura *et al*, 1991; Chi *et al*, 2001). In addition to these established clinical parameters, the identification of biochemical or molecular markers more strictly related to intrinsic tumour cell behaviour in ovarian cancer and its characteristic progression pathway may be helpful in improving the survival of patients with this disease.

Angiotensin II, a multifunctional bioactive peptide in the renin–angiotensin system (RAS), plays a fundamental role in controlling cardiovascular and renal homeostasis. Angiotensin II also acts as a potent growth factor not only for vascular smooth muscle cells, but also for certain cancer cell lines (Fujimoto *et al*, 2001; Muscella *et al*, 2002). In addition, angiotensin II stimulates angiogenesis via the upregulation of vascular endothelial growth factor (VEGF) (Le Noble *et al*, 1993; Chua *et al*, 1998; Pupilli *et al*, 1999; Tamarat

*et al*, 2002). These cellular effects of angiotensin II are mostly mediated through specific G-protein-coupled AT<sub>1</sub>R.

Recent studies have proposed the concept of a localized tissue RAS in various organs (Nielsen *et al*, 1995), and activation of the RAS has been demonstrated under neoplastic conditions (Inwang *et al*, 1997; Takeda and Kondo, 2001; Juillerat-Jeanneret *et al*, 2004). In the tumour-related RAS, angiotensin II is abundantly generated from angiotensin I by angiotensin-converting enzyme (ACE), and AT<sub>1</sub>R expression is generally upregulated. Previous studies showed that the angiotensin II–AT<sub>1</sub>R system is deeply involved in tumour growth, metastasis and angiogenesis in experimental animal models, suggesting a therapeutic potential of RAS blockade using an ACE inhibitor or AT<sub>1</sub>R blocker (Rivera *et al*, 2001; Yoshiji *et al*, 2001; Fujita *et al*, 2002; Miyajima *et al*, 2002; Egami *et al*, 2003; Uemura *et al*, 2003; Arrieta *et al*, 2005).

In gynaecological malignancies, prior studies at our laboratory demonstrated that angiotensin II stimulated *in vitro* cell proliferation, invasion, or VEGF secretion via AT<sub>1</sub>R in cervical cancer (Kikkawa *et al*, 2004; Suganuma *et al*, 2004), endometrial cancer (Watanabe *et al*, 2003) and choriocarcinoma (Ino *et al*, 2003). Recently, we were the first to show that AT<sub>1</sub>R is expressed in human ovarian cancer cells and angiotensin II enhanced tumour cell invasion and VEGF expression/secretion via AT<sub>1</sub>R (Suganuma *et al*, 2005). Furthermore, we demonstrated that AT<sub>1</sub>R blocker suppresses angiogenesis and the peritoneal dissemination of ovarian cancer in a mouse model (Suganuma *et al*, 2005). These

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results prompted us to hypothesize that angiotensin II acts as an angiogenic and tumour-progressive factor for ovarian cancer, and that AT<sub>1</sub>R may have clinical potential as a novel molecular target or as a prognostic indicator in the treatment of ovarian cancer, as well as in other gynaecological malignancies.

Based on these findings, the present study examined the immunohistochemical expression of AT<sub>1</sub>R in ovarian cancer tissues to determine whether AT<sub>1</sub>R expression is correlated with clinicopathological factors or angiogenic parameters, including VEGF expression and intratumour microvessel density (MVD). Furthermore, we assessed whether AT<sub>1</sub>R correlates with the prognosis of ovarian cancer patients.

## MATERIALS AND METHODS

### Reagents and antibodies

Rabbit polyclonal antibody against AT<sub>1</sub>R (306) was purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Rabbit anti-human VEGF polyclonal antibody (A-20) was also purchased from Santa Cruz. Mouse monoclonal antibody against human CD34, a marker of endothelial cells, was obtained from Immuno- tech (Marseille, France). Antiproliferating cell nuclear antigen (PCNA) monoclonal antibody PC10 was purchased from Dako (Glostrup, Denmark).

### Patients and tissue samples

Human epithelial ovarian cancer tissues ( $n=67$ ) were obtained from patients who underwent surgical treatment at Nagoya University Hospital between 1993 and 2002. Surgical treatment consisted of total abdominal hysterectomy and bilateral salpingo-oophorectomy, followed by surgical staging and/or debulking surgery (if necessary). All tissue samples were fixed in 10% formalin, embedded in paraffin, and routinely stained with haematoxylin and eosin for histological examination. The histological cell types and histological grade (tumour differentiation) were assigned according to the criteria of the World Health Organization (WHO) classification. Clinical staging was reviewed based on the International Federation of Gynecology and Obstetrics (FIGO) staging system. The patients received post-operative chemotherapy with platinum plus cyclophosphamide and doxorubicin (before 1997), or platinum plus paclitaxel (after 1997) for high-risk early stage (stage I with grade 3; stage IC; any stage II) or advanced diseases (stages III and IV). Tumour recurrence/progression was defined based on clinical, radiological or histological diagnosis. The use of tissue samples was approved by the Institutional Review Board (IRB) of Nagoya University Graduate School of Medicine and by individual patients, respectively.

### Immunohistochemistry

Formalin-fixed, paraffin-embedded tissue sections were cut at a thickness of 4 µm. For heat-induced epitope retrieval, deparaffinized sections in 0.01 M citrate buffer (Target Retrieval Solution pH 6.1, Dako) were treated three times at 90°C for 5 min using a microwave oven. Immunohistochemical staining was performed using the avidin-biotin immunoperoxidase technique (Histofine SAB-PO kit, Nichirei, Tokyo, Japan). Endogenous peroxidase activity was blocked by incubation with 0.3% H<sub>2</sub>O<sub>2</sub> in methanol for 15 min, and nonspecific immunoglobulin binding was blocked by incubation with 10% normal goat serum for 10 min. Sections were incubated at room temperature for 2 h with primary antibody (anti-AT<sub>1</sub>R at 1:100 dilution, anti-VEGF at 1:200, anti-CD34 at 1:40, anti-PCNA at 1:40). The sections were rinsed and incubated for 30 min with biotinylated second antibody. After washing, the sections were incubated for 30 min with horseradish peroxidase-

conjugated streptavidin, and finally treated with 3-amino-9-ethylcarbazole (AEC) in 0.01% H<sub>2</sub>O<sub>2</sub> for 10 min. The slides were counterstained with Meyer's haematoxylin. As a negative control, the primary antibody was replaced with normal rabbit IgG or mouse IgG at an appropriate dilution. As positive immunohistochemical controls, marked immunoreactivity of AT<sub>1</sub>R and VEGF in placental trophoblastic tissues was confirmed as reported previously (Shiraishi *et al*, 1996; Ino *et al*, 2003). The immunostaining intensity for AT<sub>1</sub>R and VEGF was scored semiquantitatively based on the percent positivity of stained cells on a three-tiered scale as follows: -, negative (no positive cells); +, focally or weakly positive (<50% positive cells); ++, diffusely or strongly positive (>50% positive cells). In each case, at least three different areas were evaluated and the mean of the results was considered to be the expression intensity score. The staining procedure for each antibody was repeated twice, and it was confirmed that there was no difference in the staining intensity between the two experiments. The scoring procedure was carried out twice by two independent observers (each blinded to the other's score) without any knowledge of the clinical parameters or other prognostic factors. The concordance rate was over 95% between the observers. In the case of disagreement, the slides were reviewed simultaneously by these two observers, with another, different observer, who were seated together at a multiheaded microscope in order to resolve the difference of opinion.

### Evaluation of tumour angiogenesis and proliferation

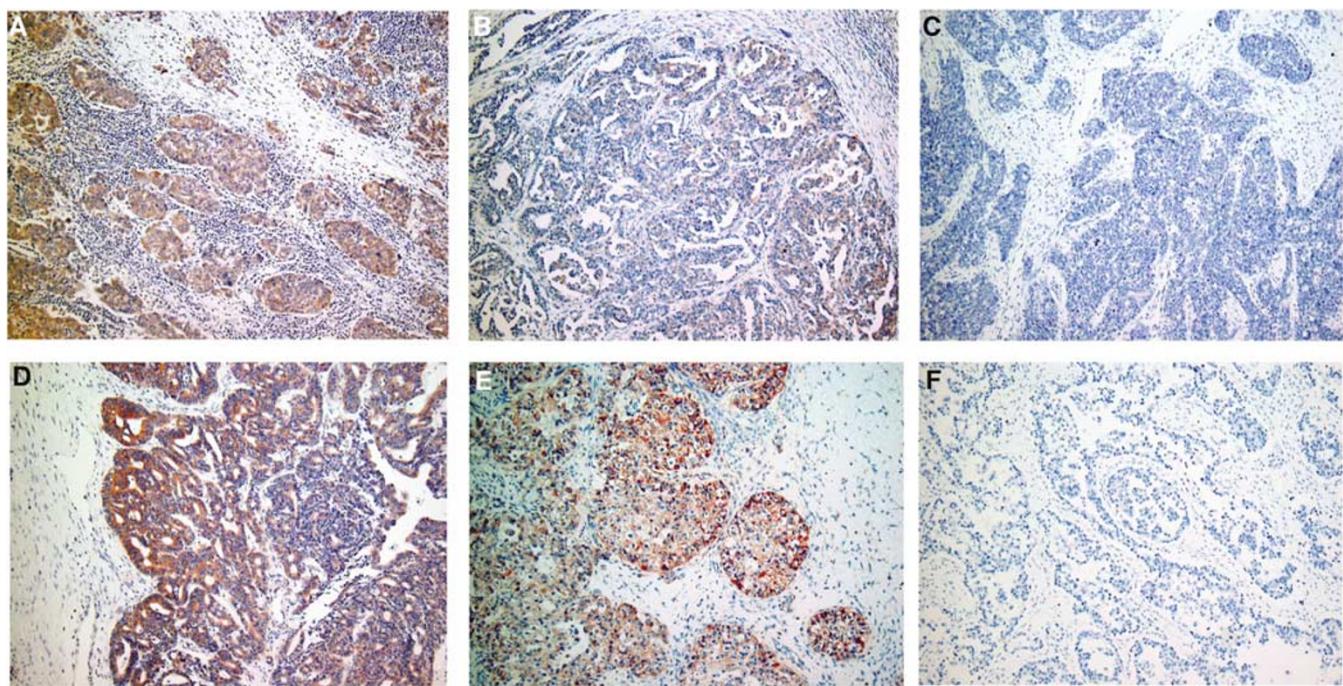
Tumour angiogenesis was assessed by counting the CD34-positive capillaries and small venules, according to the method of Weidner *et al* (1992). First, after scanning the immunostained section under a light microscope at low magnification ( $\times 40$  and  $\times 100$ ), the area within the tumour having the highest number of distinct CD34-staining microvessels ('hot spots') was selected. For the determination of intratumour MVD, all microvessels were counted within the neovascular hot spot under a light microscope at  $\times 200$  magnification ( $\times 20$  objective lens and  $\times 10$  ocular lens). Two investigators independently evaluated tumour vascularity without any information about the clinicopathological features or expression scores for AT<sub>1</sub>R and VEGF. The average number of microvessels of the three  $\times 200$  fields that were strictly confined to the hot spot area was recorded as the MVD, as described previously (Ueda *et al*, 1999).

To assess proliferation, sections were immunostained with the cellular proliferation marker PCNA as described previously (Thomas *et al*, 1995; Fujimura *et al*, 2000). The PCNA labelling index (PCNA LI) was defined as the number of tumour cells with nuclear PCNA immunostaining divided by the total number of tumour cells, and expressed as a percentage. A total of 1000 nuclei in the selected area were counted under a light microscope at high magnification ( $\times 400$  fields) and the mean percentages were recorded as the PCNA LI.

### Statistical analysis

Spearman's rank correlation test was performed to analyse the correlation between AT<sub>1</sub>R expression scores and various parameters. Fisher's exact test or chi-square ( $\chi^2$ ) test were also used to analyse the distribution of AT<sub>1</sub>R-strongly positive cases, according to clinicopathological, angiogenic and proliferative features.

Overall survival (OS) was calculated from the date of diagnosis to the date of death, and progression-free survival (PFS) was calculated from the date of diagnosis to the date of progression/recurrence or date of last follow-up. Survival analyses were performed according to the life tables method and according to the Kaplan-Meier method. Comparison of the survival between groups was performed with the log-rank test. Cox proportional-hazard analysis was used for univariate and multivariate analysis



**Figure 1** Representative immunohistochemical staining for AT<sub>1</sub>R in human ovarian cancer tissues. (**A**) Strongly positive; (**B**) weakly positive; (**C**) negative in serous adenocarcinoma; (**D**) strongly positive in endometrioid adenocarcinoma; (**E**) strongly positive in clear cell adenocarcinoma; (**F**) negative control. Original magnification,  $\times 100$  in (**A–F**).

to explore the effect of variables on survival. The SAS software (SAS Institute Inc., Cary, NC, USA) was used for all statistical analyses and a *P*-value of  $<0.05$  was considered significant.

## RESULTS

### Immunohistochemical expression of AT<sub>1</sub>R in ovarian cancer tissues

As shown in Figure 1, the immunoreactivity of AT<sub>1</sub>R was detected at variable levels, and was localized both on the membrane and in the cytoplasm of tumour cells. There was no immunoreactivity of AT<sub>1</sub>R in the tumour stroma. Of the 67 ovarian cancer specimens examined in this study, AT<sub>1</sub>R was detected in 57 (85%) cases, of which 37 (55%) were strongly positive. AT<sub>1</sub>R-negative tumours were found in only 10 cases (15%).

### Correlation of AT<sub>1</sub>R expression with clinicopathological, angiogenic and proliferative parameters

The correlation of AT<sub>1</sub>R expression with clinicopathological, angiogenic and proliferative parameters was analysed in 67 ovarian cancer tissues. Type 1 angiotensin II receptors expression did not significantly correlate with the histological subtype, FIGO stage or histological grade (tumour differentiation) (Figure 2A–C). In contrast, the AT<sub>1</sub>R expression positively correlated with the VEGF staining scores (Figure 2D). Of the VEGF-strongly positive cases ( $n=24$ ), 20 (83%) were AT<sub>1</sub>R strongly positive, while half ( $n=9$ ) of the VEGF-negative cases ( $n=18$ ) were AT<sub>1</sub>R negative (Figure 2D).

To determine the correlation between AT<sub>1</sub>R expression and tumour angiogenesis, we assessed the intratumour MVD by counting CD34-positive microvessels in the same series of ovarian cancer tissues ( $n=67$ ). Microvessel density ranged broadly from 20 to 150 (mean = 75.2, median = 68). Thus, we defined cases with

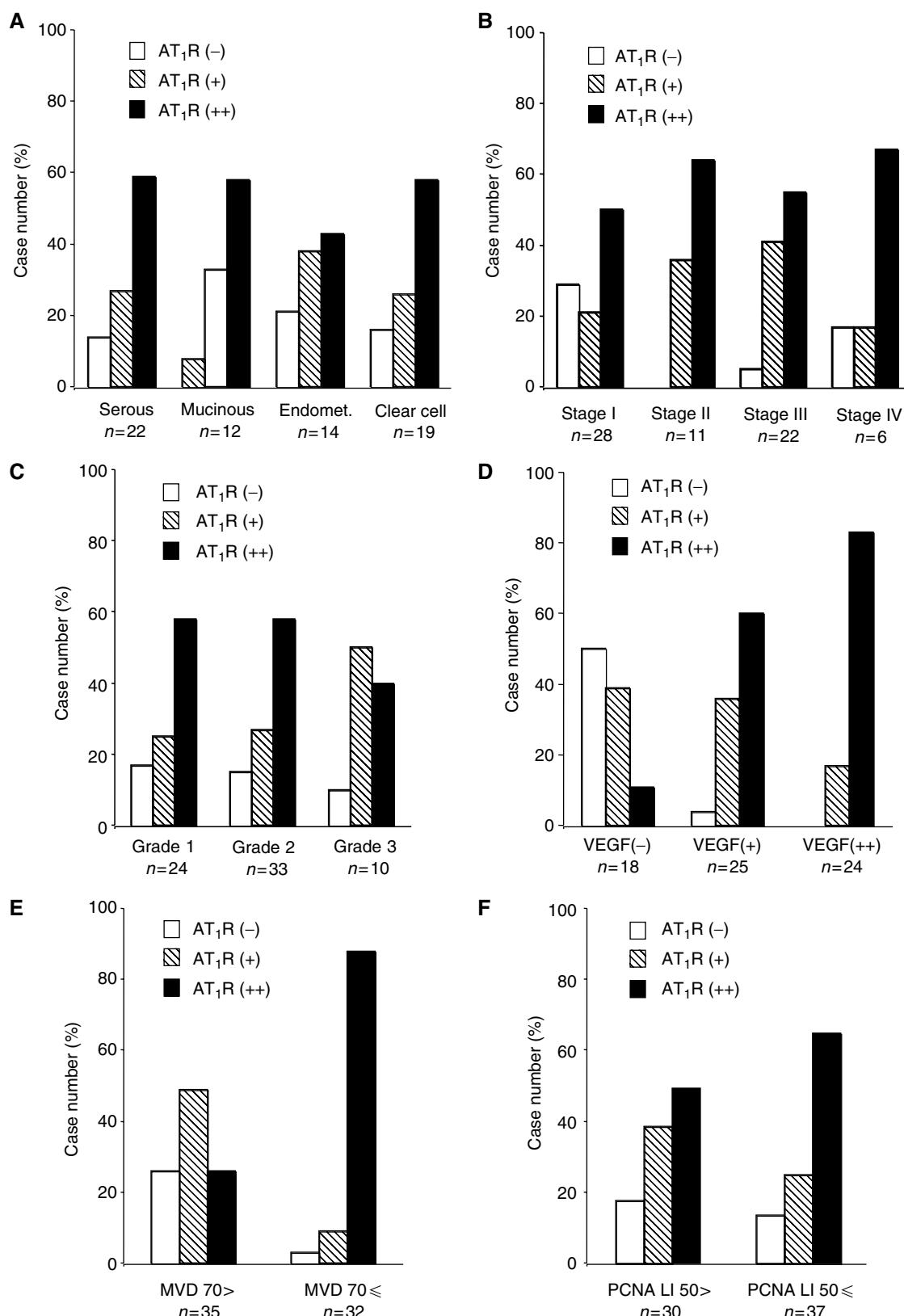
MVD number of more than 70 as high MVD ( $n=32$ ), while cases with MVD number of  $<70$  were defined as low MVD ( $n=35$ ). Interestingly, the AT<sub>1</sub>R expression score positively correlated with MVD, and 28 (87.5%) of 32 high MVD cases were AT<sub>1</sub>R strongly positive (Figure 2E). In this study, we confirmed that the MVD number did not correlate with the histological subtype, FIGO stage or histological grade, while VEGF expression scores positively correlated with the MVD number (data not shown). Taken together, our findings indicate that AT<sub>1</sub>R, as well as VEGF, are associated with tumour angiogenesis of ovarian carcinoma.

Next, we assessed the correlation between AT<sub>1</sub>R expression and tumour proliferation using immunohistochemical staining with PCNA in the same series of ovarian cancer tissues. The PCNA LI ranged from 16 to 80%, with a mean value of 50.16%, which is well consistent with the results from a previous report (Thomas *et al*, 1995). We defined cases with a PCNA LI of more than 50 as high PCNA LI ( $n=37$ ), while cases with a PCNA LI of  $<50$  were defined as low PCNA LI ( $n=30$ ). As shown in Figure 2F, the AT<sub>1</sub>R expression score did not significantly correlate with PCNA LI.

The results from correlation analyses of AT<sub>1</sub>R overexpression (strongly positive cases) with clinicopathological parameters, VEGF expression, MVD and PCNA are summarized in Table 1. These analyses demonstrated that AT<sub>1</sub>R overexpression was significantly correlated with VEGF overexpression and high MVD number, but not with the proliferation marker PCNA, nor any of the clinicopathological factors examined in this study.

### Correlation of AT<sub>1</sub>R expression with survival of ovarian cancer patients

Follow-up data were available for 58 patients (nine patients were lost to follow-up). The median follow-up was 60 months (range 1–121 months). During the follow-up period, the total number of cases in which death and progression/recurrence were observed was 19 (32.8%) and 25 (43.1%), respectively.



**Figure 2** Correlation of AT<sub>1</sub>R expression intensity with histological subtype (**A**), FIGO stage (**B**), histological grade (**C**), VEGF expression intensity (**D**), MVD number (**E**), and PCNA LI (**F**) in ovarian cancer tissues. Significant correlation ( $P<0.05$ ) was observed between AT<sub>1</sub>R expression and VEGF expression (**D**), or between AT<sub>1</sub>R expression and MVD number (**E**), while there was no significant correlation between AT<sub>1</sub>R expression and other clinicopathological factors or PCNA LI.

**Table 1** Correlation of AT<sub>1</sub>R overexpression with clinicopathologic factors, and angiogenesis/proliferation markers in ovarian cancer

	Patients		AT <sub>1</sub> R overexpression		
	No.	%	No.	%	P-value
All cases	67	100.0	37	55.2	
Histological type					
Serous	22	32.8	13	59.1	0.777 <sup>a</sup>
Mucinous	12	17.9	7	58.3	
Endometrioid	14	20.9	6	42.9	
Clear cell	19	28.4	11	57.9	
Histological grade					
G1	24	35.8	14	58.3	0.576 <sup>a</sup>
G2	33	49.3	19	57.6	
G3	10	14.9	4	40.0	
FIGO stage					
I	28	41.8	14	50.0	0.814 <sup>a</sup>
II	11	16.4	7	63.6	
III	22	32.8	12	54.5	
IV	6	9.0	4	66.7	
VEGF expression					
(-)	18	26.9	2	11.1	0.001 <sup>b</sup>
(+)	49	73.1	35	71.4	
Microvessel density					
70>	35	52.2	9	25.7	0.001 <sup>b</sup>
70≤	32	47.8	28	75.7	
PCNA LI					
50>	30	44.8	14	46.7	0.205 <sup>b</sup>
50≤	37	55.2	23	52.2	

<sup>a</sup>χ<sup>2</sup> test. <sup>b</sup>Fisher's exact test.

To evaluate the impact of AT<sub>1</sub>R expression on patient prognosis, overall survival (OS) and PFS curves were constructed using the Kaplan–Meier method. The 5-year OS rates of patients who had negative ( $n=10$ ), weakly positive ( $n=18$ ), and strongly positive ( $n=30$ ) expression for AT<sub>1</sub>R were 100, 45.8 and 55.7%, respectively (Figure 3A). The 5-year PFS rates of patients who had negative, weakly positive, and strongly positive expression for AT<sub>1</sub>R were 100, 27.5 and 41.9%, respectively (Figure 3B). Both OS and PFS in patients with positive (weak and strong) expression for AT<sub>1</sub>R were significantly lower than those in patients with negative AT<sub>1</sub>R expression ( $P=0.041$  and 0.017, respectively, by log-rank test). However, there was no significant difference in the distributions of OS and PFS between the AT<sub>1</sub>R-weakly positive and AT<sub>1</sub>R-strongly positive groups.

The correlation of VEGF expression, intratumour MVD, and PCNA LI with prognosis was also analysed. The 5-year OS rates of patients who had negative ( $n=15$ ), weakly positive ( $n=22$ ) and strongly positive ( $n=21$ ) expression for VEGF were 92.9, 48.4 and 51.3%, respectively (Figure 3C). The 5-year PFS rates of patients who had negative, weakly positive and strongly positive expression for VEGF were 69.6, 47.6 and 24.2%, respectively (Figure 3D). Both OS and PFS in patients with positive expression for VEGF were significantly lower than those in patients with negative VEGF expression ( $P=0.046$  and 0.036, respectively, by log-rank test), although there was no significant difference in the distributions of OS and PFS between the VEGF-weakly positive and strongly positive groups. In contrast to AT<sub>1</sub>R and VEGF, there was no significant difference in the distributions of OS and PFS according to the MVD number ( $P=0.203$  and 0.467, respectively), or

according to the PCNA LI ( $P=0.909$  and 0.316, respectively), as shown in Figure 3E–H. These results indicate that AT<sub>1</sub>R and VEGF, but not the MVD number and PCNA LI, significantly correlated with the impaired survival of ovarian cancer patients.

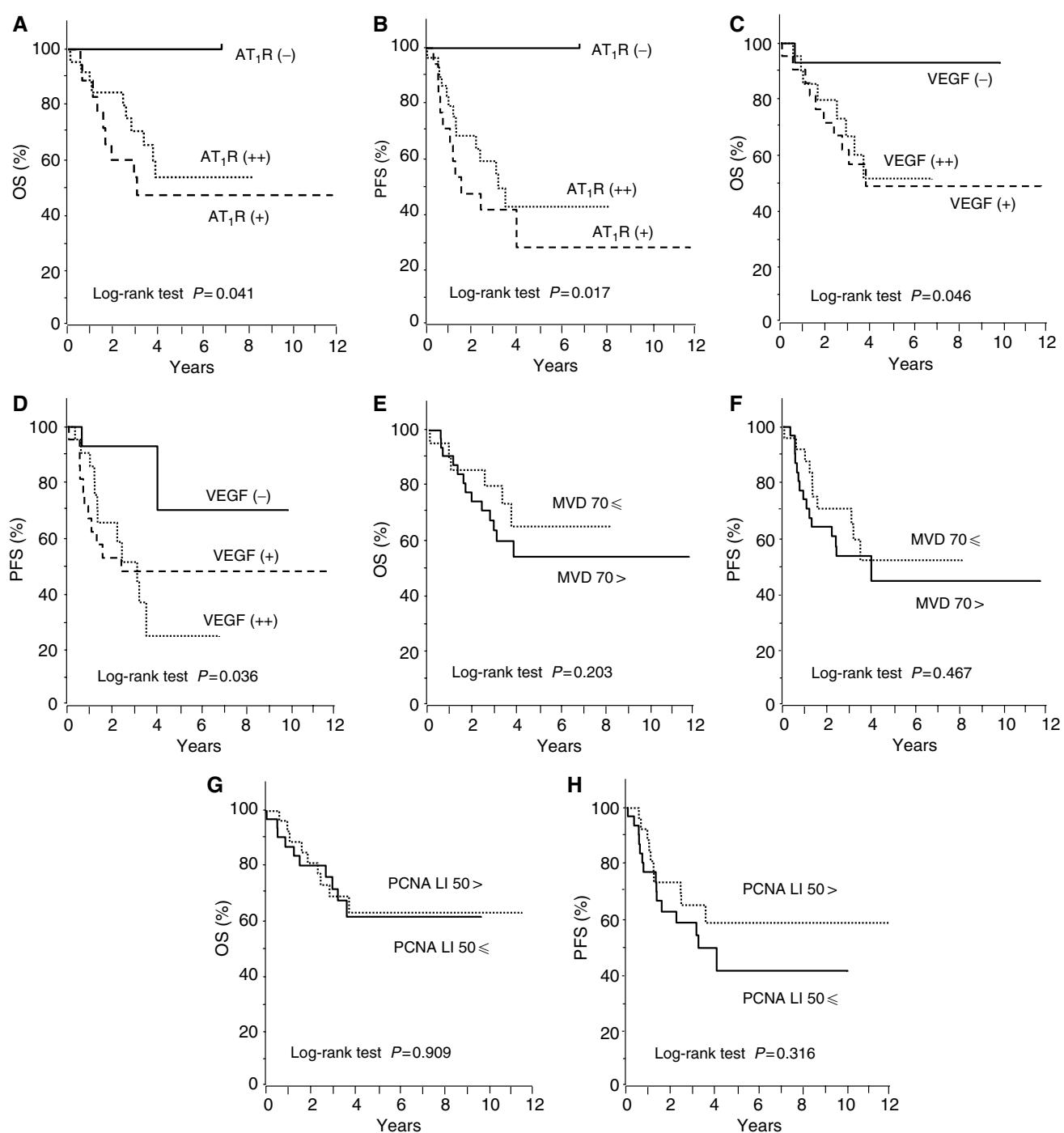
### Multivariate analysis of prognostic variables in ovarian cancer patients

Cox proportional-hazard analysis was performed to determine the impact of various factors on survival. The results of univariate/multivariate analyses of the variables, including AT<sub>1</sub>R, VEGF, MVD and PCNA LI, with respect to OS and PFS are shown in Tables 2 and 3, respectively. Because none of the patients with negative AT<sub>1</sub>R expression ( $n=10$ ) died or recurred (no risk for OS and PFS), the univariate analysis for AT<sub>1</sub>R expression (negative or weakly/strongly positive) could not be performed, and it was not entered into the multivariate analysis model. Among the other six variables, the FIGO stage ( $P=0.025$ ) and VEGF expression ( $P=0.018$ ) were statistically significant prognostic factors with respect to OS on multivariate analysis (Table 2). Similarly, the FIGO stage ( $P=0.015$ ) and VEGF expression ( $P=0.008$ ) were found to be independent prognostic factors with respect to PFS on multivariate analysis (Table 3).

### DISCUSSION

In the present study, we demonstrated the expression of the type 1 angiotensin II receptor, AT<sub>1</sub>R, in human ovarian carcinoma tissues, and its significant correlation with tumour angiogenesis and patient survival. It has been reported that AT<sub>1</sub>R is expressed in various human malignant tumour tissues, including breast cancer (Inwang *et al*, 1997), skin cancer (Takeda and Kondo, 2001), pancreatic cancer (Fujimoto *et al*, 2001), laryngeal carcinoma (Marsigliante *et al*, 1996) and prostate cancer (Uemura *et al*, 2003). Furthermore, we recently demonstrated that AT<sub>1</sub>R was expressed in gynaecological malignancies, including cervical cancer (Kikkawa *et al*, 2004), endometrial cancer (Watanabe *et al*, 2003), choriocarcinoma (Ino *et al*, 2003), and ovarian cancer (Suganuma *et al*, 2005). These findings suggest that AT<sub>1</sub>R exists in a wide spectrum of human cancers, especially in gynaecological malignancies, and that the angiotensin II-AT<sub>1</sub>R system may play significant roles in the localized RAS within these tumour tissues. Evidence for the involvement of AT<sub>1</sub>R in tumour progression, such as growth, metastasis and angiogenesis, has accumulated in various animal models (Rivera *et al*, 2001; Fujita *et al*, 2002; Miyajima *et al*, 2002; Egami *et al*, 2003; Uemura *et al*, 2003; Arietta *et al*, 2005). Until now, however, there have been no reports analysing the correlation of AT<sub>1</sub>R expression with clinical parameters, especially with patient prognosis, using a large scale of clinical samples of human cancers. Thus, the present study is the first to investigate the correlation between AT<sub>1</sub>R and clinical outcome in ovarian cancer patients.

Our immunohistochemical analysis showed that AT<sub>1</sub>R was present in 85% of ovarian carcinomas examined, and it was overexpressed in more than half (55%) of the cases. Furthermore, AT<sub>1</sub>R expression was not dependent on the histological subtype, grade or FIGO stage, although the rate of AT<sub>1</sub>R-negative cases was relatively high in FIGO stage I (eight of 28 cases: 29%) as compared to those in FIGO II–IV. Our prior studies showed that AT<sub>1</sub>R expression was almost absent in benign ovarian cystadenoma, but was dramatically upregulated with progression from borderline malignancy to invasive ovarian carcinoma (Suganuma *et al*, 2005). These results suggest the possible involvement of AT<sub>1</sub>R in the specific cell behaviours that were common to the malignant phenotypes, such as tumour cell invasion or aggressive neovascularization, rather than in the degree of tumour differentiation or histopathological subtypes.



**Figure 3** Overall survival (OS) and progression-free survival (PFS) curves drawn using the Kaplan–Meier method according to the AT<sub>1</sub>R expression (**A** and **B**), VEGF expression (**C** and **D**), MVD number (**E** and **F**), and PCNA LI (**G** and **H**). Significant differences in the OS and PFS according to the AT<sub>1</sub>R expression ( $P=0.041$  and  $0.017$ , respectively), and according to the VEGF expression ( $P=0.046$  and  $0.036$ , respectively). No significant difference in the OS and PFS according to the MVD ( $P=0.203$  and  $0.467$ , respectively) and according to the PCNA LI ( $P=0.909$  and  $0.316$ , respectively).

It is of particular interest that AT<sub>1</sub>R expression was positively correlated with VEGF expression intensity in the ovarian carcinomas examined in this study. Vascular endothelial growth factor is known to be the main angiogenic factor in ovarian cancer (Yamamoto *et al*, 1997; Fujimoto *et al*, 1998; Bamberger and Perrett, 2002). It has been shown that angiotensin II upregulated VEGF expression via AT<sub>1</sub>R (Chua *et al*, 1998; Pupilli *et al*, 1999; Tamarat *et al*, 2002), and this angiotensin II-induced VEGF

upregulation was mediated by hypoxia-inducible factor-1, even in non-hypoxic conditions (Richard *et al*, 2001). We have recently reported that angiotensin II stimulated VEGF expression and secretion in AT<sub>1</sub>R-positive ovarian cancer cells *in vitro* (Suganuma *et al*, 2005). Furthermore, we found that ACE, an enzyme producing angiotensin II in the RAS, was expressed in the tumour stroma of ovarian cancer (data not shown). Thus, the positive correlation between AT<sub>1</sub>R and VEGF observed in the present study

**Table 2** Univariate and multivariate analyses of overall survival (OS) in ovarian cancer patients

Variables	Categories	Univariate analysis			Multivariate analysis		
		Hazard ratio	95% CI	P-value	Hazard ratio	95% CI	P-value
FIGO stage	I/II III/IV	2.60	1.00–6.72	0.049	2.96	1.15–4.61	0.025
Histological grade	I or 2 3	0.19	0.02–1.56	0.122	0.22	0.03–1.71	0.147
AT <sub>1</sub> R expression	(–) (+)/++)	ND <sup>a</sup>	ND <sup>a</sup>	ND <sup>a</sup>	—	—	—
AT <sub>1</sub> R overexpression	(–)/(+) ++)	0.86	0.25–2.98	0.815	—	—	—
VEGF expression	(–) (+)/++)	5.75	0.71–46.52	0.101	12.18	1.54–96.37	0.018
MVD	<70 ≥70	0.27	0.09–0.77	0.145	0.30	0.11–0.84	0.122
PCNA LI	<50 ≥50	1.13	0.44–2.88	0.805	—	—	—

CI = confidence interval. <sup>a</sup>ND = not determined because no patients with negative AT<sub>1</sub>R expression died or recurred.

**Table 3** Univariate and multivariate analyses of progression-free survival (PFS) in ovarian cancer patients

Variables	Categories	Univariate analysis			Multivariate analysis		
		Hazard ratio	95% CI	P-value	Hazard ratio	95% CI	P-value
FIGO stage	I/II III/IV	2.29	1.02–5.14	0.044	2.70	1.21–6.04	0.015
Histological grade	I or 2 3	0.58	0.16–2.08	0.400	—	—	—
AT <sub>1</sub> R expression	(–) (+)/++)	ND <sup>a</sup>	ND <sup>a</sup>	ND <sup>a</sup>	—	—	—
AT <sub>1</sub> R overexpression	(–)/(+) ++)	0.85	0.26–2.75	0.789	—	—	—
VEGF expression	(–) (+)/++)	4.10	0.88–19.02	0.072	7.49	1.68–33.32	0.008
MVD	<70 ≥70	0.36	0.15–0.88	0.125	0.46	0.20–1.06	0.096
PCNA LI	<50 ≥50	1.49	0.65–3.42	0.343	—	—	—

CI = confidence interval. <sup>a</sup>ND = not determined because no patients with negative AT<sub>1</sub>R expression died or recurred.

not only reflects the results from the prior *in vitro* studies, but also suggests the existence of the angiotensin II-AT<sub>1</sub>R-VEGF system controlling angiogenic signals in ovarian cancer.

In addition to VEGF, the intratumour MVD number also positively correlated with AT<sub>1</sub>R expression intensity. Indeed, AT<sub>1</sub>R was overexpressed in 87.5% of the cases with high MVD numbers (more than 70). It is well known that MVD is a most reliable tool for reflecting tumour angiogenesis (Weidner *et al*, 1992) and it has been reported that VEGF expression directly correlated with increased MVD in a variety of tumours (Toi *et al*, 1996; Maeda *et al*, 1996). In the present study, we found the positive correlation of MVD number not only with VEGF, but also with AT<sub>1</sub>R. These

findings support our hypothesis that AT<sub>1</sub>R is a key molecule in tumour angiogenesis in ovarian cancer.

It has been reported that AT<sub>1</sub>R is involved in the proliferation of various cancers *in vitro* and *in vivo* (Fujimoto *et al*, 2001; Rivera *et al*, 2001; Muscella *et al*, 2002; Ino *et al*, 2003; Uemura *et al*, 2003). However, our immunohistochemical study showed that the AT<sub>1</sub>R expression score did not significantly correlate with the positivity of the cellular proliferation marker PCNA in ovarian cancer tissues. Consistently, we previously reported that angiotensin II enhanced the invasive activity and VEGF secretion, but not cell proliferation, in AT<sub>1</sub>R-positive ovarian cancer cell lines *in vitro* (Suganuma *et al*, 2005). Taken together, it appears that

AT<sub>1</sub>R is not directly involved in tumour cell proliferation in ovarian cancer, and that the angiotensin II-AT<sub>1</sub>R system plays differential roles in cancer progression, which may be dependent upon tumour type.

Our results from survival analyses demonstrated that both OS and PFS were significantly poorer in AT<sub>1</sub>R-positive cases than those in AT<sub>1</sub>R-negative cases. This provides the first evidence for the positive correlation of AT<sub>1</sub>R expression with poor clinical outcome in human malignancies. Surprisingly, we observed 100% of the 5-year OS and PFS in AT<sub>1</sub>R-negative cases, although the case number of these AT<sub>1</sub>R-non-expressing ovarian carcinomas was small ( $n=10$ ) in this study. In addition, we showed that there was no significant difference in the survival between AT<sub>1</sub>R strongly positive and weakly positive within the AT<sub>1</sub>R-expressing cases. These results might be unexpected because the immunohistochemical analysis showed a clear correlation between overexpression (strongly positive) of AT<sub>1</sub>R with high VEGF expression and high MVD numbers. One could speculate that the existence (either high or low expression) of AT<sub>1</sub>R plays a crucial role in the initiation of angiogenic signals in the primary lesions of ovarian cancer, while other various molecules are complexly involved in subsequent tumour progression, such as peritoneal dissemination or distal metastasis. Similar to AT<sub>1</sub>R, both the OS and PFS in VEGF-positive cases were significantly poorer than those in VEGF-negative cases, which is consistent with the observation in previous studies (Yamamoto *et al*, 1997; Shen *et al*, 2000). These findings suggest that AT<sub>1</sub>R, as well as VEGF, may become a prognostic marker for ovarian cancer patients; however, our multivariate analyses showed that FIGO stage and VEGF, but not AT<sub>1</sub>R, were independent prognostic factors for both OS and PFS. These results may be due, at least in part, to the small number of patient samples, because the AT<sub>1</sub>R-negative group consisted of only 10 patients, and there was no death or recurrence in this group. Further studies, including multivariate analyses using a larger sample size, are required to clarify whether AT<sub>1</sub>R can be an independent prognostic factor or not.

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