www.bjcancer.com

Low expression of polypeptide GalNAc *N*-acetylgalactosaminyl transferase-3 in lung adenocarcinoma: impact on poor prognosis and early recurrence

C Gu¹, T Oyama^{*,2}, T Osaki², J Li², M Takenoyama², H Izumi³, K Sugio², K Kohno³ and K Yasumoto²

¹Department of Thoracic and Cardiovascular Surgery, First Affiliated Hospital of Dalian Medical University, Dalian 116011, China; ²Department of Surgery II University of Occupational and Environmental Health, School of Medicine, 1-1 Iseigoaka, Yahatanishi-ku Kitakyushu 807-8555, Japan; ³Department of Molecular Biology, University of Occupational and Environmental Health, School of Medicine, 1-1 Iseigoaka, Yahatanishi-ku Kitakyushu 807-8555, Japan

Initial glycosylation of mucin-type O-linked protein is catalysed by one of the UDP-GalNAc: polypeptide *N*-acetyl-galactosaminyl transferase-3 (GalNAc-T3). O-glycosylation is important in the binding of cell adhesion molecules, cell differentiation, invasion, and metastasis in tumours. This study was designed to detect GalNAc-T3 expression in lung adenocarcinoma by using immunohistochemical staining, and to evaluate the relationship between the GalNAc-T3 expression level and prognosis and recurrence in completely resected lung adenocarcinoma patients. A low expression of GalNAc-T3 was detected in the cytoplasm of tumour cells in 79 of 148 patients (53.4%) with lung adenocarcinoma. The low expression of GalNAc-T3 was associated with poorly differentiated tumour (P < 0.0001), poor pathologic stage (P < 0.0001), lymph node metastasis (P < 0.0001), and tumour recurrence (P = 0.016). The lung carcinoma patients with low GalNAc-T3 expression had a poorer prognosis than those with high GalNAc-T3 expression of GalNAc-T3 was a significant independent factor for predicting poor prognosis and early recurrence (P = 0.006, rr = 2.87 and P = 0.019, rr = 3.05, respectively). The low expression of GalNAc-T3 may be a useful marker for predicting poor prognosis and early recurrence in completely resected lung carcinoma patients, particularly patients with stage I diseases. *British Journal of Cancer* (2004) **90**, 436–442. doi:10.1038/sj.bjc.6601531 www.bjcancer.com

Keywords: GalNAc-T3; lung adenocarcinoma; prognosis; recurrence

It is widely accepted that changes in the structure and distribution of cell surface glycoproteins are assumed to influence the biologic behaviour of tumour cells during malignant transformation and tumour progression (Nakamori et al, 1994). In the past few years, O-linked carbohydrate antigens, such as mucin antigen (MUC1), carcinoembryonic antigen (CEA), sialyl LewisX (SLX), CA19-9, Sialyl Tn (STn), and Tn have been reported to be associated with altered adhesion (Fogel et al, 1983; Rice and Bevilacqua, 1989), invasion (Bolscher et al, 1988; Matsushita et al, 1991; Nakamori et al, 1994), recurrence, and prognosis (Itzkowitz et al, 1990; Niklinski et al, 1992; Ogawa et al, 1994; Cao et al, 1995; Diez et al., 1996; Ohgami et al, 1999) in lung cancer and other tumours. Mucin-type O-linked carbohydrates may constitute up to 80% of the total mass of these glycoproteins (Clausen and Bennett, 1996); O-glycosylation has been shown to be important in the binding of cell adhesion molecules, cell differentiation, invasion, and

metastasis in tumours (Nakamori *et al*, 1994; Clausen and Bennett, 1996; Sutherlin *et al*, 1997; Nomoto *et al*, 1999). Initial glycosylation of mucin-type O-linked protein was catalysed by a family of the UDP-GalNAc: polypeptide N-acetyl-galactosaminyl transferases (GalNAc-transferase) (Hagen *et al*, 1993; Homa *et al*, 1993; Wandall *et al*, 1997). To date, seven distinct human GalNActransferase genes (from GalNAc-T1 to GalNAc-T7) have been cloned and characterised, and O-glycosylation is carried out in part by the differential expression of GalNAc-transferase in normal tissues and tumours (White *et al*, 1995; Bennett *et al*, 1996, 1998, 1999a, b; Wandall *et al*, 1997; Ten Hagen *et al*, 1998). Thus, the differential expression of O-linked carbohydrate antigens in tumours may be explained by the differential expression and activity of specific GalNAc-transferases.

GalNAc-T3 expression is restricted to cell lines derived from epithelial gland adenocarcinoma, but not cells derived from nonsecretory epithelial tissue carcinomas (Sutherlin *et al*, 1997; Nomoto *et al*, 1999). A recent study using a polyclonal antibody to GalNAc-T3 demonstrated that GalNAc-T3 is expressed in adenocarcinoma but not nonadenocarcinoma cell lines (Nomoto *et al.*, 1999). Previous immunohistochemical results suggest that the anti-GalNAc-T3 antibody may be useful for diagnostic purposes in the early stages of breast cancer, but the relationship between

^{*}Correspondence: T Oyama; E-mail: oyama@med.uoeh-u.ac.jp Supported in part by grant-in-aid for cancer research (No. 214571292) from the Ministry of Education, Science, Sports and Culture, Japan. Received 26 February 2003; revised 12 August 2003; accepted 5 November 2003

This study was a retrospective cohort and designed to detect GalNAc-T3 expression in lung adenocarcinoma by using immunohistochemical staining, and to evaluate the relationship between GalNAc-T3 expression levels and prognosis or recurrence of the patients' tumours.

MATERIALS AND METHODS

Patients and follow-up

We examined 148 of 184 (80.4%) consecutive patients with lung adenocarcinoma, who underwent complete surgical resection between July 1991 and September 1996 at the University of Occupational and Environmental Health, School of Medicine, Kitakyushu, Japan. The criteria for inclusion into the study were based on the availability of follow-up data. Clinicopathological data were obtained by retrospective chart review. Tumour stage was classified according to Revisions in the International System for Staging Lung Cancer (1997) (Mountain, 1997). There were 88 men and 60 women in this series, with a mean age of 64.8 years (range, 32–84). None of these patients received chemotherapy or radiotherapy prior to the operation. Of 148 (6.8%) patients, 10 received adjuvant chemotherapy and this adjuvant chemotherapy does not affect the prognosis of these patients.

For the postoperative follow-up, the patients were examined every month within the first year and at approximately 2- to 4month intervals thereafter. The evaluations included physical examination, chest roentgenography, analysis of blood chemistry, and carcinoembryonic antigen assay. Chest, abdominal, and brain computed tomographic scans and a bone scintiscan were performed every 6 months through the third year, and annually thereafter. If any symptoms or signs of recurrence appeared in these examinations, additional evaluations to locate the site of the recurrence were performed. Survival data were updated in April 2001. Follow-up was available for all patients, ranging from 53 to 3507 days after the primary operation (median follow-up, 48.5 months).

Western blot analysis

Cytoplasmic proteins were extracted from the frozen normal tissue and tumour tissue in adenocarcinoma patients and 5 μ g cytoplasmic proteins was electroblotted onto polyvinylidene difluoride membranes (Immobilon; Millipore, Bedford, MA, USA) after separation on 10% SDS-polyacrylamide gel electrophoresis gel. Polyclonal antibody against GalNAc-T3 was generated by multiple immunisations of a New Zealand white rabbit, using synthetic peptides as described previously (Nomoto *et al*, 1999). Immunoblot analysis was performed with a 1:5000 dilution of anti-GalNAc-T3 antibody. Detection was performed using ECL (Amersham Pharmacia Biotech, Buckinghamshire, UK). High GalNAc-T3 expression breast carcinoma given by Nomoto *et al* (1999) was used as a positive control.

Immunohistochemical staining

A formalin-fixed, paraffin-embedded, $3-\mu m$ section was obtained from each of the 148 samples of primary lesions. All specimens were stained with haematoxylin and eosin for histopathologic diagnosis. Immunohistochemical (IHC) staining was performed by a streptavidin – biotin – peroxidase complex method (Nomoto *et al*, 1999). Sections were immersed briefly in a citrate buffer (0.01 mol li⁻¹ citric acid: pH 6.0) and incubated for two 5-min intervals at 100°C in a microwave oven for antigen retrieval. They were then incubated with polyclonal GalNAc-T3 antibody diluted 1:2000 for 90 min at room temperature by using the Labeled 427

Immunostaining evaluation

All slides were evaluated for immunostaining by three observers (CG, JL, and TO) using a blind protocol design (observers had no information on clinical outcome or other clinicopathologic data). Cells were judged positive for GalNAc-T3 when the cytoplasm or cell membranes were stained. The percentage of positive cells was calculated by counting more than 1000 cells in random high-power fields (10×40), and scored according to the percentage of positive GalNAc-T3 cells: score 0, 0-5%; score 1, 6-25%; score 2, 26-50%; score 3, 51-75%; or score 4, 76-100% expression levels. To evaluate the correlation with clinicopathological characteristics, GalNAc-T3 expression scores were divided into two groups. Specimens with expression scores of 0-2 were called low expression of GalNAc-T3, and specimens with scores of 3-4 were called high expression of GalNAc-T3.

Statistical analysis

The statistical significance was evaluated using the Pearson's χ^2 test. Survival curves were plotted according to the Kaplan–Meier method (Kaplan and Meier, 1958), and differences between the curves were analysed by the log-rank test (Peto *et al*, 1977). The Cox proportional hazards model was applied to the multivariate survival analysis (Cox, 1972). The results of the Cox proportional hazards model did not seem to change when the follow-up was within 5 years. Data were analysed with the use of Abacus Concepts, Survival Tools for StatView (Abacus Concepts, Inc., Berkeley, CA, USA).

RESULTS

Western blot analysis

To determine the specificity of an anti-GalNAc-T3 antibody, Western blot analysis was performed in 10 cases with frozen normal tissue and tumour tissue from the same patient. GalNAc-T3 expression was detected in 68 kDa protein. High GalNAc-T3 expression breast carcinoma given by Nomoto *et al* (1999) was used as a positive control (lane A). Figure 1 shows the one of Western blotting analysis, in which samples were extracted from normal tissue and tumour tissue of patients with well-differentiated adenocarcinoma and poorly differentiated adenocarcinoma. GalNAc-T3 expression level of tumour tissue increased in comparison with that of normal tissue in well-differentiated adenocarcinoma, although the GalNAc-T3 expression level of tumour tissue decreased in comparison with that of normal tissue



Figure I Western blotting analysis showed GalNAc-T3 expression levels of normal tissue and tumour tissue of patients with well-differentiated adenocarcinoma and poorly differentiated adenocarcinoma. Lane A; positive control (extracted from high GalNAc-T3 expression breast carcinoma), N; normal lung tissue, T; tumour tissue, well; well-differentiated adenocarcinoma, poorly; poorly differentiated adenocarcinoma.

in poorly differentiated adenocarcinoma. The concordance rate between the results of Western blot analysis and the results of IHC detection about GalNAc-T3 expression was 80%.

Immunohistochemical detection of GalNAc-T3 expression in lung adenocarcinoma

In all 148 specimens, 69 (46.6%) stained positive for GalNAc-T3 in the cytoplasm of over 50% of tumour cells, and 79 (53.4%) showed a low expression of GalNAc-T3 in the cytoplasm. In tumour cells, the GalNAc-T3 IHC staining was usually seen in the cytoplasm or cell membranes. In a few cases, immunostaining was also observed in the nucleus as well as in a chromosome in mitosis. But the surrounding normal stromal cells did not react. In normal lung tissues, GalNAc-T3 IHC staining was often seen in the respiratory epithelium and bronchial glands. Typical appearances of staining in high expression of GalNAc-T3 and low expression of GalNAc-T3 tumours are shown in Figure 2a and b, respectively. The relations between GalNAc-T3 expression level and various clinicopathologic characteristics of the patients are summarised in Table 1. The frequency of low GalNAc-T3 expression was significantly lower in well-differentiated adenocarcinomas (27.3%) than in those with moderately and poorly differentiated adenocarcinomas (51.4 and 96.7%, respectively) (P < 0.01). Thus, GalNAc-T3 expression in tumours showed a significant relationship to histologic differentiation. As shown in Table 1, low expression of GalNAc-T3 was associated with pathologic stage (pStage) (P < 0.01), lymph node



Figure 2 GalNAc-T3 IHC detection in adenocarcinoma. (**A**) High GalNAc-T3 expression tumour cells with deep brown stained cytoplasm are shown in the case of well-differentiated adenocarcinoma (original magnification \times 400). (**B**) Low GalNAc-T3 expression tumour cells without stained cytoplasm are shown in the case of poorly differentiated adenocarcinoma (original magnification \times 400).

 Table I
 Relations between the level of GalNAC-T3 expression and clinicopathologic characteristics in 148 lung adenocarcinoma

		GalNAC-T3				
Characteristics	Total	Low	(%)	High	Р	
All cases	148	79	(53.4)	69		
Gender						
Male	88	54	(61.4)	34		
Female	60	25	(41.7)	35	0.02	
Age (years)						
< 65	68	42	(61.8)	26		
≥65	80	37	(46.3)	43	0.06	
Histologic Grade						
Well	44	12	(27.3)	32		
Moderately	74	38	(51.4)	36		
Poorly	30	29	(96.7)	1	< 0.01	
Pathologic stage			. ,			
1	78	28	(35.9)	50		
2	9	4	(44.5)	5		
3	53	42	(79.2)	11		
4	8	5	(62.5)	3	< 0.01	
рТNM			. ,			
T						
ΤI	56	24	(42.9)	32		
T2	51	26	(51.0)	25		
Т3	15	10	(66.7)	5		
T4	26	19	(73.1)	7	0.05	
Ν						
NO	91	33	(36.3)	58		
NI	14	10	(71.4)	4		
N2	41	34	(82.9)	7		
N3	2	2	(100)	0	< 0.01	
М						
MO	140	74	(52.9)	66		
MI	8	5	(62.5)	3	0.60	
Recurrence						
Without recurrence	106	50	(47.2)	56		
With recurrence	42	29	(69.1)	13	0.02	

metastasis (P < 0.01), and tumour recurrence (P = 0.016), but not distant metastasis (P = 0.60). A borderline significant *P*-value was also found in T status (P = 0.05).

Next, GalNAc-T3 expression level was compared to various clinicopathologic parameters of patients with stage I lung adenocarcinoma (tumours with no invasion of adjacent structures and no metastases to regional lymph nodes or distant organs). As shown in Table 2, among 78 patients with lung adnocarcinoma in stage I, a significant relationship was observed between GalNAc-T3 expression and histologic differentiation. In addition, a low expression of GalNAc-T3 was detected in 12 out of 21 (57.1%) cases with recurrent disease, and this finding was statistically significant (P = 0.02). Of the 12 patients, 10 experienced early tumour recurrence (within 2 years of the primary operation). Finally, the pattern of recurrence seemed to be haematogenous. In all, 10 patients had haematogenous distant recurrences, one patient had a lymphogenous recurrence, and one patient had haematogenous recurrences in combination with lymphogenous metastases. Thus, the low expression of GalNAc-T3 showed a significant relationship to early tumour recurrence.

Influence of GalNAc-T3 expression level on survival and recurrence

The influence of GalNAc-T3 expression level on the patients' overall survival was evaluated. The overall 5-year survival rate was 47.4% for the 148 patients in this study. The Kaplan-Meier survival curves demonstrated that the patients with low GalNAc-T3 expression had significantly shorter survival periods than those

		GalNAC-T3				
Characteristics	n	Low	(%)	High	Р	
All cases Histologic grade	78	28	(35.9)	50		
Well	31	6	(19.4)	25		
Moderately	38	14	(36.8)	24		
Poorly	9	8	(88.9)	1	< 0.01	
Pathologic stage			. ,			
IA	46	15	(32.6)	31		
IB	32	13	(40.6)	19	0.47	
Recurrence						
Without recurrence	57	16	(28.1)	41		
With recurrence	21	12	(57.1)	9	0.02	



Figure 3 Survival curves of adenocarcinoma patients with low GalNAc-T3 expression or high GalNAc-T3 expression. (**A**) Overall survival (n = 148), the overall 5-year survival rates in the patients with low GalNAc-T3 expression and high GalNAc-T3 expression were 25.4 and 72.1%, respectively (P < 0.01). (**B**) Survival curves of stage I lung adenocarcinoma patients (n = 78, P < 0.01).

with high GalNAc-T3 expression (P < 0.01) (Figure 3A). The patients with low GalNAc-T3 expression (score 0 and 1) also had significantly shorter survival periods than those with high GalNAc-T3 expression (score 4) (P < 0.01, data not shown) when the

survival curves of three groups patients were determined, according to the cutoff points of 25 and 75%. Among the patients with stage I (n = 78), the overall 5-year survival rates in the patients with low GalNAc-T3 expression and high GalNAc-T3 expression were 41.6 and 82%, respectively (P < 0.01) (Figure 3B). In univariate analysis, six variables (gender, pT, pN, pM, grade of differentiation, and high or low expression of GalNAc-T3) were found significantly to affect survival in 148 adenocarcinoma patients. Furthermore, a multivariate survival analysis demonstrated that five variables (pT, pN, pM, grade of differentiation, and high or low expression of GalNAc-T3) were independently associated with patient survival (Table 3). The low expression of GalNAc-T3 was estimated to be associated with an increased risk of death by a factor of 2.22 in multivariate analysis (P < 0.01). On the other hand, in univariate analysis, three variables (pStage, grade of differentiation, and high or low expression of GalNAc-T3) were found to affect significantly survival in 78 patients with stage I adenocarcinoma. Furthermore, a multivariate survival analysis demonstrated that two variables (grade of differentiation, and high or low expression of GalNAc-T3) were independently associated with patient survival and low expression of GalNAc-T3 was estimated to be associated with an increased risk of death by a factor of 2.99 in multivariate analysis (P < 0.01; data not shown).

To investigate whether GalNAc-T3 expression level is significantly correlated with recurrence in early-stage diseases, disease-free survival curves of the patients with GalNAc-T3 expression in stage I were analysed using the Kaplan–Meier method. As shown in Figure 4, the patients with low GalNAc-T3 expression had significantly shorter disease-free survival periods than those with high GalNAc-T3 expression (P < 0.01). In univariate analysis, two variables (grade of differentiation and high or low expression of GalNAc-T3) were found to affect significantly disease-free survival in 78 patients with stage I adenocarcinoma. The variable of pStage was estimated as a borderline association (P = 0.06). A multivariate diseade-free survival analysis demonstrated that the variable of grade of high or low expression of GalNAc-T3 was independently associated with disease-free survival. The low expression of

 Table 3
 Univariate and multivariate analyses using a proportional hazard model for survival of 148 lung adenocarcinoma patients

		Univariate analysis			Multivariate analysis		
Variable	n	95% CI	HR	Р	95% CI	HR	Р
Gender							
Female	60		1			I	
Male	88	1.02-2.54	1.61	0.04	0.71-1.85	1.15	0.58
рT							
ΤI	56		1			I	
Т2	51	1.39-4.45	2.49	< 0.01	1.28-4.38	2.37	< 0.01
Т3	15	1.96-8.52	4.09	< 0.01	1.06-5.37	2.38	0.04
T4	26	3.54-12.5	6.66	< 0.01	2.16-8.72	4.34	< 0.01
PN							
N0	91		1			I	
NI	14	1.86-6.94	3.59	< 0.01	0.76-3.18	1.55	0.23
N2	41	2.31-5.98	3.71	< 0.01	1.05-3.20	1.83	0.03
N3	2	1.23-21.7	5.17	0.03	2.46-59.1	12.1	< 0.01
pМ							
MO	140		I			Ι	
MI	8	1.85-9.26	4.13	< 0.01	0.93-5.18	2.19	0.07
Grade							
Well	44		I			Ι	
Moderately	74	2.09-7.66	4.00	< 0.01	1.46-6.07	2.98	< 0.01
Poorly	30	2.11-9.37	4.44	< 0.0	0.74-3.88	1.69	0.22
GalNAC-T3							
High expression	69		1			Ι	
Low expression	79	2.20-5.65	3.53	< 0.0	1.26-3.91	2.22	< 0.0

HR-hazard ratio.

439



Figure 4 Disease-free survival curves of stage I lung adenocarcinoma patients with low GalNAc-T3 expression or high GalNAc-T3 expression (n = 78). The disease-free 3-year survival rates in the patients with low GalNAc-T3 expression and high GalNAc-T3 expression were 59 and 87.9%, respectively (P < 0.01).

 Table 4
 Univariate and multivariate analyses using a proportional hazard model for disease-free survival of 78 patients with stage I adenocarcinoma

	Univariate analysis			Multivariate analysis			
Variable	n	95% CI	HR	Р	95% CI	HR	Р
þStage							
ĬĂ	46		1			1	
IB	32	0.96-5.43	2.29	0.06	0.83-5.01	2.04	0.12
Grade							
Well	31		1			1	
Moderately	38	1.12-10.3	3.40	0.03	0.87-8.39	2.70	0.09
Poorly	9	0.79-16.0	3.56	0.10	0.25-6.60	1.29	0.76
GalNAC-T3							
High expression	50		1			1	
Low expression	28	1.38-7.86	3.29	< 0.0	1.22-7.65	3.05	0.02

HR = hazard ratio.

GalNAc-T3 was estimated to be associated with an increased risk of death by a factor of 3.05 in multivariate analysis (P = 0.02) (Table 4).

DISCUSSION

Lung cancer is the leading cause of cancer-related deaths in North America, and it became the most common cause of cancer-related deaths among Japanese in 1998 (Ginsberg *et al*, 1997; Hara *et al*, 2000). Of the four main histologic types of lung cancer, adenocarcinoma is the most commonly occurring type in Japan (Hara *et al*, 2000). Lung cancer is also an aggressive carcinoma with a poor outcome, and the overall 5-year survival rate is about 20% (Landis *et al*, 1998; Hara *et al*, 2000). Therefore, it is important to evaluate the malignant potential of tumour cells for a more precise evaluation of the prognosis for patients with lung adenocarcinoma.

The current study investigated the associations between GalNAc-T3 expression level and various clinicopathologic characteristics of patients with lung adenocarcinoma. The low expression of GalNAc-T3 was associated with poorly differentiated tumour, poorly pathologic stage, lymph node metastasis and tumour recurrence. Both univariate and multivariate analyses demonstrated that, among the clinicopathologic factors examined, low expression of GalNAc-T3 was a significant independent factor for predicting poor prognosis and early recurrence.

Concerning the differentiation status of lung adenocarcinoma and GalNAc-T3 expression levels, low expression levels of GalNAc-T3 correlated with poorly differentiated adenocarcinoma and high expression levels of GalNAc-T3 correlated with well-differentiated adenocarcinoma. These findings are consistent with the report by Sutherlin *et al* (1997) that GalNAc-T3 expression is higher in welldifferentiated pancreatic adenocarcinoma cell lines. Generally, poorly differentiated adenocarcinoma are thought to have a greater malignant potential than well-differentiated adenocarcinoma. Thus, GalNAc-T3 expression level may be a marker of malignant potential in lung adenocarcinoma.

It is well known that the metastatic process consists of several stages: tumour cell escape from primary tumour, invasion of the vessels, migration, adhesion to the vascular endothelium, extravasation, and colonisation, all of which are essential for the development of clinically overt metastases. Among O-linked carbohydrate antigens, sialyl LewisX has been reported to function as a ligand of the endothelial cell adhesion molecule E-selectin (ELAM-1), which adheres human cancer cells to the vascular endothelium (Phillips et al, 1990; Walz et al, 1990; Takada et al, 1993). In addition, a mucin antigen such as MUC1 inhibits the Ecadherin-mediated cell-cell adhesion, which implies that MUC1 mucin enables adenocarcinoma cells to be detached readily from the primary tumour tissue (Wesseling et al, 1996). MUC1 polypeptide is also associated with cell surface O-linked glycoproteins such as sialyl LewisX and sialyl LewisA (Ho et al, 1995). The initial glycosylation of mucin-type O-linked carbohydrate antigens is regulated in part by the differential expression of GalNActransferase, which add GalNAc to fibronectine (Bennett et al. 1999b) and may affect the density or positions of attachment of Olinked oligosaccharides (Sutherlin et al, 1997). Differential expression of GalNAc-transferase may have a 'valvular' function in regulating O-glycosylation in mucin-type O-linked carbohydrate antigens such as sialyl LewisX and MUC1 (Wandall et al, 1997; von Mensdorff-Pouilly et al, 2000). Therefore, GalNAc-transferase expression may play a role in tumour cell escape from primary tumour, adhesion to the vascular endothelium, and extravasation during the metastatic process.

In lung cancer, even after a 'curative' resection for pathologic stage I disease, about 30% of patients experience recurrence and eventually die of the disease (Little et al, 1986; Ginsberg et al, 1997). This suggests that occult metastases are present at the time of surgical intervention. Previous studies (Gu et al 2002) showed that micrometastatic tumour cells (cytokeratin positive cells) were present in pathologic negative lymph nodes in 34.7% of stage I non-small-cell lung cancer (NSCLC) patients after complete resectioning, and patients with micrometastatic tumour cells had a poor prognosis and a high rate of recurrent disease. To see whether GalNAc-T3 expression correlates with micrometastases, micrometastatic tumour cells were detected in a total of 1436 hilar and mediastinal pathologic negative lymph nodes from 65 patients with stage I lung adenocarcinoma, using the method previously described (Gu et al, 2002), Of 65 patients, 19 exhibited lymph nodal micrometastasis, and a low expression of GalNAc-T3 was found in 12 patients (63.2%) (P = 0.003). Furthermore, this may be a partial explanation for the relationship between aberrant expression of GalNAc-T3 and early metastasis (unpublished data, with subjects different from those of this study). In the current study, among 12 out of 78 patients with low GalNAc-T3 expression and recurrent diseases in stage I lung adenocarcinoma, tumour recurrence within 2 years of the primary operation was found in 10 patients. The pattern of recurrence seemed to be haematogenous, and a low expression of GalNAc-T3 was associated with poor prognosis and early recurrence. Based on these results, it is

Molecular and Cellular Pathology

reasonable to argue that a low expression of GalNAc-T3 may be a useful indicator of early tumour recurrence in stage I lung adenocarcinoma.

In conclusion, low expression of GalNAc-T3 may be a useful marker in predicting poor prognosis and early recurrence, not only in all completely resected patients with adenocarcinoma of the lung but also in those with stage I diseases. These patients

REFERENCES

- Bennett EP, Hassan H, Clausen H (1996) cDNA cloning and expression of a novel human UDP-*N*-acetyl-alpha-D-galactosamine. Polypeptide *N*-acetylgalactosaminyltransferase, GalNAc-T3. *J Biol Chem* **271**: 17006– 17012
- Bennett EP, Hassan H, Mandel U, Mirgorodskaya E, Roepstorff P, Burchell J, Taylor-Papadimitriou J, Hollingsworth MA, Merkx G, van Kessel AG, Eiberg H, Steffensen R, Clausen H (1998) Cloning of a human UDP-*N*-acetyl-alpha-D-galactosamine: polypeptide *N*-acetylgalactosaminyltransferase that complements other GalNAc-transferases in complete O-glycosylation of the MUC1 tandem repeat. *J Biol Chem* **273:** 30472-30481
- Bennett EP, Hassan H, Hollingsworth MA, Clausen H (1999a) A novel human UDP-N-acetyl-D-galactosamine: polypeptide N-acetylgalactosaminyltransferase, GalNAc-T7, with specificity for partial GalNAcglycosylated acceptor substrates. *FEES Lett* **460**: 226–230
- Bennett EP, Hassan H, Mandel U, Hollingsworth MA, Akisawa N, Ikematsu Y, Merkx G, van Kessel AG, Olofsson S, Clausen H (1999b) Cloning and characterization of a close homologue of human UDP-*N*-acetyl- alpha-D-galactosamine: Polypeptide *N*-acetylgalactosaminyltransferase-T3, designated GalNAc-T6. Evidence for genetic but not functional redundancy. *J Biol Chem* 274: 25362-25370
- Bolscher JG, Schallier DC, van Rooy H, Storme GA, Smets LA (1988) Modification of cell surface carbohydrates and invasive behavior by an alky] lysophospholipid. *Cancer Res* **48**: 977–982
- Cao Y, Karsten UR, Liebrich W, Haensch W, Springer GF, Schlag PM (1995) Expression of Thomsen-Friedenreich-related antigens in primary and metastatic colorectal carcinomas. A reevaluation. *Cancer* 76: 1700-1708
- Clausen H, Bennett EP (1996) A family of UDP-GalNAc: polypeptide N-acetylgalactosaminyl-transferases control the initiation of mucin-type O-linked glycosylation. *Glycobiology* **6:** 635–646

Cox D (1972) Regression models and life tables. J R Stat Soc 34: 187-220

- Diez M, Torres A, Maestro ML, Ortega MD, Gomez A, Pollan M, Lopez JA, Picardo A, Hernando F, Balibrea JL (1996) Prediction of survival and recurrence by serum and cytosolic levels of CEA, CA125 and SCC antigens in resectable non-small-cell lung cancer. *Br J Cancer* **73**: 1248–1254
- Fogel M, Altevogt P, Schirrmacher V (1983) Metastatic potential severely altered by changes in tumor cell adhesiveness and cell-surface sialylation. *J Exp Med* **157**: 371-376
- Ginsberg R, Yokes E, Raben A (1997) Non-small cell lung cancer. In *Cancer, Principles and Practice of Oncology*, De Vita Jr V, Helleman S, Rosenberg S (eds) pp 858–911. Philadelphia: Lippencott-Raven
- Gu C, Osaki T, Oyama T, Inoue M, Kodate M, Dobashi K, Oka T, Yasumoto K (2002) Detection of micrometastatic tumor cells in pN0 lymph nodes of patients with completely resected nonsmall cell lung cancer: impact on recurrence and survival. *Ann Surg* 235: 133–139
- Hagen FK, Van Wuyckhuyse B, Tabak LA (1993) Purification, cloning, and expression of a bovine UDP-GalNAc: polypeptide *N*-acetyl-galactosaminyltransferase. *J Biol Chem* **268**: 18960–18965
- Hara N, Nakanishi Y, Izumi M (2000) Epidemiology of lung cancer in Japan. Nippon Rinsho 58: 1005-1011
- Ho JJ, Siddiki B, Kim YS (1995) Association of sialyl-Lewis(a) and sialyl-Lewis(x) with MUC-1 apomucin in a pancreatic cancer cell line. *Cancer Res* 55: 3659-3663
- Homa FL, Hollander T, Lehman DJ, Thomsen DR, Elhammer AP (1993) Isolation and expression of a cDNA clone encoding a bovine UDP-GalNAc: polypeptide *N*-acetylgalactosaminyltransferase. *J Biol Chem* **268**: 12609–12616
- Itzkowitz SH, Bloom EJ, Kokal WA, Modin G, Hakomori S, Kim YS (1990) Sialosyl-Tn. A novel mucin antigen associated with prognosis in colorectal cancer patients. *Cancer* **66**: 1960–1966

need to be followed up carefully after surgery. At present, postoperative adjuvant chemotherapy is not a routine standard therapy for completely resected NSCLC patients, because it has not been shown to improve patient outcomes consistently. However, by assessing the GalNAc-T3 expression level, it would be possible to select patients who might benefit most from adjuvant chemotherapy.

- Kaplan E, Meier P (1958) Nonparametric estimation from incomplete observations. J Am Stat Assoc 53: 457-481
- Landis SH, Murray T, Bolden S, Wingo PA (1998) Cancer statistics, 1998. CA Cancer J Clin 48: 6-29
- Little AG, DeMeester TR, Ferguson MK, Skinner DB, Hoffman PC, Skosey C, Blough RR, Golomb HM (1986) Modified stage I (T1N0M0, T2N0M0), non-small cell lung cancer: treatment results, recurrence patterns, and adjuvant immunotherapy. *Surgery* **100**: 618–621
- Matsushita Y, Nakamori S, Seftor EA, Hendrix MJ, Irimura T (1991) Human colon carcinoma cells with increased invasive capacity obtained by selection for sialyl-dimeric LeX antigen. *Exp Cell Res* 196: 20-25
- Mountain C (1997) Revisions in the International System for Staging Lung Cancer. Chest 111: 1710–1717
- Nakamori S, Ota DM, Cleary KR, Shirotani K, Irimura T (1994) MUC1 mucin expression as a marker of progression and metastasis of human colorectal carcinoma. *Gastroenterology* **106:** 353–361
- Niklinski J, Furman M, Laudanski J, Kozlowski M. (1992) Prognostic value of pretreatment CEA, SCC-Ag and CA 19-9 levels in sera of patients with non-small cell lung cancer. *Eur J Cancer Prev* 1: 401–416
- Nomoto M, Izumi H, Ise T, Kato K, Takano H, Nagatani G, Shibao K, Ohta R, Imamura T, Kuwano M, Matsuo K, Yamada Y, Itoh H, Kohno K (1999) Structural basis for the regulation of UDP-*N*acetyl-alpha-D-galactosamine: polypeptide *N*-acetylgalactosaminyl transferase-3 gene expression in adenocarcinoma cells. *Cancer Res* 59: 6214-6222
- Ogawa J, Sano A, Koide S, Shohtsu A (1994) Relation between recurrence and expression of proliferating cell nuclear antigen, sialyl LewisX, and sialyl Lewis(a) in lung cancer. J Thorac Cardiovasc Surg 108: 329-336
- Ohgami A, Tsuda T, Osaki T, Mitsudomi T, Morimoto Y, Higashi T, Yasumoto K (1999) MUC1 mucin mRNA expression in stage I lung adenocarcinoma and its association with early recurrence. *Ann Thorac Surg* 67: 810-814
- Peto R, Pike MC, Armitage P, Breslow NE, Cox DR, Howard SV, Mantel N, McPherson K, Peto J, Smith PG (1977) Design and analysis of randomised clinical trials requiring prolonged observation of each patient. Br J Cancer 35: 1-39
- Phillips ML, Nudelman E, Gaeta FC, Perez M, Singhal AK, Hakomori S, Paulson JC (1990) ELAM-1 mediates cell adhesion by recognition of a carbohydrate ligand, sialyl-Lex. *Science* **250**: 1130-1132
- Rice GE, Bevilacqua MP (1989) An inducible endothelial cell surface glycoprotein mediates melanoma adhesion. *Science* **246**: 1303–1306
- Sutherlin ME, Nishimori I, Caffrey T, Bennett EP, Hassan H, Mandel U, Mack D, Iwamura T, Clausen H, Hollingsworth MA (1997) Expression of three UDP-N-acetyl-alpha-D-galactosamine: polypeptide GalNAc N-acetylgalactosaminyl transferases in adenocarcinoma cell lines. Cancer Res 57: 4744-4748
- Takada A, Ohmori K, Yoneda T, Tsuyuoka K, Hasegawa A, Kiso M, Kannagi R (1993) Contribution of carbohydrate antigens sialyl Lewis A and sialyl Lewis X to adhesion of human cancer cells to vascular endothelium. *Cancer Res* **53**: 354-361
- Ten Hagen KG, Hagen FK, Balys MM, Beres TM, Van Wuyckhuyse B, Tabak LA (1998) Cloning and expression of a novel, tissue specifically expressed member of the UDP-GalNAc:polypeptide *N*-acetylgalactosa-minyltransferase family. *J Biol Chem* **273**: 27749–27754
- von Mensdorff-Pouilly S, Petrakou E, Kenemans P, van Uffelen K, Verstraeten AA, Snijdewint FG, van Kamp GJ, Schol DJ, Reis CA, Price MR, Livingston PO, Hilgers J (2000) Reactivity of natural and induced human antibodies to MUC1 mucin with MUC1 peptides and *n*-acetylgalactosamine (GalNAc) peptides. *Int J Cancer* 86: 702-712

- Walz G, Aruffo A, Kolanus W, Bevilacqua M, Seed B (1990) Recognition by ELAM-1 of the sialyl-Lex determinant on myeloid and tumor cells. *Science* 250: 1132-1135
- Wandall HH, Hassan H, Mirgorodskaya E et al. (1997) Substrate specificities of three members of the human UDP-N-acetyl- alpha-D-galactosamine: Polypeptide N-acetylgalactosaminyltransferase family, GalNAc-T1, -T2, and -T3. Biol Chem 272: 23503-23514
- Wesseling J, van der Valk SW, Hilkens J (1996) A mechanism for inhibition of E-cadherin-mediated cell-cell adhesion by the membrane-associated mucin episialin/MUC1. *Mol Biol Cell* **7**: 565-577
- White T, Bennett EP, Takio K, Sorensen T, Bonding N, Clausen H (1995) Purification and cDNA cloning of a human UDP-N-acetyl-alpha-Dgalactosamine: polypeptide N-acetylgalactosaminyltransferase. J Biol Chem 270: 24156-24165