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The downstream modulator of interferon- γ , STAT1 is not genetically associated to the Dutch coeliac disease population

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Coeliac disease (CD) is a complex genetic disorder. Its etiology is owing to multiple genes and environmental factors, such as gluten. The first event in the pathogenesis of CD after the ingestion of gluten is the activation of a Th1 immune response that leads to villous atrophy. Although this immune response seems crucial to the disease's development, only the HLA-DQ2/DQ8 genes have been identified as causative immune genes related to CD. Recently, the activation of the transcription factor STAT1 and changes in its expression levels have confirmed the participation of the Janus kinase-signal transducer and activator of transcription pathway in CD. Furthermore, as the *STAT-1* gene is a positional candidate located in the CELIAC3 locus on chromosome 2, we speculate that alterations in this gene could be primarily responsible for the aberrant immune response that characterizes CD. Based on this functional and genetic evidence, we investigated the primary contribution of *STAT-1* to CD. We performed a comprehensive genetic association study using five tag SNPs fully covering the *STAT-1* gene in a Dutch cohort of 355 independent CD cases and 360 healthy controls. Neither the alleles, nor the genotypes in the case-control genetic association studies, nor the haplotype analysis showed any association to the *STAT-1* gene in the Dutch CD population. Our results do not point to a primary involvement of the *STAT-1* gene in the Dutch CD population.

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Introduction

Coeliac disease (CD) is a complex genetic disorder in which the main causative environmental factor is the dietary protein gluten. This is present in common Western foods, such as bread, pasta and many processed products. When

gluten is ingested by coeliac patients, it provokes an inflammatory reaction in the small intestine that leads to crypt hyperplasia and villous atrophy.¹ Although the exact nature of this immune response remains unknown, there is much evidence to suggest that gluten promotes a Th1 immune response mostly mediated by high levels of interferon-gamma (IFN- γ)² and $\alpha\beta$ CD4+ T cells that recognize gluten peptides through HLA-DQ2/DQ8 molecules.³ This observation is further supported by genetic data showing a primary association between the genes encoding the HLA-DQ2/DQ8 molecules on chromosome 6 and all the CD populations tested to date.^{4,5} Despite the fact that these results indicate an essential role for HLA in

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the etiopathogenesis of CD, the HLA-DQ2/DQ8 genes only explain approximately 40% of the familial clustering to CD,^{5,6} implying that other non-HLA genes must also be associated with CD. As the Th1 immune response is such a crucial factor in the pathogenesis of CD, the search for other primary genes should focus on genes linked to the Th1 immune response.

The Janus kinase-signal transducer and activator of transcription (JAK-STAT) pathway plays an important role in regulating the immune response. This pathway mediates the early Th1 proinflammatory IFN- γ -directed response by activating gene transcription through different STAT proteins (Figure 1a). The activation of an aberrant Th1 immune reaction initiated by the presence of gluten is a crucial event in the pathogenesis of CD.⁷ Some studies have shown the importance of the JAK-STAT pathway in the pathogenesis of CD.^{8–10} One interesting activator of the JAK-STAT pathway in CD might be IFN- γ , which is also highly expressed in the intestine of coeliac individuals.² The exact role of IFN- γ in the pathogenesis of CD has not been elucidated, but it might contribute to the maintenance of the chronic inflammatory response through the JAK-STAT pathway by regulating the expression and the activation of STAT1.

Although IFNG might play an important role in CD pathogenesis,² genetic association studies have failed to show that this gene makes a primary contribution to the disease.^{2,11} Hence, it seems attractive to test the *STAT1* gene, as it is one of IFN- γ first downstream targets. The *STAT1* gene is both a positional and functional candidate gene. The *STAT1* gene is located in the CELIAC3 locus on chromosome 2q23–32,¹² which also encompasses the

ICOS-CTLA4-CD28 gene cluster. Several studies have investigated the contribution of the *CTLA4* gene in CD as this locus has been repeatedly shown to be associated with autoimmune disorders.¹³ However, the genome-wide linkage study performed in the Dutch population did not show significant linkage to this region.⁵ Initial genetic association studies on the *CTLA4* gene in the Dutch population showed borderline significance for the CT60-G allele ($P=0.048$), but this did not remain significant after correcting for multiple testing¹⁴ and could also not be replicated in an expanded cohort.¹⁵ Moreover, the weak association reported by some groups is insufficient to explain the linkage peak. It is likely that there is another gene from this region associated with CD susceptibility. Furthermore, linkage studies performed in this region have shown that the markers with the highest significant values are located outside the *ICOS/CTLA4/CD28* gene boundaries and toward the centromeric side of the *ICOS/CTLA4/CD28* genes, where the *STAT1* gene is located^{4,12,16} (Figure 2). The *STAT1* gene is located 12.9 Mb centromeric to the *CTLA4* gene, 3.7 Mb from D2S117, which was identified in a meta-analysis performed in European populations ($P=0.023$),⁴ 2.2 Mb from D2S1776, which showed an lod score of 2.3 in a genome-wide linkage study carried out in the Finish population¹² and 10.9 Mb from D2S2214 found in a meta-analysis that included five North European population ($P=0.0001$)¹⁶ (Figure 2).

From a functional point of view, the STAT1 protein has been shown to be activated in the mononuclear cells of the lamina propria and epithelial cells of CD biopsies.⁸ These findings are further substantiated by mRNA expression

Figure 1 Main components of the JAK-STAT pathway and differentially expressed genes in our expression studies. (a) The JAK-STAT pathway mediates the early Th1 proinflammatory IFN- γ -directed response by activating gene transcription through the STAT1 protein. (b) mRNA expression levels of the *STAT1* gene, IFNG, as well as multiple downstream targets of the JAK-STAT1 cascade on biopsies of CD patients.

Figure 2 Overview of the markers and their location in the CELIAC3 region. These have been obtained from three reported studies in CD.^{4,12,16} The position of the markers and genes is given in bps according to Ensembl v37 Feb 2006. The figure is not drawn to scale.

studies on biopsies of CD patients, with upregulation of the *STAT1* gene, IFNG, as well as multiple downstream targets of this signal-transduction cascade (Figure 1b) (Diosdado *et al* in preparation). These observations suggest that *STAT1* might be critical in controlling the immunological activity in CD and, hence, any genetic change that alters its transcriptional activity could explain the detrimental immune response seen in CD. Based on this evidence, we carried out a comprehensive approach using tag SNPs to investigate the genetic contribution of the non-HLA gene *STAT1* in the Dutch CD population in a case-control cohort.

Subjects and methods

DNA samples for genetic association studies

For the genetic association study, we collected a group of 355 independent CD cases and 360 independent blood bank controls, all of Dutch Caucasian origin. All CD patients were diagnosed according to the ESPGHAN criteria¹⁷ and were selected for showing a Marsh III lesion at initial diagnosis. Ninety-three percent of the patients were HLA-DQ2 positive. We collected blood samples and isolated DNA according to standard laboratory procedures.⁵

Informed consent was obtained from all individuals and the study was approved by the Medical Ethics Committee of the University Medical Centre, Utrecht, The Netherlands.

Power calculations

We have more than 80% power to detect a disease variant with minor allele frequencies ranging from 0.1 to 0.3, assuming a relative risk of 1.7, a dominant inheritance, a disease prevalence of the 0.1, a D' of 1 and an equal frequency of the tested SNP and the high-risk variant. In case of a different frequency of the high-risk allele compared to the tested SNP or an imperfect tagging, the power drops. In case of a codominant effect, the power increases.

SNP selection and data analysis

To select the tag SNPs, we downloaded the genotype information on the SNPs for the *STAT1* gene from the HAPMAP database release 18 phase I (<http://www.hapmap.org/>) (typed in the CEPH population of Utah residents

with ancestors from Northern and Western Europe). We uploaded this to the Tagger program (<http://www.broad.mit.edu/mpg/tagger/>). This software returned six nonredundant tag SNPs with a minor allele frequency (MAF) $\geq 10\%$ and an $r^2 \geq 0.8$ that covered the *STAT1* gene. For these SNPs, two assay-on-demand probes: rs3771300 (C_22274194_10) and rs1914408 (C_1737196_10); and four assay-by-design probes: rs7562024 (C_1737208_10), rs1547550 (C_1737199_10), rs13010343 (C_1737197_10) and rs2030171 (C_1737225_20) were obtained. All the tested SNPs were analyzed on an ABI Prism 7900 HT system. All reagents and equipment were purchased from Applied Biosystems (Foster, CA, USA) and used according to their protocols.

Hardy-Weinberg equilibrium was evaluated separately in the case and control groups, for all tested SNPs (data not shown). Differences in allele frequencies and genotypes were compared between cases and controls using a χ^2 test.

Results

To study the *STAT1* gene as a causative factor in CD, we selected six tag SNPs for further genetic association studies (Table 1). Although the SNP rs1547550 could not be tested owing to manufacturing failures, the other five tag SNPs captured 90% of the untyped SNPs present in the gene with an $r^2 > 0.8$ and MAF $> 10\%$. All SNPs were in HWE (data not shown). The association studies showed no statistical difference between the cases and controls for either allele (Table 1a) or genotype frequencies (Table 1b). Haplotype analysis did not improve the results (data not shown). We therefore concluded that the *STAT1* gene is not associated to CD in the Dutch population.

Discussion

The CELIAC3 region has been found to be significantly linked to CD in the Scandinavian population and was also reported in a meta-analysis carried out by a European consortium on CD.^{4,12} This locus contains a number of interesting candidate genes for CD, but so far attention has mainly been focused on the *CTLA4* gene, as it has shown association to other autoimmune disorders.¹³ Nevertheless, genetic studies in CD have been rather inconclusive and do not clearly point to a genetic association with

Table 1 *P*-values obtained from the genetic association study in the *STAT1* gene using tag SNPs on 355 CD patients and 360 controls. (a) Tag SNP information and association studies on allele frequencies. (b) Association studies on genotype frequencies

SNP rs number	Bp position	SNP type	MAF ^a	Cases (%)	Controls (%)	χ ²	P-value
(a)							
rs3771300	191661102	T/G	G	48.0	48.5	1.78	0.1821
rs1914408	191665482	C/T	T	25.5	24.9	0.078	0.7807
rs13010343	191668951	A/G	A	15.8	12.4	3.36	0.0668
rs7562024	191681027	C/T	T	35.9	37.1	0.226	0.6344
rs2030171	191694669	A/G	A	32.9	31.8	0.206	0.6502
SNP rs number	Genotype	Cases (%)	Controls (%)	χ ^{2b}	P-value ^b		
(b)							
rs3771300	GG	89 (25.6)	96 (26.8)	1.68	0.19		
	GT	155 (44.7)	177 (49.4)				
	TT	103 (29.7)	85 (23.7)				
rs1914408	CC	194 (55.9)	200 (56.8)	0.00	0.96		
	CT	129 (37.2)	129 (36.6)				
	TT	24 (6.9)	23 (6.5)				
rs13010343	AA	250 (71.8)	271 (77.2)	1.64	0.20		
	AG	86 (24.7)	73 (20.8)				
	GG	12 (3.4)	7 (2.0)				
rs7562024	CC	145 (41.8)	142 (40.7)	0.03	0.87		
	CT	155 (44.7)	155 (44.4)				
	TT	47 (13.5)	52 (14.9)				
rs2030171	AA	35 (10.2)	38 (10.9)	0.24	0.63		
	AG	155 (45.3)	145 (41.7)				
	GG	152 (44.4)	165(47.4)				

^aMinor allele frequency.^bOverall χ^2 *P*-value of all three genotypes.

CTLA4.^{12,14–16,18–29} Therefore, it is tempting to speculate that another gene in this region may be the causative one in CD pathogenesis. In this study, we selected as a candidate the *STAT1* gene. Its protein is one of the main downstream modulators of IFN- γ signaling through the JAK-STAT pathway. The *STAT1* gene is located in the CELIAC3 region on chromosome 2q23–32, and its participation in the pathogenesis of CD is suggested by persistent activation at the lamina propria, and the upregulation of IFNG, several components of the JAK-STAT pathway and its downstream targets.^{8–10}

We therefore conducted genetic association studies in a Dutch case–control cohort to determine the contribution of the *STAT1* gene in the etiology of CD. Our association studies did not reach statistical significance for any of the tested tag SNPs. Although we cannot exclude a minor effect of this gene in the Dutch population, our results gave no evidence for *STAT1* being a strong causative factor in the Dutch CD population.

Despite the results from the genetic studies on IFNG and *STAT1*, the importance of the JAK-STAT1 pathway in the immune response to gluten is still remarkable. However, at this point, we cannot exclude that upregulation of the

downstream STAT-1 targets is secondary to the high IFN- γ production in the lamina propria of CD patients. Unfortunately, there have been no functional experiments to determine the exact nature of the stimuli that initiate the JAK-STAT1 pathway in CD. It would be logical to assume that the major stimulator of this pathway is IFNG, as large amounts of it are seen in the biopsies of CD patients, but other cytokines such as IL-6 can also activate this pathway.³⁰ Additional studies on the JAK-STAT1 pathway, revealing the range of genes regulated by this transcription factor under the presence of gluten, are important, and should reveal new aspects of the immune response that occurs in CD. We still need to determine whether other STAT (STAT1–6) and JAK proteins are activated in the coeliac intestine, and especially, which molecules of the regulatory mechanisms of this pathway (the suppressor of cytokine signaling (SOCS) family, the protein inhibitor of activated STAT (PIAS) proteins and protein tyrosine phosphatases (PTPs)) participate in the pathogenesis of CD. Interestingly, some of the molecules of the regulatory mechanisms have already been related to immune diseases, like the regulatory molecule protein, tyrosine phosphatase receptor type C (*PTPRC*) that has been associated to

multiple sclerosis.³¹ Identifying other members of the JAK-STAT pathway participating in CD, and the molecules that maintain its balance, might therefore reveal potential candidate genes for further genetic association studies in CD.

In conclusion, although functional studies imply that STAT1 plays an important role in regulating the extensive activation of the adaptive immune response in the lamina propria of CD, genetic association studies do not show any significant association of this gene to the Dutch CD population.

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References

- Marsh MN: Gluten, major histocompatibility complex, and the small intestine. A molecular and immunobiologic approach to the spectrum of gluten sensitivity ('celiac sprue'). *Gastroenterology* 1992; **102**: 330–354.
- Wapenaar MC, van Belzen MJ, Fransen JH *et al*: The interferon gamma gene in celiac disease: augmented expression correlates with tissue damage but no evidence for genetic susceptibility. *J Autoimmun* 2004; **23**: 183–190.
- Sollid LM: Coeliac disease: dissecting a complex inflammatory disorder. *Nat Rev Immunol* 2002; **2**: 647–655.
- Babron MC, Nilsson S, Adamovic S *et al*: Meta and pooled analysis of European coeliac disease data. *Eur J Hum Genet* 2003; **11**: 828–834.
- Van Belzen MJ, Meijer JW, Sandkuijl LA *et al*: A major non-HLA locus in celiac disease maps to chromosome 19. *Gastroenterology* 2003; **125**: 1032–1041.
- Bevan S, Popat S, Braegger CP *et al*: Contribution of the MHC region to the familial risk of coeliac disease. *J Med Genet* 1999; **36**: 687–690.
- Sollid LM: Molecular basis of celiac disease. *Annu Rev Immunol* 2000; **18**: 53–81.
- Mazzarella G, MacDonald TT, Salvati VM *et al*: Constitutive activation of the signal transducer and activator of transcription pathway in celiac disease lesions. *Am J Pathol* 2003; **162**: 1845–1855.
- Monteleone G, Pender SL, Wathen NC, MacDonald TT: Interferon-alpha drives T cell-mediated immunopathology in the intestine. *Eur J Immunol* 2001; **31**: 2247–2255.
- Monteleone I, Monteleone G, Del Vecchio Blanco G *et al*: Regulation of the T helper cell type 1 transcription factor T-bet in coeliac disease mucosa. *Gut* 2004; **53**: 1090–1095.
- Woolley N, Mustalahti K, Maki M, Partanen J: Cytokine gene polymorphisms and genetic association with coeliac disease in the Finnish population. *Scand J Immunol* 2005; **61**: 51–56.
- Rioux JD, Karinen H, Kocher K *et al*: Genomewide search and association studies in a Finnish celiac disease population: identification of a novel locus and replication of the HLA and CTLA4 loci. *Am J Med Genet A* 2004; **130**: 345–350.
- Kristiansen OP, Larsen ZM, Pociot F: CTLA-4 in autoimmune diseases – a general susceptibility gene to autoimmunity? *Genes Immun* 2000; **1**: 170–184.
- van Belzen MJ, Mulder CJ, Zhernakova A, Pearson PL, Houwen RH, Wijmenga C: CTLA4 +49 A/G and CT60 polymorphisms in Dutch coeliac disease patients. *Eur J Hum Genet* 2004; **12**: 782–785.
- Zhernakova A, Eerligh P, Barrera P *et al*: CTLA4 is differentially associated with autoimmune diseases in the Dutch population. *Hum Genet* 2005; **118**: 58–66.
- Popat S, Hearle N, Hogberg L *et al*: Variation in the CTLA4/CD28 gene region confers an increased risk of coeliac disease. *Ann Hum Genet* 2002; **66**: 125–137.
- Revised criteria for diagnosis of coeliac disease: Report of working group of European society of paediatric gastroenterology and nutrition. *Arch Dis Child* 1990; **65**: 909–911.
- Hunt KA, McGovern DP, Kumar PJ *et al*: A common CTLA4 haplotype associated with coeliac disease. *Eur J Hum Genet* 2005; **13**: 440–444.
- Naluai AT, Nilsson S, Samuelsson L *et al*: The CTLA4/CD28 gene region on chromosome 2q33 confers susceptibility to celiac disease in a way possibly distinct from that of type 1 diabetes and other chronic inflammatory disorders. *Tissue Antigens* 2000; **56**: 350–355.
- Djilali-Saiah I, Schmitz J, Harfouch-Hammoud E, Mougenot JF, Bach JF, Caillat-Zucman S: CTLA-4 gene polymorphism is associated with predisposition to coeliac disease. *Gut* 1998; **43**: 187–189.
- Clot F, Fulchignoni-Lataud MC, Renoux C *et al*: Linkage and association study of the CTLA-4 region in coeliac disease for Italian and Tunisian populations. *Tissue Antigens* 1999; **54**: 527–530.
- Holopainen P, Arvas M, Sistonen P *et al*: CD28/CTLA4 gene region on chromosome 2q33 confers genetic susceptibility to celiac disease. A linkage and family-based association study. *Tissue Antigens* 1999; **53**: 470–475.
- King AL, Moodie SJ, Fraser JS *et al*: CTLA-4/CD28 gene region is associated with genetic susceptibility to coeliac disease in UK families. *J Med Genet* 2002; **39**: 51–54.
- King AL, Moodie SJ, Fraser JS *et al*: Coeliac disease: investigation of proposed causal variants in the CTLA4 gene region. *Eur J Immunogenet* 2003; **30**: 427–432.
- Mora B, Bonamico M, Indovina P *et al*: CTLA-4 +49 A/G dimorphism in Italian patients with celiac disease. *Hum Immunol* 2003; **64**: 297–301.
- Popat S, Hearle N, Wixey J *et al*: Analysis of the CTLA4 gene in Swedish coeliac disease patients. *Scand J Gastroenterol* 2002; **37**: 28–31.
- Martin-Pagola A, Perez de Nanclares G, Vitoria JC *et al*: No association of CTLA4 gene with celiac disease in the Basque population. *J Pediatr Gastroenterol Nutr* 2003; **37**: 142–145.
- Amundsen SS, Naluai AT, Ascher H *et al*: Genetic analysis of the CD28/CTLA4/ICOS (CELIAC3) region in coeliac disease. *Tissue Antigens* 2004; **64**: 593–599.
- Haimila K, Smedberg T, Mustalahti K, Maki M, Partanen J, Holopainen P: Genetic association of coeliac disease susceptibility to polymorphisms in the ICOS gene on chromosome 2q33. *Genes Immun* 2004; **5**: 85–92.
- Shuai K, Liu B: Regulation of JAK-STAT signalling in the immune system. *Nat Rev Immunol* 2003; **3**: 900–911.
- Jacobsen M, Schweer D, Ziegler A *et al*: A point mutation in PTPRC is associated with the development of multiple sclerosis. *Nat Genet* 2000; **26**: 495–499.