

## Appendix 2

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**Table 1** Mutations found at a frequency higher than 1% in Europe<sup>5-7</sup>

Country										
Albania	dF508 (70.0)									
Austria	dF508 (63.7)	G542X (2.1)	R1162X (1.9)	G551D (1.1)						
Belarus	dF508 (63.0)	N1303K (2.7)	G542X (2.1)	W1282X (1.7)						
Belgium	dF508 (75.5)	N1303K (2.9)	G542X (2.7)	W1282X (1.5)	S1251N (1.3)	1717-1G>A (1.1)				
Bulgaria	dF508 (61.0)	N1303K (409)	G542X (3.6)	R347P (2.0)	1677delTA (2.0)	R1070Q (1.5)	W1282X (1.3)	G1244V+S912L (1.0)		
Croatia	dF508 (64.5)	N1303K (3.6)	G542X (3.3)	G551D (1.1)						
Cyprus	dF508 (46.7)	L346P (16.7)	1677delTA (6.7)							
Czechia	dF508 (71.6)	G551D (4.0)	N1303K (3.0)	G542X (2.2)	1898+1G>A (2.0)	2143delT (1.2)	CFTRdele2,3(21kb) (4.6)			
Denmark	dF508 (87.2)	394delTT (1.9)								
Estonia	dF508 (54.0)	394delTT (15.0)								
Finland	dF508 (46.2)	394delTT (28.8)								
France	dF508 (66.8)	G542X (3.1)	1717-1G>A (1.6)	N1303K (1.4)						
Germany	dF508 (73.2)	R553X (2.7)	R347P (1.3)	G551D (1.3)	N1303K (1.2)	G542X (1.2)	3849+10kbC>T (1.2)	CFTRdele2,3(21kb) (1.5)		
Greece	dF508 (52.2)	621+1G>T (4.5)	G542X (3.9)	N1303K (3.3)	2183AA>G (1.8)	2789+5G>A (1.8)	E822K (1.6)	R117H (1.2)	R344W (1.2) 3272-26A>G (1.0) R1158X (1.0) G85E (1.0)	
Hungary	dF508 (54.7)	G542X (2.2)								
Ireland	dF508 (72.7)	G551D (6.9)								
Israel	dF508 (32.2)	W1282X (36.2)	G542X (5.4)	3849+10kbC>T (4.6)	405+1G>A (3.8)	N1303K (3.0)	Q359K+T360K (1.9)	S549R (1.1)		
Italy	dF508 (51.1)	G542X (4.8)								
Latvia	dF508 (54.2)	3849+10kbC>T (12.5)	N1303K (8.3)	W1282X (4.2)						
Lithuania	dF508 (30.9)	R553X (4.8)								
Macedonia	dF508 (56.0)	G542X (3.3)								
North Africa	dF508 (32.0)	N1303K (10.2)	W1282X (8.2)	711+1G>T (7.5)	G542X (4.8)	R1162X (2.7)	L227R (1.4)	S549R (1.4)	S549N (1.4) S549I (1.4) G551D (1.4) S945L (1.4)	
Northern Ireland	dF508 (68.0)	G551D (5.1)								
Norway	dF508 (66.7)	394delTT (4.2)								
Poland	dF508 (52.9)	3849+10kbC>T (2.6)	G542X (2.5)	N1303K (1.7)	1717-1G>A (1.7)	R553X (1.0)	CFTdele2,3(21kb) (2.0)			
Portugal	dF508 (52.3)	R1066C (3.5)								
Romania	dF508 (27.0)									
Russia P	dF508 (45.0)	W1282X (2.1)	N1303K (1.6)	1677delTA (1.6)	2143delT (1.0)	CFTRdele2,3(21kb) (7.5)				
Russia M	dF508 (54.5)	N1303K (2.6)	2143delT (2.0)	2184insA (2.0)	W1282X (2.0)	G542X (1.8)	3849+10kbC>T (1.8)	CFTRdele2,3(21kb) (8.4)		
Slovakia	dF508 (59.4)	G542X (5.6)								
Spain	dF508 (54.4)	G542X (7.7)	N1303K (2.5)	1811+1.6kbA>G (1.5)	R1162X (1.3)	712-1G>T (1.1)	1609delCA (1.0)			
Sweden	dF508 (73.3)	394delTT (9.7)								
Switzerland	dF508 (43.2)	R553X (24.2)								
The Netherlands	dF508 (74.4)	A455E (3.3)								
Turkey	dF508 (34.8)	N1303K (6.4)	1677delTA (2.8)	E92X (2.8)	R347H (2.8)	G542X (2.8)	K68N (1.4)	2043delG (1.4)	2183AA>G (1.4) 2789+5G>A (1.4)	
Ukraine	dF508 (50.0)									
United Kingdom	dF508 (75.3)	G551D (3.1)								
Yugoslavia	dF508 (66.3)	G542X (5.3)								

Sources: Deltas C., Doerk T., Graham C., Macek M. Jr., Pacheco. P. (Personal communication, January 1999).

**Table 2** Overview of total sensitivity for mutations found at a frequency higher than 1% (January 1999)

Country	Total sensitivity %	No. mutations	% mutations not found
Albania	70	1	28.1
Austria	68.8	4	35
Belarus	69.5	4	29.5
Belgium	85	6	8.4
Bulgaria	77.3	8	14.5
Croatia	72.5	4	25
Cyprus	90	8	10
Czechia	88.6	7	4.9
Denmark	90.1	3	4
Estonia	69.4	2	13.5
Finland	75	2	21.2
France	72.9	4	17.5
Germany	83.6	8	16.4
Greece	74.5	12	13.7
Hungary	57.9	3	41.1
Ireland	82.6	4	13.9
Israel	88.2	8	8.4
Italy	70.3	8	22.7
Latvia	79.2	4	20.8
Lithuania	38.1	3	66
Macedonia	68.3	7	50
North Africa	73.8	12	24.5
Northern Ireland	86.2	7	13.8
Norway	75.1	4	23.8
Poland	64.4	7	31.1
Portugal	64.8	7	27.7
Romania	27	1	57.3
Russia (sint Petersburg)	58.8	6	52.8
Russia (Moscow)	75.1	8	
Slovakia	77.7	6	21
Spain	69.5	7	17.9
Sweden	89.6	5	7.3
Switzerland	89.5	6	9.5
The Netherlands	81.7	5	13.5
Turkey	58	10	41.1
Ukraine	50	1	49.1
United Kingdom	80.1	3	13.9
Yugoslavia	71.6	2	26.4

**Table 3** Intragenic polymorphisms in the *CFTR* gene

Polymorphic locus	Reference
IVS6a-GATT	Dörk <i>et al.</i> <sup>45</sup>
1001+11C→T	Cuppens <i>et al.</i> <sup>46</sup>
IVS8-CA	Morral <i>et al.</i> <sup>47</sup>
Tn (5T-7T-9T)	Cuppens <i>et al.</i> <sup>46</sup>
TUB9 (intron 9 1525-61 A or G)	Dörk <i>et al.</i> <sup>45</sup>
M470V	Dörk <i>et al.</i> <sup>45</sup> , Cuppens <i>et al.</i> <sup>46</sup>
T854T	Dörk <i>et al.</i> <sup>45</sup> , Cuppens <i>et al.</i> <sup>46</sup>
IVS17B-TA	Morral <i>et al.</i> <sup>47</sup>
IVS17B-CA	Morral <i>et al.</i> <sup>47</sup>
TUB18 (intron 18 3601-65 A or C)	Dörk <i>et al.</i> <sup>45</sup>
TUB20 (intron 20 4006-200 A or G)	Dörk <i>et al.</i> <sup>45</sup>
Q1463Q	Cuppens <i>et al.</i> <sup>46</sup>

**Table 4** Cost comparative study for the most frequently used *CFTR* mutation detection methods (January 1999)

Mutation detection method	Time scheme			Price of material and supplies for analysing 5 samples	No. mutations
	Working time	Waiting time	Total time		
INNO-LiPA CF2	40min–2h	6h–8h	6h40min–10h	157.3–200 Euro	8 mutations
Elucigene CF12	30min–2h	5h30min–6h	6h–8h	157.3–200 Euro	12 mutations
OLA cystic fibrosis assay	1h30min–3h	4h30min–10h30min	7h–12h30min	237.5–275 Euro	31 mutations
Restriction enzyme analysis	1h15min–2h	5h–10h	7h–13h	16.2–21.3 Euro	1 mutation
Heteroduplex analysis	30min–1h	6h–8h	7h–12h15min	10–12.5 Euro	1 or 2 mutations
DGGE	1h30min–2h	6h–20h	8h–22h	20–22.5 Euro	1 exon or multiplex
SSCP	2h30min–3h	10h–22h	12h30min–25h	16.2–18.7 Euro	1 exon

These results were obtained from the comparative study of the actual cost and hands-on time for different CF mutation detection techniques. The data obtained from the workshop on DNA extraction and CF mutation detection technique held in Leuven, September 1996 were compared with the information obtained from a detailed questionnaire which was sent to laboratories which participate in the European Quality control trial 1998 (except for the OLA kit).

For each technique, the total working time and the costs were evaluated. The total time for the different techniques was divided into the working time i.e. the time needed for one person to do the practical work, and the waiting time, i.e. the time when the samples are in a machine or during a chemical reaction without any handling by a person. The costs for the *CFTR* mutation detection techniques were costs for material and supplies. Material and supplies were valued on the basis of replacement prices (Western European prices).

**Table 5** Features of the most common CF mutation detection techniques (January 1999)

<i>Mutation specific methods</i>	
Heteroduplex Analysis	Useful for small deletions/insertions Mostly used for dF508/dI507 Other insertions/deletions eg 394delTT, 1461insAGAT Visualisation with either ethidium bromide or silver staining Cheap, easy to use
Restriction Enzyme Analysis	Ideally used for confirmation, verification or mutations detected by other methods Can be used for local specific mutation Testing of relatives for 'family' mutations Used in restriction generating PCR Specificity not absolute Relatively expensive
ARMS	Allele-specific PCR, useful for isolated (local specific) mutations Result may rely on a absence of PCR product Can be useful in determining maternal contamination May be applied to pooled samples, in a research setting only! Difficulty in designing primers Cheap once primers have been designed
RG PCR	Useful for isolated (local specific) mutations May be useful for certain applications
Reverse/Normal Dot Blots	Developed according to local population frequencies Useful for large numbers of samples Visualisation/labelling methods Difficult to establish initially May require radioactivity
INNO-LiPA	See Table 6 for mutations tested INNO-LiPA <i>CFTR</i> 12 and INNO-LiPA <i>CFTR</i> 17+Tn (latter includes polyT) INNO-LiPA will distinguish heterozygotes (Wild-type probes included for every mutation detected) No special equipment required, except for a shaking waterbath More robust with respect to methodology High specificity Sensitivity generally high (European-wide) but not locally variable Relatively expensive (although not per mutation) Labour costs reduced Automatable with the auto-LiPA
Elucigene	See Table 6 for mutations tested Elucigene does not generate normal alleles for mutations except dF508 More robust with respect to methodology Elucigene may be sensitive to DNA sample type High specificity Sensitivity generally high (European-wide) but not locally variable Relatively expensive (although not per mutation) Labour costs reduced
OLA	See Table 6 for mutation tested OLA will distinguish heterozygotes Requirement fo expensive equipment ABI 310 or 377 More robust with respect to methodology High specificity Sensitivity generally high (European-wide) but not locally variable Relatively expensive (although not per mutation) Labour costs reduced
<i>Generic methods</i>	
SSCP DGGE	Cheap, once established Difficult to set up (primer design) High pick-up rate, especially when multiplexed
Sequencing	For absolute confirmation of new findings Exclusion of polymorphisms
<i>Polymorphisms</i>	
For Prenatal Diagnosis where one or no mutations have been found For assessment of maternal cell contamination PolyT interpretation – TG <sub>m</sub> For confirmation of zygosity Exclusion of uniparental disomy	

**Table 6** Mutations detected by commercial kits  
(November 1999)

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INNO-LIPA	
CF2:	dF508, dI507, G542X, 1717-1G>A, G551D, R553X, W128X, N1303K
CFTR12:	dF508, dI507, G542X, 1717-1G>A, G551D, R553X, W1282X, N1303K, S1251N, R560T, 3905insT, Q552X
CFTR17+Tn:	394delTT, G85E, 621+1G>T, R117H, 1078delT, R347P, R334W, E60X, 2183AA>G, 2184delA, 711+5G>A, 2789+5G to A, R1162X, 3659delC, 3849+10kbC>T, 2143delT, A455E, (5T/7T/9T)
Elucigene	
CF4:	dF508, 542X, G551D, 621+1G>T
C12	dF508, G542X, G551D, N1303K, W1282X, 1717-1G>A, R553X, 621+1G>T, R117H, R1162X, 3849+10kbC>T, R334W
CF20:	1717-1G>A, G542X, W1282X, N1303K, dF508, 3849+10kbC>T, 621+1G>T, R553X, G551D, R117H, R1160X, R334W, A455E, 2183AA>G, 3659delC, 1078delT, dI507, R345P, S1251N, E60X
CF Poly-T	5T/7T/9T
OLA	
CF OLA assay:	dF508, F508C, dI507, Q493X, V520F, 1717-1G>A, G542X, G551D, R553X, R560T, S549R, S549N, 3849+10kbC>T, 3849+4A>G, R1162X, 3659delC, W1282X, 3905insT, N1303K, G85E, 621+1G>T, R117H, Y122X, 711+1G>T, 1078delT, R347P, R347H, R334W, A455E, 1898+1G>A, 2183AA>G, 2789+5G>A

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