Pseudomonas aeruginosa septicaemia from an oral source

M. A. Gosney, A. J. Preston, J. Corkhill, B. Millns, and M. V. Martin, 5

Oral colonisation with aerobic Gram-negative bacilli (AGNB) is abnormal and usually indicates a medically compromised state in the host. It has been postulated that oral colonisation with AGNB may predispose a patient to serious systemic infection, but proof of this assertion is lacking. This report describes an elderly patient who had oral colonisation of *Pseudomonas aeruginosa* and developed septicaemia from an identical strain of this bacterium.

An 84-year-old female, with chronic obstructive pulmonary disease, was admitted to hospital with an acute right lobar pneumonia. Sputum samples grew mixed respiratory flora but no AGNB. She was treated with 1 g daily intravenous ceftriaxone and responded well and, after 6 days she was discharged. Within 24 hours, she was readmitted with a high fever and signs of septicaemia. Blood and oral cultures grew pure growths of Ps aeruginosa (the patient did not wear dentures); three further sputum samples grew no Ps aeruginosa. She was again treated with 1 g daily intravenous ceftriaxone for 6 days and recovered. Her mouth swabs yielded heavy growths of Ps aeruginosa and she was selectively decontaminated with oral nystatin pastilles (100,000 units), polymyxie E (2 mg) and tobramycin (18.8 mg) four times daily for five days. This selective decontamination regime was obtained from the Pharmacy, Glasgow Royal Infirmary, Glasgow where it is routinely used. The patient has had no further episodes of septicaemia. On both admissions, sputum and urine samples were negative for AGNB.

Chromosomal DNA from a control *Ps aeruginosa* (NCTC 10662) and the oral and blood isolates were compared after endonuclease digestion, using pulsed-field gel electrophoresis (PFGE). DNA was digested with both Spe 1 and Xba 1 restriction endonuclease and the fragments separated by PFGE. The fragments were stained with ethidium bromide and compared. The two strains were found to be indistinguishable (Fig. 1).

This patient had her oral cavity sampled as part of a study (for which she had given informed consent) looking at the incidence of oral AGNB in elderly patients admitted to hospital with acute illness.1 The fact that she developed a Ps aeruginosa septicaemia with concomitant oral carriage of the same microorganism allowed the two cultures to be compared. Indistinguishable strains of Ps aeruginosa were isolated from her oral cavity and blood. It is therefore likely that the mouth flora was the source of her septicaemia. No report was found in the literature describing the use of DNA restriction fragment length polymorphisms to compare oral and systemic strains of *Ps aeruginosa*. This report has additionally shown that the mouth may be a source of systemic infection.

Comment

The patient's septicaemia was managed using cephalosporins and she made an uneventful recovery. Her mouth was abnormally colonised by *Ps aeruginosa*² as this bacteria was consistently found on every occasion when the mouth was sampled prior to decontamination (results not shown). It is therefore probable that the mouth was the source of this patient's systemic infection because elderly debilitated people are known to be colonised with AGNB.^{3,4} No significant pathology was present in her mouth, but this is often not necessary for spontaneous

In brief

- The mouths of elderly medically compromised patients can become abnormally colonised by aerobic Gram-negative bacilli
- In rare cases this can lead to septicaemia
- The colonisation can be prevented or eliminated by selective oral use of non-absorbable antibiotics (selective decontamination)

¹Senior Lecturer/Honarary Consultant, Department of Geriatric Medicine, ²Lecturer in Restorative Dentistry, Leeds Dental Institute, Clarendon Way, Leeds L52 9LU, ⁴Laboratory Scientist, ⁵Senior Lecturer, Department of Clinical Dental Sciences, ³Senior Laboratory Scientist, Department of Medical Microbiology and Genitourinary Medicine, University of Liverpool, Liverpool L69 3BX REFEREED PAPER Received 01.12.98; accepted 11.02.99

© British Dental Journal 1999; **187**: 639–640

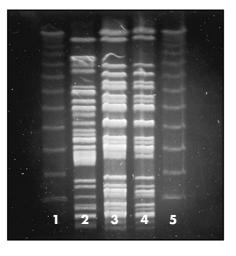


Fig. 1 This figure shows the DNA from the two Ps aeruginosa strains from the mouth and the blood. The DNA has been split into DNA fragments following digestion with the restriction endonucleases (A) SPE 1 and (B) Xba 1. The DNA fragments have been separated by pulsed field gel electrophoresis and can be compared. For control purposes two DNA standards have been included together with DNA from a type strain of Ps aeruginosa. Lanes 1 and 5 contain DNA standards; lane 2 Ps aeruginosa (NCTC 10662); lane 3 Ps aeruginosa (blood isolate) and lane 4 Ps aeruginosa (mouth isolate)

PRACTICE case study

bacterial entry into the blood stream.⁵ Elderly patients do acquire oral Gramnegative bacteria sometimes from nosocomial sources.^{3,4} It is possible that the *Ps* aeruginosa came from the lungs as this patient had chronic obstructive pulmonary disease, but this is unlikely as three sputum samples did not yield any Gram-negative bacilli. To prevent systemic re-infection it was decided to selectively decontaminate her mouth of AGNB using non-absorbable antibiotics: this was successful.⁶ The non-absorbable antibiotics chosen were tobramycin, polymyxin (colistin) and amphotericin. Tobramycin and polymyxin stop the colonisation of the mouth by Gramnegative bacilli and when used together no resistant strains will be selected to either antibiotic.⁶ Amphotericin was also used as this is not absorbed, and this prevents opportunistic overgrowth of yeasts.

Oral carriage of AGNB is abnormal and this case shows that this could lead to serious infection. Selective decontamination should be considered in compromised patients with oral carriage of AGNB to prevent serious systemic infection.

 Preston A J, Gosney M A, Noon, S, Martin M V. Oral flora of elderly patients following acute medical admission. *Gerontology* 1999; 45: 49-52.

- 2 Marsh P D, Martin M V. Oral Microbiology. 4th ed. London: Chapman and Hall, 1999.
- 3 Craven D E, Steger K A, Barat L M, Duncan R A. Nosocomial pneumonia: epidemiology and infection control. *Intensive Care Med* 1992; 18: S3-S9.
- 4 Klastersky J. Nosocomial infections due to Gram-negative bacilli in compromised hosts: Considerations for prevention and therapy. *Rev Inf Dis* 1985; 7: S552-S558.
- 5 Shanson D C. Antibiotic prophylaxis of infective endocarditis in the United Kingdom and Europe. *J Antimicrob Ag Chemotherap* 1987; 8 Suppl. A: 117-121.
- 6 Baxby D, Van Saene H K F, Stoutenbeek C P, Zandstra D F. Selective decontamination of the digestive tract: 13 years on, what is and what is not. *Intensive Care Med* 1996; 22: 699-706.

Submitting illustrations to the BDJ

Authors submitting manuscripts for publication in the BDJ are reminded that two copies of all illustrations must be supplied. This will assist in speeding up the refereeing process — manuscripts and illustrations are always sent to referees and a set of illustrations is required at the editorial office for reference. If you choose to submit 35-mm transparencies then one set of these and one set of prints (colour or black & white) will be acceptable.

We strongly advise authors to keep copies of all illustrations submitted in case letters are lost or damaged in transit. Please do not send glass-mounted transparencies — even when securely wrapped the glass will often arrive cracked or shattered, resulting in damage to the surface of the slide which usually renders it unsuitable for reproduction.