12. vom Baur, E. et al. *EMBO J.* **15**, 110–124 (1995).
13. Cavaillès, V. et al. *EMBO J.* **14**, 3741–3751 (1995).
14. Halachmi, S. et al. *Science* **264**, 1455–1458 (1994).
15. Le Douarin, B. et al. *EMBO J.* **14**, 2020–2033 (1995).
16. Schüle, R. & Evans, R. M. *Trends Genet.* **7**, 377–381 (1991).
17. Arias, J. et al. *Nature* **370**, 226–229 (1994).
18. Hill, C. S. & Treisman, R. *Cell* **80**, 199–211 (1995).
19. Schulman, I. G., Chakravarti, D., Juguilon, H., Romo, A. & Evans, R. M. *Proc. Natl Acad. Sci. USA* **92**, 8288–8292 (1995).
20. Parker, D. et al. *Mol. Cell. Biol.* **16**, 694–703 (1996).
21. Forman, B. M., Umesono, K., Chen, J. & Evans, R. M. *Cell* **81**, 541–550 (1995).
22. Forman, B. M. et al. *Cell* **81**, 687–693 (1995).
23. Umesono, K., Murakami, K. K., Thompson, C. C. & Evans, R. M. *Cell* **65**, 1255–1266 (1991).
24. Hollenberg, S. M., Giguère, V., Segui, P. & Evans, R. M. *Cell* **49**, 39–46 (1987).
25. Kamei, Y. et al. *Cell* **85**, 403–414 (1996).
26. Kwok, R. P. S. et al. *Nature* **380**, 642–646 (1996).

ACKNOWLEDGEMENTS. D.C. and V.L. contributed equally to this work. We thank members of R.M.E.'s laboratory for reagents and for critically reading the manuscript, and T.-P. Yao, D. Livingston and M. Brown for communication of unpublished results. D.C. is supported by a postdoctoral fellowship from the Jane Coffin Childs Memorial Fund for Medical Research. V.L. is supported by a post-doctoral fellowship from the American Cancer Society. I.G.S. is supported by a postdoctoral fellowship from the California division of the American Cancer Society and funds from the J. Aron foundation. R.M.E. is an Investigator of the Howard Hughes Medical Institute at the Salk Institute. This work was supported by grants from the NIH (R.M.E.), The Jane Coffin Childs Memorial Fund for Medical Research (D.C.) and The American Cancer Society (V.L.).

CORRESPONDENCE and requests for materials should be addressed to R.M.E. (e-mail: evans@salk.edu).

CORRECTIONS

Activation of complement by an IgG molecule without a genetic hinge

Ole Henrik Brekke, Terje E. Michaelsen, Randi Sandlin & Inger Sandlie

Nature **363**, 628–630 (1993)

WE reported the construction of a chimaeric mouse–human mutant IgG3 molecule, HM-1 (IgG₃231Cys232), with all hinge exons deleted, but with an introduced cysteine following Ala 231 encoded by the C_H2 exon. However, we have recently noted that the mutant that was tested had four amino acids, namely Ala-Ala-Cys-Ala, introduced after Ala 231. This mutant was also produced in the laboratory, and at some point following *in vitro* mutagenesis and sequencing there had been a switching of tubes. Our original conclusion, namely that the presence of neither a long upper hinge nor a core hinge is necessary for complement-mediated lysis (CML), is still valid. IgG₃231AlaAlaCysAla232 has an upper hinge of three amino acids, and the core hinge is substituted with a single cysteine (see Table 1 below).

IgG₃231Cys232 has recently been tested on non-reducing SDS–PAGE and found to contain mostly H–L half-molecules. It has an upper hinge of one amino acid, the core hinge is substituted with a single cysteine, and the lower hinge lacks an alanine compared to the wild-type sequence. Obviously, it lacks residues necessary for effective disulphide-bond formation. IgG₃231Cys232 has some activity in CML at high antigen density, but no activity at low antigen density. □

CORRECTION

A comparative embryological study of two ornithischian dinosaurs

J. R. Horner & D. B. Weishampel

Nature **332**, 256–257 (1988)

IN this Letter we erroneously referred an embryo in an egg to the ornithopod dinosaur *Orodromeus makelai*. Further preparation of the embryo has revealed its identity to be that of the theropod dinosaur *Troodon cf. formosus*.

The unusual teeth of *Troodon*, possessing cylindrical roots similar to ornithischians, exacerbated the identity problem in these embryos and added to an already long history of confusion¹. Additional preparation has revealed that the dentary union lacks a prementary, precluding the specimen from the Ornithischia. Shape of the humerus and configuration of the metatarsals are identical with corresponding adult and juvenile elements of *Troodon cf. formosus* (in collections of the Museum of the Rockies, Montana State University, Bozeman, Montana, USA). Absence of a fourth trochanter on the femora, originally interpreted as not having ossified until post-eclosion activity, is instead indicative of its theropod affinities. □

1. Currie, P. J. *Vertebr. Paleontol.* **7**, 72–81 (1987).

Inhibition of an inwardly rectifying K⁺ channel by G-protein α -subunits

Wolfgang Schreibmayer, Carmen W. Dessauer, Dmitry Vorobiov, Alfred G. Gilman, Henry A. Lester, Norman Davidson & Nathan Dascal

Nature **380**, 624–627 (1996)

IN the Correspondence and materials section of this letter, which lists the nucleotide positions of antisense oligodeoxynucleotides (ODNs) used in this work, the nucleotide positions of anti-G_{q11} ODN were 546–517 (and not 918–891 as published), and for anti-G_{q13} ODN they were 335–306 (and not 366–335). The rest are correct. □

TABLE 1 Amino-acid sequences of the hinges of human wild-type and hinge-mutant IgG3 molecules

	Upper hinge	Genetic hinge Middle hinge	Lower hinge	s-s	CML
IgG ₃	216 ELKTPGLGDTTHT	CPRCP (EPKSCDTPPPCPRCP) ₃	231 APELLGGP	+	+
IgG ₃ 231Cys232	A	C	PELLGGP	–	–(+)
IgG ₃ 231AlaAlaCysAla232	AAA	C	APELLGGP	+	+
m0			APELLGGP	–	–

CML activity and the presence of covalently closed H₂L₂ molecules (s-s) are indicated as plus and minus signs, respectively. m0, Hinge-deleted IgG3.