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Breast carcinoma with a high histologic grade is highly invasive and metastatic. One reason for its irregular morphology is the formation of excessive protrusions due to assemblages of branched actin filament networks. In mammalian cells, the actin-related protein 2 and 3 (Arp2/3) complex initiates actin assembly to form lamellipodial protrusions by binding to the Wiskott-Aldrich syndrome (WASP)/WASP family verprolinehomologous protein2 (WAVE2), a member of the WASP protein family. In this study, the localization Arp2 and WAVE2 in breast carcinoma was investigated to clarify whether coexpression of the two proteins is associated with histologic grade or patient outcome. Immunohistochemical staining of Arp2 and WAVE2 was performed on mirror specimens of 197 breast carcinomas, and the association between coexpression of the two proteins and clinicopathologic factors was examined. Kaplan-Meier disease-free survival and overall survival curves were analyzed to determine the prognostic significance of Arp2 and WAVE2 coexpression in breast carcinoma. Coexpression of Arp2 and WAVE2 was detected in 64 (36%) of 179 invasive ductal carcinomas and in 2 (11%) of 18 ductal carcinomas in situ, but was not detected in any adjacent non-cancerous tissue. The proportion of cancer cells expressing both Arp2 and WAVE2 was significantly higher in cases with high histologic grade (P < 0.0001), and cases with lymph node metastasis (P = 0.0150). The patients whose cancer cells showed such coexpression had shorter disease-free (P=0.0002) and overall survival (P=0.0122) than patients whose cancer cells expressed only one or none of Arp2 and WAVE2. Multivariate Cox regression analysis revealed that coexpression of Arp2 and WAVE2 is an independent factor for both tumor recurrence (P = 0.0308) and death (P = 0.0455). These results indicate that coexpression of Arp2 and WAVE2 is a significant prognostic factor that is closely associated with aggressive morphology of invasive ductal carcinoma of the breast. Modern Pathology (2007) 20, 339-343. doi:10.1038/modpathol.3800741; published online 2 February 2007

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Actin is one of the most abundant proteins in eukaryotes, being a component of the cytoskeleton that determines and maintains cell shape. Eukaryotic cells can change their shape dramatically and migrate as a result of actin polymerization and/or depolymerization. Cell migration is initiated by plasma membrane protrusions, in the form of filopodia and lamellipodia, which are also known as invadopodia in the case of cancer cells. Among the variously shaped membrane protrusions, flattened lamellipodia are likely to play the most important role in directional movement.¹

Lamellipodia are composed of dense networks of actin filaments that are branched at about 70° . The

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actin-related protein 2 and 3 complex (Arp2/3 complex) is present at the corners of branched actin filaments, and is indispensable for lamellipodium formation.^{2,3} The Arp2/3 complex rapidly catalyzes actin filament nucleation and elongation, and is also essential for efficient remodeling of actin filaments by ATP hydrolysis.^{4–6}

The Arp2/3 complex is activated by binding to the Wiskott–Aldrich syndrome protein (WASP) family proteins. There are five WASP family proteins; WASP, neural WASP (N-WASP), and WASP family verprolin-homologous protein (WAVE)1–3, known to be a suppressor of cAMP-receptors (SCAR1-3). They are very potent cellular nucleating factors, and share a common C-terminal verprolin-homology-connecting acidic (VCA) domain that is responsible for Arp2/3 complex activation. However, WASP and WAVE proteins differ in both the signaling inputs they receive and their cellular outputs, which include adhesion, endocytosis, trafficking, and migration.

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Among the WASP family proteins, WAVE2 has a central role in the formation of lamellipodia and the initiation of amoeboid movement.^{7,8} Interaction of the Arp2/3 complex and WAVE2 occurs at the leading edge downstream in the signaling pathway responsible for directional movements in response to various stimuli such as epidermal growth factor (EGF) and platelet-derived growth factor (PDGF).^{7,9} Activation of these receptors generates phosphatidylinositol^{3–5} triphosphate (PIP3), which in turn activates Rac on the cell membrane. WAVE2 is a PIP3-binding protein and can regulate activation of the Arp2/3 complex from the signals of PIP3 and activated Rac.¹⁰

The majority of WAVE2 in the cell is complexed with Abl-interactor-1 (Abi1), Nck-associated protein-1 (Nap1), a small protein HSPC300, and p53inducible protein-121 (PIR121)/Sra1.¹¹ Although the activity of the WAVE2 complex has yet to be examined, it is considered to be strictly regulated by other molecules that act as different signaling inputs regulating actin dynamics.

Thus, the binding of the Arp2/3 complex and WAVE2 is a final intracellular signal that initiates actin polymerization for the formation of lamellipodial protrusions. The signal is considered to be integrated by various signaling inputs. In order to clarify the clinical significance of this cellular signal, we previously examined the coexpression of Arp2 and WAVE2 in human lung adenocarcinoma by immunostaining using mirror specimens, and the coexpression was found to be correlated with poorer patient outcome.¹² In the present study, immunostaining of Arp2 and WAVE2 was performed in specimens of breast cancer and the correlation of coexpression of these proteins with histologic grade and patient outcome was examined.

Materials and methods

Study Subjects

The subjects of this study were 197 Japanese women who underwent modified radical mastectomy for breast cancer at the Tokyo Medical University Hospital between 1988 and 2002. The median follow-up period for all patients was 7.7 years, and that of living patients was 8.6 years (range, 3.5–13.6 years). As adjuvant chemotherapy between 1988 and 1989, 63 patients of invasive ductal carcinoma received orally a combination of uracil and tegafur, often referred to as UFT, (250 mg of tegafur per square meter of body-surface area per day) for two years after the surgery. From 1990 to 2002 UFT plus tamoxifen (20 mg/day) were given to the patients with ER-positive invasive ductal carcinoma and only oral UFT for those with ER-negative invasive ductal carcinoma. Other 18 patients of ductal carcinoma *in situ* did not receive any chemotherapy. Blocks of formalin-fixed, paraffin-embedded tissue from these patients were retrieved from the Department of Diagnostic Pathology, and clinicopathological data, such as age at the time of diagnosis, tumor size, and axillary lymph node status, were obtained from the medical records. To confirm the presence of sufficient tumor tissue and to ensure contemporary uniform grading, paraffin-embedded tissue blocks were sectioned, stained with hematoxylin–eosin, reviewed for the presence of tumor, and histological grade was determined by a single pathologist, as outlined previously.¹³ Informed consent was obtained from each patient.

Immunohistochemistry

One or two representative blocks of each tumor were selected for immunohistochemical analysis of Arp2 and WAVE2. From all paraffin blocks, $4-\mu$ m-thick sections were cut and placed on silane-coated slides. In order to clarify the localization of Arp2 and WAVE2, we prepared mirror sections from a pair of consecutive specimens. These were placed on slides with the common cross-section turned upwards, so that they would share the same cutting surface. The features in such specimens appear reversed, and the specimens are therefore known as mirror specimens. Immunohistochemical procedure was described in our previous paper.¹² Briefly, antigen retrieval was achieved by heating the slides in an autoclave at 110°C for 10 min in 1 mM-EDTA (pH8.0). After cooling, pairs of mirror specimens were incubated overnight at room temperature with the primary antibody, goat polyclonal anti-human Arp2 antibody, at a dilution of 1:50 (k-15, Santa Cruz Biotechnology, Santa Cruz, CA, USA) and goat polyclonal anti-human WAVE2 antibody at a dilution of 1:50 (c-14, Santa Cruz Biotechnology), respectively.

If more than 5% of cancer cells expressed both Arp2 and WAVE2 in their cytoplasm, Arp2 and WAVE2 were judged to be positive for coexpression in the tumor.

Statistical Analysis

Correlations between the coexpression of Arp2 and WAVE2 and clinicopathologic factors were evaluated by χ^2 tests, at a significance level of P < 0.05. Univariate and multivariate survival analyses were performed to evaluate the prognostic significance of coexpression and the effect of other factors, such as age, tumor size, axillary lymph node status, and histologic grade, on the clinical outcome.

Results

Immunohistochemistry

Coexpression of Arp2 and WAVE2 was detected in 64 (36%) of 179 invasive ductal carcinomas and in 2 (11%) of 18 ductal carcinomas *in situ*, and there was

Factors	Cases	Coexpression (%)	P-value
Tumor size			
T1	59	16 (27)	
T2	99	41 (41)	
Т3	21	7 (33)	NS
State of axillary ly	mph node		
Metastasis	68	32 (47)	
No metastasis	102	29 (28)	0.015
Histologic grade			
Grade 1	25	1 (4)	
Grade 2	93	27 (29)	
Grade 3	61	36 (59)	< 0.0001
Invasive type			
DCIS	18	2 (11)	
IDC	179	64 (36)	0.0333

a significant difference in the frequency of coexpression between invasive carcinoma and noninvasive carcinoma (Table 1, P = 0.0333). The cancer cells showed coexpression most frequently at the invading front (Figure 1). In most cases judged positive for the coexpression, number of cancer cells expressing both Arp2 and WAVE2 were no more that 10% of whole cancer cells. Either Arp2 or WAVE2 was focally stained in normal epithelial cells or stromal cells, and no normal cells showed distinct coexpression of Arp2 and WAVE2 except for macrophages.

Association between Coexpression and Clinicopathologic Factors

Among 61 invasive ductal carcinomas with a high histologic grade (grade 3), 36 (59%) were coexpression-positive (Table 1). However, 27 (29%) of 93 grade 2 and 1 (4%) of 25 grade 1 tumors were judged to be positive for localization. Thus the coexpression was strongly associated with a high histologic grade (P<0.0001).

Cancer cells with positive coexpression frequently metastasized to the axillary lymph nodes (P=0.0150). There was no correlation between coexpression and T factor.

Table 2 shows the association between coexpression and established biologic markers of breast cancer. Coexpression was strongly associated with positive expression of Her2/*neu* (P<0.0001). It was inversely correlated with both estrogen (P=0.0027) and progesterone receptors (P=0.0025). Nuclear p53 immunoreactivity was not correlated with coexpression in this study.

Association between Coexpression and Patient Outcome

Figure 2 shows the Kaplan–Meier disease-free survival (DFS) curves of patients with tumors



Figure 1 A case of invasive ductal carcinoma in which mirror sections were stained for Arp2 and WAVE2. Specimens A was stained for Arp2, and the corresponding mirror sections B was stained for WAVE2. Staining of both Arp2 and WAVE2 are present homogeneously in the cytoplasm of the same cancer cells.

showing positive coexpression and those negative for coexpression. Their DFS curves showed a significant difference as estimated by the log-rank test (P=0.0002). Similarly, histologic grade and axillary lymph node status were significant prognostic factors for DFS. Among them, multivariate Cox regression analysis demonstrated that coexpression of Arp2 and WAVE2 (P=0.0002) and axillary lymph node status had independent prognostic value for DFS (P<0.0155) (Table 3).

There was also a significant difference in the overall survival curves between the positive and negative groups (Figure 2, P=0.0122), and coexpression was found to be an independent prognostic factor for overall survival by multivariate analysis (Table 4, P=0.0455). Of 63 invasive ductal carcinoma cases between 1988 and 1989 that were given oral UFT as adjuvant chemotherapy, cases with positive coexpression had shorter disease-free (P<0.0001) and overall survival (P=0.0002) than those without coexpression of Arp2 and WAVE2.

Factors	Cases	Coexpression (%)	P-value
Her2/neu			
0 and 1+	132	32 (24)	
2+ and 3+	47	32 (68)	< 0.0001
ER			
Negative	58	30 (52)	
Positive	121	34 (28)	0.0027
PR			
Negative	73	36 (49)	
Positive	106	28 (26)	0.0025
p53			
Negative	119	38 (32)	
Positive	60	26 (43)	0.1408



Figure 2 Kaplan-Meier analysis in the Arp2 and WAVE2 coexpression-positive group and -negative group. (a) Kaplan-Meier DFS curves for 179 patients stratified by an Arp2 and WAVE2 coexpression-positive group and -negative group. (b) Kaplan-Meier overall survival curves for 179 patients in an Arp2 and WAVE2 coexpression-positive group and -negative group. *P*-values were calculated using the log-rank test.

Similarly, 116 patients between 1990 and 2002 with coexpression had shorter disease-free (P=0.0317), however, there was no difference in overall survival (P=0.1832) between two groups.

The physiological role of WAVE2 bound to the Arp2/3 complex in the formation of lamellipodia, which initiate migration, has been established.¹⁴ In the present study, we demonstrated coexpression of Arp2 and WAVE2 in breast cancer tissue. As a preliminary experiment, we prepared cell blocks of breast cancer cell lines such as MCF7, SK-BR-3, and AU-565, and subjected them to Arp2 and WAVE2 immunostaining. The staining pattern of Arp2 and WAVE2 coexpression in these cell lines was predominant at the leading edge, as revealed by double staining and confocal laser scanning microscopy. We considered that such coexpression in breast cancer cells corresponds to binding of WAVE2 to the Arp2/3 complex at the leading edge, and that this is at least partly involved in cancer cell migration that facilitates invasion and metastasis. Our findings that the coexpression was frequently detected at the invading front and was significantly correlated with axillary lymph-node metastasis support the involvement of coexpression in the process of invasion and/ or metastasis.

This study revealed that WAVE2:Arp2/3 coexpression is an independent prognostic factor of both DFS and overall survival, and functionally represents a signal of lamellipodium formation and cell migration. Such coexpression was detected in 59% of carcinomas with high histologic grade. Excessive formation of lamellipodial protrusions is suggested to induce an aggressive morphology and active movement of cancer cells. However, ductal carcinomas with high histologic grade also contain cancer cells with a round shape. Other signals of cellular migration such as the Rho/ROCK pathway and expression of cortactin may be enhanced in such cancer cells.¹⁵⁻¹⁸ If it were possible to eliminate both signals of cellular migration, then the ability of cancer for invasion might become easier to understand.

WAVE2:Arp2/3 coexpression is considered to be an intracellular signal occurring downstream of tyrosine kinase receptors such as EGF and PDGF. Our data showed that such coexpression is significantly associated with Her2/*neu* overexpression, suggesting that the coexpression is also downstream from Her2/*neu* stimuli. Currently, trastuzumab is a powerful therapeutic agent for advanced breast cancer. WAVE2:Arp2/3 coexpression may eliminate the ability of trastuzumab to suppress invasion and/or metastasis, although further examination to confirm the direct link between Her2/*neu* overexpression and coexpression is necessary.

The coexpression of Arp2 and WAVE2 was detected most frequently at the invading front, and was not present diffusely throughout the cancer tissue. In human breast cancer cell lines, EGF stimulation does not induce WAVE2:Arp2/3 coexpression in all cells but only in a proportion of them. These findings suggest that WAVE2:Arp2/3
 Table 3 Multivariate conditional logistic regression analysis for prediction of tumor recurrence

Factor	Univariate P-value	Multivariate	
		Hazard rate	P-value
Axillary lymph node status Histologic grade, G1 and G2 vs G3	0.0155 0.0089	3.939	0.0472
Arp2-WAVE2, negative vs positive	0.0002	4.665	0.0308

Factor	Univariate P-value	Multivariate	
		Hazard rate	P-value
ER, negative <i>vs</i> positive Arp2-WAVE2, negative <i>vs</i> positive	$0.0469 \\ 0.0122$	2.66	0.0455

coexpression is regulated by other stimuli, for example, cell-to-cell contact, and that cancer cells appear to be enhanced by coexpression because adjacent non-cancerous breast epithelial cells showed no coexpression. Clarification of the function of the WAVE2 complex may be a key to resolving this issue and for finding molecules that could be useful as therapeutic targets.

We propose that coexpression of Arp2 and WAVE2 is a new prognostic factor in breast cancer, which partly reflects cell migration ability. Such coexpression may prove promising for clinical manipulation with a view to future therapy.

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