Intercellular junctions in Ewing sarcoma/ primitive neuroectodermal tumor: additional evidence of epithelial differentiation

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Ewing sarcoma/primitive neuroectodermal tumor (ES/PNET) has recently been shown to frequently express cytokeratins, suggesting partial epithelial differentiation. Older ultrastructural studies have documented primitive cell-cell junctions in ES/PNET, reportedly resembling poorly formed desmosomes. Recently, paraffinreactive antibodies have become available to proteins found in a variety of intercellular junctions indicative of epithelial differentiation, including tight junctions, desmosomes and adherens junctions. We examined intercellular junction protein expression in a large number of genetically confirmed ES/PNET. Formalin-fixed, paraffin-embedded sections from 23 primary and seven recurrent or metastatic cases of genetically confirmed ES/PNET were immunostained for claudin-1 and occludin (tight junction structural proteins), zonula occludens-1 (ZO-1, tight junction linker protein), desmoglein 1/2 (desmosomal adherens protein), desmoplakin (desmosomal structural protein) and E-cadherin (epithelial adherens junction protein), using steam heatinduced epitope retrieval and the Dako Envision system. Cases with >5% positive cells were scored as 'positive'. Normal colonic epithelium and skin served as external positive controls. Claudin-1 was expressed by 19 of 30 specimens (63%), ZO-1 was expressed by 15 of 29 specimens (51%), and occludin was expressed by three of 28 specimens (11%). In 28 specimens all three tight junction markers were evaluable. In all, 15 samples (54%) expressed only one tight junction marker, and 10 samples (36%) expressed two tight junction markers. No case expressed all three tight junction markers. Desmoglein was expressed in one of 30 (3%) samples. Desmoplakin was expressed in two of 28 (7%) samples. E-cadherin was negative in all cases. Our data suggest that many of the previously described cell-cell junctions in ES/PNET are poorly formed tight junctions, given the high frequency of claudin-1 and ZO-1 expression. This may underestimate the true frequency of tight junction protein expression in ES/PNET, as there are at least 20 different claudins and other ZO proteins. These tight junctions are almost certainly abnormal, given the absence of occludin expression in most cases. Desmosomal and adherens junction protein expression was rare to absent. Our findings provide additional evidence that ES/PNET frequently show partial epithelial differentiation.

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Ewing sarcoma/primitive neuroectodermal tumor (ES/PNET) is a primitive bone and soft tissue sarcoma that may occur in both children and adults. Although ES/PNET has traditionally been considered to be of neuroectodermal derivation, it generally does not show evidence of any specific line of differentiation, either by ultrastructural or immunohistochemical methods.^{1–9}

Over the last two decades it has been discovered that over 85% of ES/PNET have a specific and diagnostic translocation, t(11;22)(q24;q12) that results in the formation of an EWS/FLI-1 fusion protein, with a minority of ES/PNET having other specific translocations, including t(21;22), t(17;22), and t(2;22).⁹⁻¹¹ These ES/PNET-specific genetic events can be identified either through traditional cytogenetic methods or with the assistance of

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molecular genetic techniques, such as RT-PCR and FISH, and are undoubtedly critical to its pathogenesis.

Recently, it has been shown that up to 25% of ES/ PNET express cytokeratins, suggestive of partial epithelial differentiation.^{12,13} These findings are intriguing, inasmuch as older ultrastructural studies of ES/PNET have documented primitive intercellular junctions,^{5,7} and a recent study has documented particularly frequent intercellular junction formation in cytokeratin-positive ES/PNET.¹⁴ A single immunohistochemical study has found expression of desmosomal proteins, such as desmoplakin and plakoglobin (α -catenin) in these primitive junctions and suggested that this finding was indicative of at least some epithelial differentiation in ES/PNET.¹⁵

Paraffin-reactive antibodies have lately become available to proteins found in a variety of intercellular junctions, including tight junctions, desmosomes, and adherens junctions. We studied a large number of genetically confirmed cases of ES/PNET with markers to the tight junction structural proteins claudin-1 and occludin, the tight junction linker protein zonula occludens-1 (ZO-1), the desmosomal cadherins desmoglein 1/2, the desmosomal structural protein desmoplakin, and the epithelial cell adherens junction protein E-cadherin, in order to more fully characterize their intercellular junctions.

Materials and methods

Formalin-fixed, paraffin-embedded blocks from 31 specimens (24 primaries, seven recurrences/metastases from these 24 primaries) of ES/PNET were retrieved from the archives of Emory University, the University of Washington Medical Center, and the Cleveland Clinic Foundation. All cases had previously been shown to be positive for the ES/PNET-specific translocations t(11;22) (23 cases) or t(21;22) (one case) by traditional cytogenetic study, RT-PCR or FISH (data not shown). Outcome data were not available.

All cases were immunostained at Emory University with the following commercially available antibodies: claudin-1 (polyclonal, 1:70, Zymed Laboratories); ZO-1 (Clone 1, 1:400, Transduction Laboratories); occludin (Clone OC-3F10, 1:100, Santa Cruz Biotechnology); desmoglein 1/2 (DG 3.10, 1:15, Research Diagnostics); and E-cadherin (ECCD-2, 1:200, Zymed Laboratories). Steam heatinduced epitope retrieval and the Dako Envision system were used. Normal colonic epithelium, skin and endothelial cells served as positive controls. Negative controls consisted of substitution of buffer for the primary antibody. Cases were scored as 'positive' if greater than 5% of cells stained positively. Only membranous staining was scored. This 5% cutoff was arrived at after initial review of

 Table 1
 Immunohistochemical results

Specimen number	Claudin-1	ZO-1	Occludin	Desmoplakin	Desmoglein 1/2	E-Cadherin
1	Negative	Negative	Negative	Negative	Negative	Negative
2	Negative	Negative	Negative	Negative	Negative	Negative
3	Negative	Positive	No tumor	Negative	Negative	Negative
4	No tumor	No tumor	No tumor	Negative	Negative	Negative
5	Positive	No tumor	No tumor	Positive	No tumor	Negative
6	Positive	Negative	Negative	Negative	Negative	Negative
7	Positive	Negative	Negative	Negative	Negative	Negative
8	Positive	Negative	Negative	Negative	Negative	Negative
9	Positive	Negative	Negative	Negative	Negative	Negative
10	Positive	Negative	Negative	Negative	Negative	Negative
11	Positive	Negative	Negative	Negative	Negative	Negative
12	Positive	Positive	Negative	Negative	Negative	Negative
13	Positive	Positive	Negative	Positive	Negative	Negative
14	Positive	Positive	Negative	Negative	Negative	Negative
15	Positive	Negative	Negative	Negative	Negative	Negative
16	Positive	Negative	Negative	Negative	Negative	Negative
17	Negative	Negative	Negative	Negative	Negative	Negative
18a	Negative	Positive	Positive	Negative	Negative	Negative
18b	Negative	Positive	Positive	Negative	Negative	Negative
18c	Negative	Positive	Negative	Negative	Negative	Negative
18d	Positive	Positive	Negative	Positive	Positive	Negative
19	Positive	Positive	Negative	Negative	Negative	Negative
20a	Negative	Positive	Negative	Negative	Negative	Negative
20b	Negative	Positive	Negative	Negative	Negative	Negative
21a	Positive	Positive	Negative	Negative	Negative	Negative
21b	Positive	Negative	Positive	Positive	Negative	Negative
22a	Positive	Positive	Negative	Negative	Negative	Negative
22b	Positive	Negative	Negative	Negative	Negative	Negative
23	Negative	Positive	Negative	Negative	Negative	Negative
24	Positive	Positive	Negative	Positive	Negative	Negative

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the immunostained slides, which showed these cell junction markers to frequently be expressed in a patchy fashion in clusters of cells, rather than diffusely. All case were independently scored by two observers (ANS and ALF) and any differences were resolved by consensus.

Immunostaining for desmoplakin was performed at the University of Virginia (DP 2.17, 1:10, Research Diagnostics Inc.) using microwave oven-induced epitope retrieval and the Dako Envision system. Normal epidermal keratinocytes served as a positive external control.

Results

The immunohistochemical results are detailed in Table 1. In all cases, positive staining for the various proteins was membranous and circumferential (non-polarized). In general, positive cases showed expression of ZO-1 and claudin-1 in > 50% of cells. In contrast, expression of occludin, desmoglein and desmoplakin was almost always confined to scattered clusters of positively staining cells.

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Of the tight junction markers, claudin-1 was expressed by 19 of 30 specimens (63%), ZO-1 was expressed by 15 of 29 specimens (51%), and occludin was expressed by three of 28 specimens (11%). In 28 specimens all three tight junction markers were evaluable. In all, 15 samples (54%) expressed only one tight junction marker, and 10 samples (36%) expressed two tight junction markers. No case expressed all three tight junction markers (Figures 1-3). Desmoglein 1/2 was expressed in one of 30 (3%) cases (Figure 2). Desmoplakin was expressed by two of 28 (7%) of cases; one of these cases (an 'adamantinoma-like' ES/PNET) also expressed ZO-1, claudin-1 and desmoglein 1/2, whereas the other desmoplakinpositive case (a conventional ES/PNET) expressed ZO-1 and claudin-1, but not desmoglein 1/2 or occludin (Figure 4). E-cadherin was negative in all 30 samples tested.

Discussion

Intercellular junctions allow individual cells to remain structurally linked to each other, while



Figure 1 Typical ES/PNET (a) showing strong expression of ZO-1 (b) and claudin-1 (c), but not occludin (d). The absence of occludin expression is suggestive of abnormal tight junction formation.



Figure 2 'Adamantinoma-like' ES/PNET showing pronounced nested growth and peripheral nuclear palisading (a). This case showed strong expression of ZO-1 (b), claudin-1 (c) and desmoglein 1/2 (d). This case also showed focal expression of desmoplakin (not shown).

controlling the diffusion of solutes between adjacent cells. Such mechanical linkages among cells create boundaries and compartments, which are fundamental in multicellular organisms. Several types of intercellular junctions exist including tight junctions (most apically located), adherens junctions and desmosomes (most basally located).

Although Friedman and Gold² did not identify intercellular junctions in their seminal 1968 ultrastructural study of ES/PNET, such junctions were noted shortly thereafter by a number of investigators, including Hou-Jensen *et al*,³ Takayama and Sugawa¹⁶ and Llombart-Bosch *et al*.⁵ In general, these junctions have been described as rudimentary, resembling desmosomes or 'desmosome-like junctions', although tight junction-like structures have also rarely been reported.⁵ Perhaps the most detailed ultrastructural study of intercellular junctions in ES/PNET is that of Navas-Palacios *et al*,⁷ who noted intercellular junctions in 70% of 33 studied ES/ PNET, with well-developed desmosomes and tight junctions noted in only 9% of cases, and primitive junctions of indeterminate type present in over 90% of cases. Most recently, Srivastava *et al*¹⁴ have documented cell–cell junctions of uncertain type in 12 cases of cytokeratin-positive ES/PNET.

The overwhelming majority of studies of ES/ PNET over the last 20 years have concentrated on its immunophenotype and the molecular genetic events underlying its pathogenesis, and relatively little has been added to our understanding of these enigmatic intercellular junctions. The only study we are aware of that has attempted to further characterize the intercellular junctions of ES/PNET is that of Moll et al,¹⁵ who demonstrated at least focal expression of desmoplakin (a desmosomal protein), vinculin (a desmosomal and adherens junction protein) and plakoglobin (a desmosomal protein) in all 11 ES/PNET studied. The desmosomal cadherin, desmoglein, was negative in the three studied cases, and tight junction-related and adherens junction-related proteins were not studied.

In epithelial cells, tight junctions define membrane polarity, form an intercellular seal, and

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Figure 3 Typical ES/PNET (a), negative for ZO-1 (b), but positive for claudin-1 (c) and occludin (d).

regulate paracellular transport of water, solutes and cells.¹⁷ The molecular constituents of tight junctions include integral transmembrane proteins, such as the claudin family, occludin, and junctional adhesion molecule (JAM), and scaffolding or linker proteins, such as the zonula occludens family.¹⁸ Tight junctions also appear to play a critical role in signal transduction through interactions with the actin cytoskeleton.¹⁷ Tight junctions are largely restricted to epithelial cells, although they are also present in endothelium and perineurial cells.¹⁹

The claudins are a family of approximately 20 homologous proteins that help determine tight junction structure and permeability, and which appear to be differentially expressed in tissues, with claudin-1 expression, for example, being relatively widespread among epithelia, and claudin-3 expression being confined to lung and liver epithelia.¹⁸ Occludin, a structural and functional component of the tight junction, has been identified in most epithelial tight junctions, at both the gene and protein level, although its role in assembly and maintenance of tight junctions is still undefined.^{20–22}

As far as is known, all normal tight junctions contain occludin, although there is some experimental evidence to suggest that occludin may not be obligatory for tight junction formation.²⁰ ZO-1 (zonula occludens-1) is a scaffolding protein that links the transmembrane tight junction proteins to cytoplasmic proteins and the actin cytoskeleton.^{23,24} Unlike the claudins and occludin, ZO-1 is not confined to the tight junction and also plays a role in the formation of the adherens junction.^{25,26}. Thus, ZO-1 is expressed in a variety of nonepithelial cell types including Schwann cells, dermal fibroblasts, and astrocytes.^{27–29}

Although Navas-Palacios noted tight junctions in only 9% of ES/PNET by electron microscopy, our data strongly suggests that these junctions are far more common. Specifically, we noted expression of the tight junction-related proteins claudin-1 and ZO-1 in over 50% of samples respectively, with coexpression in nearly 40%, and expression of at least one protein in close to 90% of samples. Although it might be argued that ZO-1 expression is not necessarily indicative of tight junction formation, given its other roles in cell signaling,



Figure 4 Typical ES/PNET (a) with strong expression of desmoplakin (b). This was the only case to show this degree of desmoplakin expression.

claudin expression is thought to be tight junctionspecific. In fact, our study may underestimate the true frequency of tight junction protein expression in ES/PNET, as there are at least 20 different claudins and 3 ZO proteins. Interestingly, occludin was present in only 11% of samples, implying that these tight junctions are abnormal, inasmuch as occludin is thought to be present in all known normal tight junctions. These tight junctions have a circumferential, nonpolarized, pattern of distribution, providing further evidence that they are abnormal. We have recently shown a similarly abnormal distribution of tight junctions in another primitive sarcoma, synovial sarcoma.³⁰

Desmosomes, along with adherens junctions, comprise the major adhesive junctions of epithelial cells, although they are also present in a variety of nonepithelial tissues, including myocardium, meningeal cells, and ovarian granulosa cells.^{15,31,32} Desmosomes are comprised of three domains; an adhesive transmembrane region, an outer dense protein plaque, and a less dense inner plaque.³³ The principal components of the adhesive region are the desmosomal cadherins, desmoglein and desmocollin, each of which have three isoforms, with desmoglein and desmocollin present in all desmosomes, and the other isoforms having more restricted patterns of expression.³³ The outer dense plaque contains plakophilin and plakoglobin, which bind to the major component of the inner plaque, desmoplakin. Desmoplakin is required for the formation of desmosomes and attaches the desmosome to the intermediate filament skeleton.³³

We have been able to partially confirm the prior observations of Moll et al with regards to desmosomal protein expression. We observed desmoplakin expression in only 7% of cases, a lower percentage than previously noted. The lower sensitivity of our study is most likely explained by our use of formalin-fixed, paraffin-embedded tissues, as compared with the frozen tissues used by Moll *et al.*¹⁵ As with occludin, the absence of desmoglein 1/2 in all but one case also suggests that the desmosomes formed by ES/PNET are abnormal, as all normal desmosomes should contain desmoglein 2 (Dr Otmar Huber, personal communication). The only tumor in this series which expressed both desmoplakin and desmoglein 1/2 was an 'adamantinomalike' ES/PNET, which also expressed high molecular weight cytokeratins (data not shown), suggestive of 'complete' epithelial differentiation. There does not appear to be any correlation between expression of cytokeratins and desmosomal proteins, as a significant subset of the desmosomal proteinnegative cases in the present series were known to express low-molecular weight cytokeratins (data not shown).

The adherens junction plays an important role not only in intercellular adhesion, but also in signal transduction to the actin cytoskeleton.³⁴ The transmembrane region of the adherens junction is composed mostly of cadherins, including E-cadherin (epithelia), N-cadherin (neural), R-cadherin (retinal), and P-cadherin (placental).³⁵ The cadherins in turn bind to α -catenin and β -catenin, which attach to actin filaments.^{34,35}

Although expression of E-cadherin has been documented in a single ES/PNET cell culture,³⁶ we are unaware of any other previous study of Ecadherin expression in ES/PNET. We were not able to document E-cadherin expression in any of the 30 samples studied, strongly suggesting that ES/PNET do not form epithelial-type adherens junctions. No previous studies appear to have examined ES/PNET for expression of other cadherins, such as Ncadherin, and this may prove to be an interesting area for future investigation.

In summary, we have found evidence of tight junction protein expression in over 50% of genetically confirmed ES/PNET, with expression of desmosomal proteins in a small number, and no expression of epithelial adherens junction markers. These observations support, we believe, the evolving viewpoint that ES/PNET often show epithelial differentiation, especially when taken in the context of the prior work of Moll and colleagues, the recent observations of cytokeratin expression in a significant number of ES/PNET, and the extremely rare but documented capacity of ES/PNET for complex epithelial differentiation, in the form of so-called 'adamantinoma-like' ES/PNET. Our finding of pan-cytokeratin (AE1/AE3) positivity in 11 of the 24 (46%) cases included in this study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a significant of the percent find in the study (data not a study (data not a study stu

so-called 'adamantinoma-like' ES/PNET. Our finding of pan-cytokeratin (AE1/AE3) positivity in 11 of the 24 (46%) cases included in this study (data not shown) and the recent findings of Srivastava et al¹⁴ of both cytokeratin expression and cell-cell junction formation in 12 cases of ES/PNET also support this concept. The minor differences in junctional protein expression we observed between primary and recurrent/ metastatic tumors is most likely related to sampling variation, although studies of other primary and metastatic ES/PNET are necessary to more fully address this issue. It remains to be seen whether epithelial differentiation in ES/PNET carries any prognostic significance or provides any information about response to different therapeutic protocols.

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