experiments containing GroES yielded very similar data (Fig. $3 d$ ). Thus, $\beta$-actin released from GroEL is recognized and cycled by c-cpn at least as efficiently as $\beta$-actin diluted from denaturant. Similar data were obtained using $\alpha$-tubulin as target protein (Fig. $3 e, f$ ). We conclude that GroEL can form a binary complex with labelled unfolded $\beta$-actin or $\alpha$-tubulin and releases them in an ATP-dependent manner, but cannot aid their proper folding, with or without GroES.

We also found that mt-cpn, like GroEL, cannot help the productive folding of $\beta$-actin or $\alpha$-tubulin (data not shown); however, mt-cpn can cycle these target proteins (although to different extents compared with GroEL) (Fig. $3 g-j$ ). We conclude that actin and tubulin target proteins can both be discharged from GroEL or mt-cpn in the presence of ATP in a spectrum of states that, though recognizable by c-cpn, cannot proceed directly to the native state. Rather, they are confined to an endless cycle of binding to and release from different GroEL or mt-cpn molecules.

To look for differences in the state of chaperonin-bound target proteins, we compared their sensitivity to two different proteases (Fig. 4). In the absence of ATP, $\beta$-actin bound to GroEL, $\mathrm{mt}-\mathrm{cpn}$ or $\mathrm{c}-\mathrm{cpn}$ has a similar low resistance to protease, suggesting that the three chaperonins have the same ability to unfold proteins. However, protease-resistant states do accumulate when $\beta$-actin is cycled by GroEL in the presence of GroES (Fig. 4f). The protease sensitivity of $\beta$-actin cycled by c-cpn cannot be interpreted because of the accumulation of protease-resistant native product, but when $\alpha$ - or $\beta$-tubulin is cycled by c-cpn, protease-resistant states are generated (G.T. et al., manuscript in preparation). Thus, c-cpn and GroEL/GroES produce more native-like spectra of folding intermediates than either GroEL alone or mt-cpn, with or without cpn10.

It has been proposed that chaperonins function by unfolding kinetically trapped intermediates ${ }^{7,8}$. However, they do not function in a manner that mimics unfolding by urea or guanidine, because all three chaperonins can aid the folding of target proteins (even following a single round of interaction) under conditions in which no spontaneous folding would occur upon dilution from denaturant. These considerations and the data presented here imply that a distinct set of folding intermediates is released from different chaperonins. Our data suggest that chaperonin-bound actin or tubulin target proteins are extensively unfolded; however, when cycled by GroEL/GroES, and even more so when cycled by c-cpn, significant higher order structure is acquired. Thus, GroEL, mt-cpn and c-cpn seem to differ not in their ability to unfold or cycle proteins, but in their ability to generate productive folding intermediates.

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## ERRATUM

# Glycogen synthase kinase-3 and dorsoventral patterning in Xenopus embryos 

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In the legends to Figs $2 d$ and $3 b$ of this Article, the shading of histogram bars was incorrectly designated. These should be: white bar, normal embryo; hatched bar, incomplete axis duplication; black bar, complete axis duplication.

## CORRECTIONS

# XLas is a new type of $G$ protein 

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We wish to report a discrepancy in the predicted size of XLas and its apparent size on SDS-PAGE. To express XLas, we transfected PC12 cells with a composite clone assembled from clones CDM8-XL, RACE I. 24 and RACE II. 7 (see Methods in Fig. $2 c$ legend) which contains the open reading frame that runs from the ATG at nucleotides $46-48$ to the CTC at nucleotides 2,5812,583 (Fig. 2c). Surprisingly, this composite clone, which encodes a polypeptide of relative molecular mass 92 K , yields a protein with an apparent mobility of 124 K on SDS-PAGE. Expression in PC12 and CHO cells of the clone CDM8-XL (nucleotides 380-2,956; Fig. 2c), which encodes a polypeptide of relative molecular mass 78 K , yields a protein with an apparent electrophoretic mobility of 94 K , identical to the endogenous XLas of PCl 2 cells. Thus it appears that translation of XLas starts at the ATG at nucleotides 439-441, resulting in a $715-$ amino-acid protein of relative molecular mass 78 K (XL portion, 367 amino acids; relative molecular mass 37 K ) with an anomalous electrophoretic mobility. This does not affect any of the conclusions of our study, except that the N -terminal domain preceding the EPAA repeats (Fig. 2c,d) is shorter than we assumed. The significance of the open reading frame starting at the ATG at nucleotides $46-48$ remains to be investigated.

# An Early Miocene anthropoid skull from the Chilean Andes 

John J. Flynn, André R. Wyss, Reynaldo Charrier \& Carl C. Swisher

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THE captions of parts $b$ and $c$ of Fig. 1 legend in this Letter were transposed as published.


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