sary and sufficient condition for a sequence to fold rapidly in the present model is that the native state is a pronounced energy minimum"² and "the features that depend only on the lower discrete part of the spectrum can be characterized by use of the compact self-avoiding chains alone, neglecting the non-compact conformations"². But this conclusion is contradicted by 11 of the sequences they studied for which lowest-energy conformations are not maximally compact². The true denatured ensemble is generally much larger than the maximally compact ensemble, so neither the first excited state "energy gap" ΔE_{10} , defined by them, nor the temperature, T_x , which they define using only the maximally compact ensemble², is precisely related to true thermodynamic stability. Furthermore, the correlation they observed between their energy gap and folding kinetics is only a weak trend, and the energy-gap condition they use is not sufficient to discriminate between folding and non-folding sequences (see Fig. 7 of ref. 2). In general, folding rate depends on the entire energy landscape³⁻⁶, not just the energy gap in a highly restricted ensemble.

Ultimately, the Levinthal problem is not that a protein has too many degrees of freedom. It is the shape, not the size alone, of the conformational energy landscape that matters^{4-6,9}. Many large land-

- Šali, A., Shakhnovich, E. I. & Karplus, M. Nature 369, 248–251 (1994).
- Šali, A., Shakhnovich, E. I. & Karplus, M. J. molec. Biol. 2. 235, 1614-1636 (1994).
- 3. Socci, N. D. & Onuchic, J. N. J. chem. Phys. 101, 1519-1528 (1994).
- 4. Camacho, C. J. & Thirumalai, D. 19. Proc. natn. Acad. Sci. U.S.A. 90, 6369-6372 (1993). 5. Chan, H. S. & Dill, K. A. J. chem. Phys. 100
- 9238-9257 (1994).
- 6. Bryngelson, J. D., Onuchic, J. N., Socci, N. D. & Wolynes, P. G. Proteins: Struct. Funct. Genet. (in the press)
- 7. Taketomi, H., Ueda, Y. & Gō, N. Int. J. Peptide Protein Res. 7, 445-459 (1975).
- Zwanzig, R., Szabo, A. & Bagchi, B. Proc. natn. Acad. Sci. U.S.A. 89, 20–22 (1992). Dill, K. A. Curr. Opin. struct. Biol. 3, 99-103 (1993).
- 10. Honeycutt, J. D. & Thirumalai, D. Proc. natn. Acad. Sci. IJ.S.A. 87, 3526-3529 (1990).
- 11. Skolnick, J., Kolinski, A. A. Rev. phys. Chem. 40, 207-235 (1989).
- 12. Baldwin, R. L. Nature 369, 183-184 (1994). Bryngelson, J. D. & Wolynes, P. G. Proc. natn. Acad. Sci. U.S.A. 84, 7524–7528 (1987).
- 14. Guo, Z., Thirumalai, D. & Honeycutt, J. D. J. chem. Phys.
- 97, 525–535 (1992). 15. Leopold, P. E., Montal, M. & Onuchic, J. N. Proc. natn. Acad. Sci. U.S.A. 89, 8721-8725 (1992).
- 16. Miller, R. *J. et al. chem. Phys.* **96**, 768–780 (1992). 17. Goldstein, R. A., Luthey-Schulten, Z. A. & Wolynes, P. G. Proc. natn. Acad. Sci. U.S.A. 89, 9029-9033
- (1.992). Shakhnovich, E., Farztdinov, G., Gutin, A. M. & Karplus, 18. M. Phys. Rev. Lett. 67, 1665-1668 (1991).
- Karplus, M., Caflish, A., Sali, A. & Shakhnovich, E. in Modelling of Biomolecular Structures and Mechanisms (eds Pullman, A. et al.) 69-84 (Kluwer Academic, Dordrecht, 1994).
- 20. Abkevich, V. I., Gutin, A. M. & Shakhnovich, E. J. chem. Phys. 101, 6052-6062 (1994). 21. Camacho, C. J. & Thirumalai, D. Proc. natn. Acad. Sci.
- U.S.A. 90, 6369-6372 (1993). 22. Miyazawa, S. & Jernigan, R. L. Macromolecules 18,
- 534-552 (1985). 23. Go, N. & Abe, H. Avd. Biophysics 18, 149-164 (1984).
- Karplus, M. & Šali, A. Curr. Opin. struct. Biol. 5, 58-73 (1995).

NATURE · VOL 373 · 23 FEBRUARY 1995

ensemble, is a necessary and sufficient condition for rapid folding in the model study. It is necessary because no sequence without such a minimum folds to the native state, and is sufficient because all sequences with such a minimum do fold (Fig. 7 of ref. 2). As to the 11 out of 200 sequences that have their minimum outside the fully compact set, none satisfied the energy condition nor did they fold repeatedly either to the lowest fully compact state or to the lowest energy state found by a Monte Carlo simulation. Thus, these sequences confirm and generalize the folding criterion². Further, the use of the energy condition for quantitatively determining the folding rate has been demonstrated²⁰. There is a strong correlation between the results from the fully compact states and the complete set of states (ref. 2 and Fig. 17 therein).

The nature of the configuration space, as well as the number of conformers, is important for the Levinthal paradox^{1,2}. Surfaces can be constructed for which resolution of the paradox is trivial, but this is not true for the 27-mer since only a fraction of sequences fold rapidly. The large size of the configuration space is necessary for the existence of a paradox. The 27-mer model has 10¹⁶ configurations and requires fewer than 5×10^7 Monte Carlo steps to find the native state. Short oligomers that have been extensively studied on a two-dimensional square lattice^{16,21} may be too small; for example, more Monte Carlo steps (10^5 or more) than there are configurations (4 \times 10⁴) were required for folding a 13-mer¹⁶

The aim of the lattice simulations was to use random interactions so as to determine what differentiates folding from non-folding sequences. The exact choice of parameters was not important, as long as a reasonable set was used. The 27-mer parameters² correspond to the Miyazawa and Jemigan set²² in terms of the magnitude of the interaction energies and their standard deviation.

As to Baldwin's statement in News and Views that we presented a "new view" of protein folding, we agree that some of the concepts in refs 1 and 2 were presaged in earlier work of Go and Abe23 and of others cited in refs 1 and 2. The 27-mer model studies^{1,2,18} provided the first demonstration that the energy-gap condition and a detailed mechanism for resolving the Levinthal paradox could be found a posteriori in computer experiments without having to be introduced explicitly a priori to achieve folding²⁴.

M. Karplus

A. Šali

E. Shakhnovich

Department of Chemistry, Harvard University.

Cambridge, Massachusetts 02138, USA

California 94143-1204, USA

refs 3, 5, 6).

Hue Sun Chan

USA; Carlos J. Camacho, Facultad de Fisica, PUC, Casilla 306, Santiago 22, Chile; Ken A. Dill, Department of Pharmaceutical Chemistry, University of California, San Francisco, California 94143-1204, USA; José N. Onuchic & Nicholas D. Socci, Department of Physics, University of California at San Diego, La Jolla, California 92093-0319, USA; D. Thirumalai, Institute for Physical Science and Technology, University of Maryland, College Park, Maryland 20742, USA; and Peter G. Wolynes, School of Chemical Sciences, University of Illinois, Urbana, Illinois 61801, USA

better than earlier models. Baldwin in

News and Views¹² has said that Šali et al.

were using a potential function of the

Miyazawa-Jernigan type, picked from the

pairwise interactions in the protein data-

base. But, as Šali et al. have noted¹, the

terms are picked from a random gaussian

distribution, not from the databank. Their

potential function is not particularly

physical, as correlations among contact

energies of different pairs of amino-acid

residues are neglected. It is unclear

whether the potential is any more or less

protein-like than any of the potentials

al. as an important "new view" of protein

folding. Naturally, lattice models are

useful for addressing general physical

principles of protein folding, even though

they involve considerable simplification.

However, it is clear from many earlier

efforts, including some that used compa-

rable lattice simulations, that many of the

ideas Baldwin cites as "new" are already

in the literature (refs 4, 10, 13-18, and refs

therein, and reviewed more recently in

Department of Pharmaceutical Chemistry,

This note was written with the assistance and concurrence

of: Joseph D. Bryngelson, NIH, Bethesda, Maryland 20892,

University of California at San Francisco,

Baldwin¹² describes the work of Šali et

used in previous works.

KARPLUS ET AL. REPLY - Chan et al. raise several questions, all of which have simple answers. The object of our study^{1,2} was to examine a large number of sequences and to separate those that fold from those that do not. Consequently, a temperature slightly above $T_{\rm m}$ the midpoint of the folding transition, was used to speed up the reaction. If folding to the native state were always possible under the simulation conditions, as Chan et al. imply there would not have been any non-folding sequences and our computer experiment would have failed. But this is not the case. Further, the same folding kinetics is observed throughout the temperature range where the true native state stability varies from 1 to 40%19

A pronounced energy gap between the native and first excited state (equations