proton transfer. In this case, the pentavalent intermediate would be stabilized by hydrogen bonds to the side-chain amide of Gln 200 with an orientation opposite to that in $G_{1\alpha}$ · GDP · AIF₄ · H₂O. In both mechanisms, the γ -phosphate is the ultimate general base, as proposed for p21^{ras} (refs 11 and 12, and A. Wittinghofer, personal communication), Arg 174 facilitates production of the leaving group⁴, and Gln 200 stabilizes the pentavalent intermediate as proposed for p21ras (ref. 13) and subsequently for $G_{i\alpha 1}$ (ref. 14). Finally, many of the critical interactions involved in the catalytic models presented here are facilitated by rearrangements within regions known to affect GTPase-activating protein (GAP) binding and activity¹⁵¹⁸. We speculate that GAPs accelerate GTPase rates by reorienting these regions in order to stabilize the pentavalent intermediate in a manner similar to that described here.

Mutational analysis is consistent with these mechanisms. Substitution of Gln 61 in $p21^{ras}$ (Gln 200 in $G_{t\alpha}$) is a frequent mutation¹⁹, and 17 different substitutions of Gln 61 lead to a similar ~10-fold reduction in the intrinsic GTPase rate²⁰. Substitution of the homologous Gln 227 in $G_{s\alpha}$ also reduces the intrinsic rate of GTP hydrolysis and is associated with thyroid²¹ and pituitary tumours²². Substitution of Gln 61 with either a glutamic acid residue¹² or an unnatural amino acid containing a nitro group²³ in place of the amide has little effect on the intrinsic or GAP-stimulated hydrolysis rate. These groups could partially substitute for the amide by hydrogen-bonding to a protonated phosphate. Substitutions of Gly 12 in p21ras (Gly 38 in $G_{t\alpha}$, conserved throughout the GTPase superfamily and the site of the most frequent oncogenic mutation in p21^{ras} (ref. 19), also disrupt intrinsic GTPase activity. Like the GTP-bound forms of $p21^{ras}$ and $G_{t\alpha}$, Gly 38 in $G_{t\alpha} \cdot \text{GDP} \cdot \text{AlF}_4^- \cdot \text{H}_2\text{O}$ does not have unusual $\phi - \psi$ angles. However, the structure of $G_{1\alpha} \cdot GDP \cdot AlF_4 \cdot H_2O$ shows that a β -substituent would weaken the stabilization of the pentavalent intermediate by sterically hindering the interaction of Gln 200 with a protonated, equatorial phosphate oxygen as well as the watermediated hydrogen bond between Gln 200 and Glu 39. Other residues described as interacting with GDP · AlF₄ · H₂O (Gly 199 for example) are also conserved in G-protein families and affect GTPase rates²³. In general, the ability of aluminium fluoride to affect numerous enzymes^{24,25} that catalyse the hydrolysis of nucleotides, including tubulin, myosin, F1-ATPase, hexokinase, and actin suggests that our conclusions may be applicable to many other systems.

◄ FIG. 3 Stereochemistry of the GTPase mechanism. a, Interactions involving GTP γ S or GDP·AIF₄⁻·H₂O at the active site of G_{ta} immediately relevant to the GTPase mechanism. GDP·AIF₄·H₂O mimics the pentavalent intermediate; that is, the square planar arrangement of fluoride ions coordinating the Al³⁺ ion is replaced by a triangular planar arrangement of oxygen atoms surrounding the γ -phosphorus. The positively charged guanidinium group of Arg 174 would increase the transfer of negative charge from the attacking hydroxyl to the $\beta - \gamma$ bridging oxygen, facilitating the collapse of the pentavalent intermediate to a terminal β phosphate and a free orthophosphate. Omitted for clarity are supporting ligands found in similar conformations in both $G_{t\alpha}$ ·GTP γ S and GtaGDP·AIF4 · H2O. b, Concerted mechanism for GTP hydrolysis in which transfer of a proton to the γ -phosphate is coupled to the deprotonation of the attacking water. Tautomerization of GIn 200 enhances the nucleophilicity of the attacking water, reduces the negative charge of the γ -phosphate and stabilizes the pentavalent intermediate through two hydrogen bonds. c, Alternative mechanism for GTP hydrolysis involving direct transfer of a proton from the attacking water molecule to the γ -phosphate oxygen. This transfer is aided by the peptide NH of Gln 200 which is polarized by its contact with Arg 204. Here the pentavalent intermediate is stabilized by hydrogen bonds to a reorientated GIn 200 side-chain amide.

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ERRATUM

Interaction between two homeodomain proteins is specified by a short C-terminal tail

Martha R. Stark & Alexander D. Johnson

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PART c of Fig. 1 of this Letter was accidentally omitted and is now shown here.