

- European Commission *The Evaluation of Tests for the Diagnosis of Transmissible Spongiform Encephalopathy in Bovines* (http://europa.eu.int/comm/food/fs/bse/bse12_en.html).
- Grassi, J. *et al. Arch. Virol.* **16** (suppl.), 197–205 (2000).
- Wells, G. A. H. *et al. Vet. Rec.* **142**, 103–106 (1998).

Genome evolution

Gene capture in archaeal chromosomes

Free genetic elements can be readily integrated into bacterial chromosomes, but so far, with the exception of one virus, there has been no evidence that this happens in Archaea — the other domain of microorganisms. Here we show that site-specific integration of different genetic elements into archaeal chromosomes is a general phenomenon, albeit rare, which requires an archaeal integrase and produces a partitioned integrase gene in the chromosome. The process is distinct from bacterial mechanisms and has implications for how horizontal gene transfer might occur across the boundaries of the domains of life.

Many bacterial integrases belong to a superfamily of tyrosine recombinase enzymes¹ and mediate site-specific recombination between small extra-chromosomal DNA elements and chromosomes², facilitating horizontal gene transfer between bacteria². The only known archaeal integrase effects site-specific and reversible integration of the SSV1 virus into the chromosome of *Sulfolobus shibatae*^{3,4} (Fig. 1a, top).

We have previously identified a region of the *Sulfolobus solfataricus* genome that shows high sequence similarity to the *Sulfolobus* plasmid pHEN7 and to other members of the pRN plasmid family^{5–7}. This chromosomal region is flanked by open-reading

frames (ORFs) with extensive sequence similarity to the amino-terminal (66 amino acids) and carboxy-terminal (269 amino acids) regions of the SSV1 integrase (Fig. 1a). The former, Int(N), overlaps with the downstream half of a gene encoding tRNA^{Val} and shows 71% sequence identity to the 45-base-pair target site (*att*) of the SSV1 integrase⁴ (within the downstream half of a tRNA^{Arg} gene), whereas the latter, Int(C), contains an identical *att* site (Fig. 1a, b).

Excision from the chromosome by recombination at the direct *att* repeat and circularization creates a plasmid, pXQ1, containing an intact integrase gene and one copy of the *att* site, similar to that of the SSV1 virus⁴. Thus, with the aid of the encoded integrase, pXQ1 could have integrated into the chromosome. As neither the free form of pXQ1 nor the empty chromosomal site of *Sulfolobus* were detectable by polymerase chain reaction (our unpublished results), we infer that the absence of an intact integrase gene precludes integrase-mediated excision from the chromosome.

A search for more homologues of the SSV1 integrase encoded in the chromosome of *S. solfataricus*⁵ revealed three Int(N)s and one Int(C). The sequences encoding Int(N) overlap with downstream halves of tRNA genes and all the sequences contain *att* sites. By analogy to the SSV1 virus, a genetic element XQ2 (67.7 kilobases) can be formed by recombination and excision at the *attL* and *attR* sites (Fig. 1a, b).

Four of the ORFs encoded in XQ2 were assigned to genes encoding the enzymes dTDP–glucose-4,6-dehydratase, glucose-1-phosphate–thymidyl transferase, dTDP–4-dehydrorhamnose reductase and dTDP–4-dehydrorhamnose-3,5-epimerase, all of which are involved in central metabolism,

and three of which are generally clustered in one operon. However, there are two additional copies of dTDP–glucose-4,6-dehydratase and another copy of each of the other enzymes encoded within the *S. solfataricus* chromosome⁵ that show 76–84% sequence identity and 86–96% similarity. It is therefore likely that XQ2 entered the chromosome, aided by its own integrase, to produce a gene-capture event.

We searched the complete genome sequences of seven other archaea for integrase homologues⁸. Those from the euryarchaeote *Pyrococcus horikoshii*⁹ and the crenarchaeote *Aeropyrum pernix*¹⁰ each had two Int(N), overlapping downstream halves of tRNA genes, and two and one copy of Int(C), respectively. The genome of *Pyrococcus* OT3 has two partitioned integrase genes¹¹. Complementary fragments of the integrase genes contain *att* sites and border regions of 21.5 and 4 kilobases in *P. horikoshii* and 17.5 kilobases in *A. pernix*. Compatible with the idea that these are inserts from unknown organisms, none of their ORFs reveals any hits in the GenBank/EMBL databases⁸. We have detected a total of seven partitioned integrase genes, on average one per genome. Their sequences vary markedly in the amino-terminal region but are less variable in the carboxy-terminal region (22–33%/41–51% identity/similarity), indicating that other integrase genes may still remain to be discovered.

We conclude that chromosomal integration in archaea differs from that in bacteria in producing partitioned integrase genes and that integration can be reversed only if the intact integrase is produced⁴. ‘Curing’ the cell of the free genetic element carrying the intact integrase gene can thus lead to gene capture by the archaeal chromosome. Although archaeal integrases differ from bacterial integrases in lacking a conserved motif¹ and in having *att* sites in the coding region, they may still facilitate gene transfer between archaea, bacteria and eukaryotes¹².

Qunxin She*, Xu Peng*, Wolfram Zillig†, Roger A. Garrett*

*Microbial Genome Group, Institute of Molecular Biology, University of Copenhagen, Sølvgade 83H, DK-1307 Copenhagen K, Denmark

†Max-Planck-Institut für Biochemie, D-82152 Martinsried, Germany

- Nunes-Düby, S. E. *et al. Nucleic Acids Res.* **26**, 391–406 (1998).
- Rowe-Magnus, D. A. & Mazel, D. *Curr. Opin. Microbiol.* **2**, 483–488 (1999).
- Reiter, W.-D. & Palm, P. *Mol. Gen. Genet.* **221**, 65–71 (1990).
- Muskhelishvili, G., Palm, P. & Zillig, W. *Mol. Gen. Genet.* **237**, 334–342 (1993).
- She, Q. *et al. DNA Sequence* **11**, 183–192 (2000).
- Kletzin, A., Lieke, A., Ulrich, T., Charlebois, R. L. & Sensen, C. W. *Genetics* **152**, 1307–1314 (1999).
- Peng, X., Holz, I., Zillig, W., Garrett, R. A. & She, Q. *J. Mol. Biol.* **303**, 449–454 (2000).
- Altschul, S. F. *et al. Nucleic Acids Res.* **25**, 3389–3402 (1997).
- Kawarabayasi, Y. *et al. DNA Res.* **5**, 55–76 (1998).
- Kawarabayasi, Y. *et al. DNA Res.* **6**, 83–101 (1999).
- Makino, S., Amano, N., Koike, H. & Suzuki, M. *Proc. Jpn. Acad. Ser. B* **75**, 166–171 (1999).
- Groth, A. C., Olivares, E. C., Thyagarajan, B. & Calos, M. P. *Proc. Natl Acad. Sci. USA* **97**, 5995–6000 (2000).

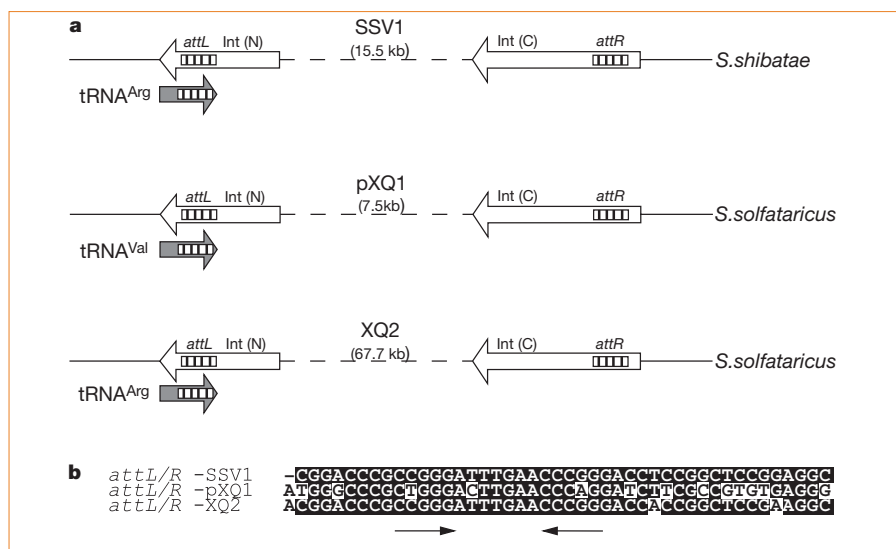


Figure 1 Site-specific integration of genetic elements into archaeal chromosomes. **a**, Organization of the SSV1 virus and the putative genetic elements pXQ1 and XQ2 in the chromosomes of *S. shibatae* and *S. solfataricus*. The relative positions of the partitioned integrase genes, the *att* sites (45 base pairs) and the overlapping tRNA genes are indicated. **b**, Alignment of the direct repeats *attL* and *attR* from virus SSV1 and from pXQ1 and XQ2. Nucleotides are shaded if two or more positions are identical. Arrows indicate a 5-base-pair inverted repeat.