

Genome stability

Transgenerational mutation by radiation

Parental exposure to ionizing radiation increases the frequency of germline mutations detectable in the next generation¹. Parental exposure can also increase the rate of mutation in somatic cells^{2,3} and confer a predisposition to cancer⁴⁻⁶ in offspring, suggesting that there could be an indirect effect of radiation on somatic genome stability that is transmissible through the germ line of the irradiated parents. We have found that this indirect effect extends to the germ line of unexposed first-generation offspring in mice, as revealed by an increased instability of repeat-DNA sequences in their descendants.

CBA/H male mice were exposed to fission neutrons and mated to unexposed females (Fig. 1a). Exposure resulted in a 6-fold increase in paternal mutation rate at expanded simple-tandem repeat (ESTR) loci^{7,8} detectable in first-generation (F₁) offspring (Table 1). Breeding from these unexposed F₁ mice revealed that germline mutation rates remained significantly high in transmissions from both F₁ males and F₁ females (6-fold and 3.5-fold increase, respectively, compared to unexposed control families and to the alleles transmitted from control breeding partners; not shown).

This increase was in part due to increased mutational mosaicism in the germ line of the F₁ mice, with more than 50% of *de novo* mutations detected in F₂ mice being shared by two or more littermates, compared with less than 25% in the offspring of both non-irradiated mice and F₀ mice exposed to X-rays⁹ or fission neutrons (Table 1 and Fig. 1b). However, singleton mutations (those present in only one member of a litter) were also more frequent in these F₂ mice than in the controls, significantly so for male transmissions (Table 1). We conclude that paternal exposure to radiation results in the destabilization of ESTR loci in the germ line of offspring, and that some of the resulting mutations occur sufficiently early in germline development for significant levels of mosaicism to arise.

This remarkable stimulation of muta-

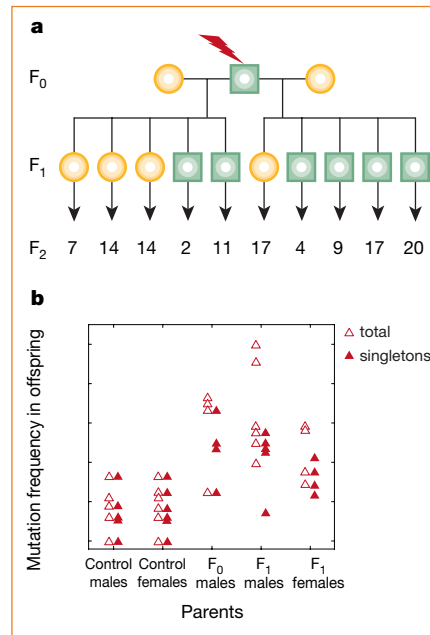


Figure 1 Expanded simple-tandem repeat mutation in the descendants of irradiated male mice. **a**, Design of the transgenerational study: the number of F₂ offspring derived from each F₁ mouse is indicated. **b**, Frequency of ESTR mutation in offspring of F₀ and F₁ males and females. Four CBA/H males were given whole-body chronic irradiation of 0.5 Gy of fission neutrons (²⁵²Cf source, 0.03 Gy min⁻¹) and 10 weeks later mated to non-irradiated CBA/H females, ensuring that litters were derived from irradiated stem cells. Litters from one irradiated male were used for subsequent breeding of F₂ mice. DNA profiles were produced⁹ using the mouse single-locus ESTR probes *Ms6-hm* and *Hm-2* (refs 7,8).

tion in the F₁ germ line following paternal irradiation is reminiscent of delayed radiation-induced genome instability in somatic cells, where radiation can induce genetic changes in some of the descendants of a single irradiated cell, an effect that can persist for many cell generations¹⁰. By analogy, exposure of F₀ mice must create a signal in the F₀ germ line that is transmitted by a single sperm to the F₁ offspring, in which instability arises many cell generations later and is detected in some but not all cells in the developing germ line.

It is unlikely that the negligible cytoplasmic component of the mature sperm could carry substantial amounts of long-lived free radicals or other radiation-induced species into the egg, so the transmitted signal is probably DNA-dependent. It is most

unlikely that this signal is radiation-induced damage at the ESTR loci themselves, because it is transmissible through meiosis and mitosis and appears to operate *in trans* in the F₁ germ line, affecting alleles both from the exposed F₀ male and from the unexposed F₀ female (data not shown).

Increased ESTR mutation rates appear to be uniform in the germ line of all F₁ offspring derived from 10 independent sperm from the irradiated F₀ male (Fig. 1b; χ^2 test for homogeneity, $P > 0.5$), suggesting that radiation-induced *de novo* mutations in DNA-repair genes are not the cause of instability in the F₁ germ line. The alternative, therefore, is a radiation-exposure signal inherited through sperm in an epigenetic fashion, perhaps through changes in DNA methylation. Methylation is transmissible through many cell divisions¹¹ and can influence the activity of DNA-repair systems¹². If radiation-induced DNA damage in the germ line triggers an epigenetic signal that influences DNA repair and can survive the reprogramming of DNA methylation during spermatogenesis and early development, then subsequent activation of this signal in the germ line could lead to the observed transgenerational mutagenesis.

These findings have potential implications for risk evaluation in humans. So far, the main concern of radiation protection is focused on the direct mutagenic effects of ionizing radiation in exposed individuals¹. However, the persistence of instability into the germ line of unexposed offspring of irradiated mice raises the issue of delayed genetic risk, as well as the potential for radiation to cause mutation clustering and to contribute to mosaicism in germ cells, long recognized as an important mechanism in the origin of human genetic disorders¹³.

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Table 1 Mutation in two mouse generations at ESTR loci *Ms6-hm* and *Hm-2*

Group	Offspring (litters)	All mutations				Singletons			
		Mutations	Rate	Ratio*	P†	Mutations‡	Rate	Ratio*	P†
Control males	74 (12)	8	0.0540	-	-	6 (75%)	0.0405	-	-
Control females	74 (12)	9	0.0608	-	-	7 (77.8%)	0.0473	-	-
F ₀ males, 0.5 Gy of neutrons	34 (7)	22	0.3235	5.98	8.62 × 10⁻⁷	16 (72.7%)	0.2353	5.80	6.59 × 10⁻⁵
F ₁ males	63 (11)	38	0.3016	5.58	5.81 × 10⁻⁸	14 (36.8%)	0.1111	2.74	0.0444
F ₁ females	52 (6)	22	0.2115	3.45	0.0007	11 (50%)	0.1058	2.24	0.1291

*Ratio to control.

†Probability of difference from the control group (Fisher's exact test, two-tailed; statistically significant values are in bold).

‡Percentage of singleton mutations is given in parentheses.