

Transcription

Gene silencing in worms and fungi

The introduction into cells of foreign nucleic acid molecules can induce sequence-specific gene silencing in some organisms. Here we show that two distantly related organisms, the nematode *Caenorhabditis elegans* and the fungus *Neurospora crassa*, which have quite different mechanisms of gene silencing, both use a similar protein to control the process. This suggests that they may share an ancestral mechanism that evolved to protect the genome against invasion by foreign DNA.

In a process known as dsRNA interference (HDGS), gene expression can be prevented by either DNA or RNA molecules. For example, double-stranded (ds) RNA can inhibit gene expression¹ in *C. elegans*, *Drosophila* and *Trypanosoma*, and transgenic DNA can induce gene silencing in plants² and fungi³.

In a process known as dsRNA interference, dsRNA silences the expression of endogenous *C. elegans* genes after they have been transcribed by inducing sequence-specific degradation of homologous messenger RNA molecules⁴, preventing their translation into protein. Plants and fungi also use post-transcriptional gene silencing (PTGS), but this is induced by transgenic DNA. Again, transcription of the target gene is unaffected but its transcripts do not accumulate as a consequence of rapid degradation.

There are several indications that RNA intermediates are involved even in transgene-induced gene silencing — transgenes produce RNA molecules known as aberrant RNAs, which somehow manage to induce messenger RNA degradation⁵. The two mechanisms of PTGS and dsRNA interference both involve sequence-specific degradation of homologous messenger RNA and an ability of silencing to spread from cell to cell.

Although HDGS phenomena have been considered to be related mechanistically and evolutionarily, no experimental evidence has been forthcoming to support this idea, principally because so far it has not been possible to compare the underlying genetic mechanisms. Genetic characterization of PTGS has been reported for *Arabidopsis thaliana*⁶ and *N. crassa*⁶, and mutants unable to perform dsRNA interference have been isolated from *C. elegans*^{7,8}. These mutants should enable the different components of the cell's gene-silencing machinery to be identified. By comparing the sequences of the respective protein products, we have found that the *qde-2* gene, which controls transgene-induced gene silencing in *N. crassa*⁶, is homologous

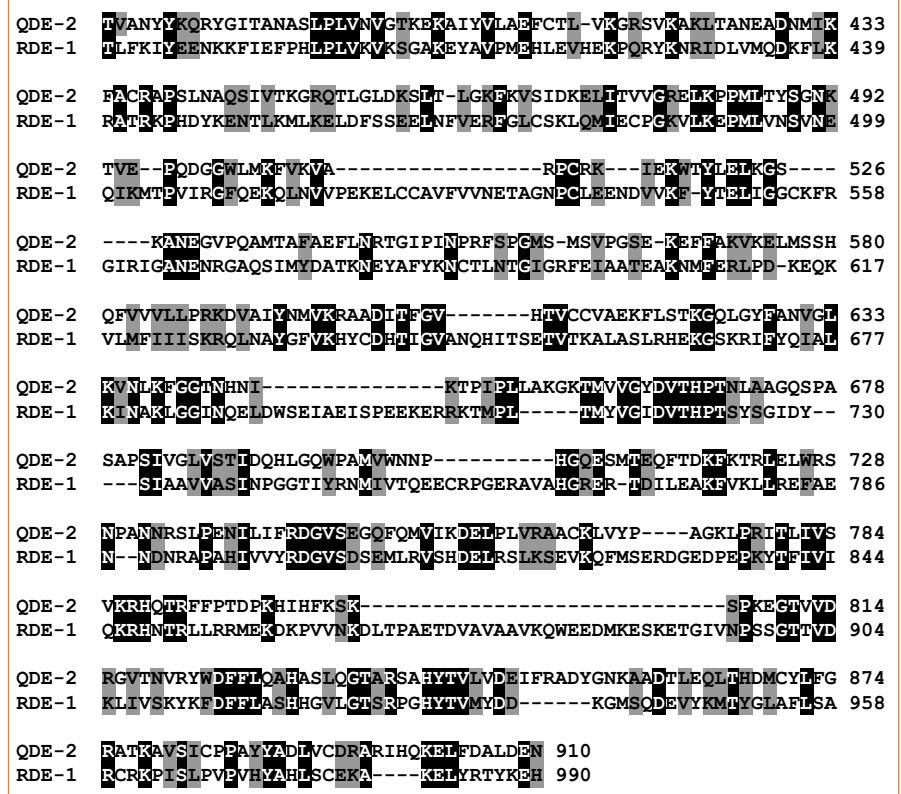


Figure 1 Sequence alignment of the QDE-2 protein (Genbank accession number AF217760) with RDE-1 protein (AF180730) of *C. elegans*. The conserved carboxy-terminus region (expected value 10⁻²³, 38% similarity) is shown. Identical residues are in black; conservative substitutions are in grey.

to the *rde-1* gene⁷, which is essential for dsRNA interference in *C. elegans* (Fig. 1).

This finding is, to our knowledge, the first experimental evidence indicating that dsRNA interference and PTGS induced by transgenic DNA share a common genetic mechanism. It supports the idea that HDGS phenomena evolved from an ancestral mechanism aimed to protect the genome against transposons and viruses. Our results also suggest that dsRNA molecules might participate in PTGS in fungi.

dsRNA could be produced directly from integrated transgenes as a result of the presence of inverted repeats, or as an outcome of transcription from convergent inverted promoters. Alternatively, transgenic single-stranded aberrant RNA may be used as a template by QDE1, a putative RNA-dependent RNA polymerase⁹, to produce dsRNAs. Further analysis should define the mechanistic function of *rde-1* and *qde-2* in gene silencing and confirm whether the homology between these two genes effectively corresponds to a cognate step in the two gene-silencing mechanisms.

As well as including *rde-1*, the *qde-2* gene family has members in both plants

and animals: in *A. thaliana* and *Drosophila*, *qde-2* homologues have been implicated in the regulation of development¹⁰, and the rabbit *qde-2* homologue, eIF2C, forms part of a protein complex that stimulates the start of translation¹¹. As all the *qde-2* homologues have not yet been biochemically characterized, it is possible that these evolutionarily related genes could be required for different biological processes.

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