

the statistical analysis and M. Ondrak and P. Schefter for technical assistance.

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Oestradiol and its biosynthesis in *Phaseolus vulgaris* L.

UNTIL recently the presence of steroidal oestrogens in plants has been a matter for debate¹, and any physiological significance associated with them has been generally disregarded^{2,3}. Although steroidal oestrogens have been found in extracts of a few species⁴, the identification of oestrone in *Phaseolus* was based on equivocal procedures⁵. Moreover, as with any compound present in low concentration, it is possible that at least some of the oestrogens identified in plant extracts are attributable to external contamination. This risk can be eliminated by demonstrating the incorporation of radioactive label from a known precursor into the compounds of interest. We report here the incorporation of radioactive isotopes from (3RS)-2-¹⁴C-mevalonic acid DBED (MVA), 4-¹⁴C-oestrone and potassium 6,7-³H-oestrone sulphate into oestradiol by *Phaseolus vulgaris* L., cultivar Canadian Wonder. This represents the first case known to us of successful labelling of an identified steroidal oestrogen from ¹⁴C-MVA in a higher plant. It seems therefore that this class of mammalian sex hormone is produced in plants as well as animals.

Eight-day-old seedlings of *P. vulgaris* were sectioned under water at the base of the hypocotyl, stood in aqueous solutions of the various radioactive substances and allowed to transpire for 5 d (ref. 6). They were then planted in potting compost and grown for 2–5 weeks to facilitate normal growth and development. Harvested plants were solvent extracted (chloroform:methanol::2:1) under reflux in a Soxhlet apparatus for 8 h. The extract was reduced to dryness *in vacuo* and taken up in ether. The phenolic fraction was isolated by partitioning with 1N NaOH, followed by acidification of the aqueous phase and repartitioning into ether. Thin-layer chromatography (TLC) and column chromatography on Sephadex LH-20 were used to isolate those fractions expected to contain oestrone (E₁) and oestradiol (E₂).

The chromatographic mobilities of metabolites were compared with authentic compounds using TLC on three solvent systems (chloroform:methanol::95:5; benzene:propan-2-ol::95:5; petroleum ether:ethyl acetate::75:25), column chromatography on LH-20 of the underivatized extracts and radio-gas-liquid chromatography (radio GLC) on OV-101 of the trimethylsilyl ether derivatives. Samples believed to contain oestradiol were diluted with non-radioactive carrier oestradiol and recrystallised to constant specific activity.

Using these techniques in combination, ¹⁴C was observed to be incorporated into oestradiol from MVA and from 4-¹⁴C-oestrone. Similarly, oestradiol could be labelled from the

³H-oestrone sulphate. Attempts to label oestrone have not been conclusive; the rate of incorporation from radio-active MVA into the putative oestrone peak was much less than that observed for oestradiol.

One previous report describes the incorporation of ¹⁴C-MVA into a phenolic fraction by the higher plant *Haplo-pappus heterophyllus*, but the radioactive phenolic material did not correspond to any common steroidal oestrogen⁷. There have been many studies of the biotransformation of the mammalian steroid hormones progesterone and testosterone^{4,8}, but our findings seem to represent the first report of a metabolic transformation of an oestrogen in plants—reduction of the 17-keto steroid E₁ to the 17β-hydroxyl compound E₂.

Evidence of unlabelled oestradiol in extracts of seeds, vegetative and flowering plants was obtained using a modified radioimmunoassay (RIA) technique⁹ and combined gas chromatography-mass spectrometry (GC-MS). The GC-MS analysis¹⁰ involved continuous monitoring at the collector of the ion abundance at m/e 416 (molecular ion of the bis-trimethylsilyl derivative of oestradiol) as generated in the ion source from the GLC effluent of an SE-30 column operated at 250 °C. This process, commonly referred to as single-ion monitoring, enabled us to use the GC-MS system as a semi-specific detector for oestradiol.

This identification contrasts with the results of Kopcewicz⁵ who presented TLC evidence for oestrone in *P. vulgaris* but could not detect oestradiol in the extracts. Indeed, this is only the second report of oestradiol detected as a plant steroid, Awad¹¹ having isolated both oestrone and oestradiol from seeds of *Prunus*.

The verification of the endogenous biosynthesis of a steroid oestrogen in *P. vulgaris* opens the way to further studies of the suggested correlation between the quantitative levels of these compounds in this species and the onset of flowering⁵. Preliminary data obtained by RIA suggest that oestradiol is present at 2–10 μg kg⁻¹ fresh weight in seeds and leaves. It should also be possible to examine the overall pathway of biosynthesis of steroids in plants in order to investigate the presumed homology between plants and animals⁴. In addition, the potential for hormonal heterophyly¹² with respect to oestrogens in plants requires attention.

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