Eron, L., and Block, R., Proc. natn. Acad. Sci. U.S.A., 68, 1828-1832 (1971).
 Chen, B., de Crombrugghe, B., Anderson, W. B., Gottesman, M. E., and Pastan, I., Nature new Biol., 233, 67-70 (1971).
 de Crombrugghe, B., and Chen, B., Gottesman, M., Pastan, I., Varmus, H. E., Emmer, M., and Perlman, R. L., Nature new Biol., 230, 37-40 (1971).
 de Crombrugghe, B., Chen, B., Anderson, W., Nissley, P., Gottesman, M., and Pastan, I., Nature new Biol., 231, 139-142 (1971).
 Ohki, M., and Mitsui, H., Nature, 252, 64-66 (1974).
 Ohki, M., J. molec. Biol., 68, 249-264 (1972).
 Ohki, M., Doi, O., and Nojima, S., J. Bact., 110, 864-869 (1972).
 Wilson, G., Rose, S. P., and Fox, C. F., Biochem. biophys. Res. Commun., 38, 617-623 (1970).
 Pardee, A. B., Jacob, F., and Monod, J., J. molec. Biol., 1, 165-178 (1959). 617-623 (1970).
 Pardee, A. B., Jacob, F., and Monod, J., J. molec. Biol., 1, 165-178 (1959).
 Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. I., J. biol. Chem., 193, 265-275 (1951).
 Haseltine, B., and Muller-Hill, B., in Experiments in Molecular Genetics (edit. by Miller, J. H.), 319 (Cold Spring Harb. Lab., New York, 1972).
 Perlman, R. L., and Pastan, I., Biochem. biophys. Res. Commun., 30, 656-664 (1968). (1968).
Ullmann, A., and Monod, J., FEBS Lett., 2, 57-60 (1968).
Schwartz, D., and Beckwith, J. R., in The Lactose Operon (edit. by Beckwith, J. R., and Zipser, D.) 417 (Cold Spring Harb. Lab., New York, 1970).
Perlman, R. L., and Pastan, I., Biochem. biophys. Res. Commun., 37, 151-157 (1969).

C_H3 domain of IgG as binding site to Fc

receptor on mouse lymphocytes

18 Sankaran, L., and Pogell, B. M., Nature new Biol., 245, 257-260 (1973).

MEMBRANE receptors recognising the Fc portion of immunoglobulin molecules (Fc receptors) are found in many cells of the immune system¹⁻⁴. Fc receptors on lymphocytes are readily detected by a rosette test3,5,6 and this reaction is inhibited by pretreatment of the lymphocytes with IgG (ref. 3). IgG proteins lacking almost the entire C_H1 and C_H3 homology regions have been obtained from mutant cell lines of MOPC 21, a plasmacytoma secreting IgG1 (refs 7 and 8) and the extent of the deletions determined (Fig. 1). To identify that part of the IgG molecule which interacts with the Fc receptor, we have tested the ability of these IgG proteins to inhibit Fc rosette formation on murine lymph node cells. We show (Table 1) that an intact C_H3 region is essential for the binding of IgG to Fc receptors on lymph node cells.

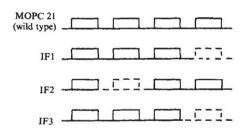


Fig. 1 Structure of the mutant proteins. Dotted lines indicate deletions in the heavy chains, shown diagrammatically, with the four intra-chain disulphide bridges. IF1, IF2 and IF3 were cloned tissue culture lines of the mouse plasmacytoma MOPC 21. IF2 has a deletion in the heavy chain of residues 121 to 214 corresponding to the wild type protein^{8,16}, that is, of almost the entire C_H1 region. IF1 has a deletion of residue 358 onwards¹⁶ and IF3 of residue 342 onwards (K. A. and C. Milstein, unpublished). The secreted protein used in the experiments was purified from the serum of mice bearing the respective tumours, using ammonium sulphate precipitation and DEAE cellulose chromatography. The purity was checked using electrophoresis on cellulose acetate strips, and isoelectric focusing on polyacrylamide gels.

The ability of IF2 protein to inhibit Fc rosette formation shows that the CH1 region is not the site through which IgG binds to Fc receptors. The failure of IF1 and IF3 proteins to inhibit Fc rosettes shows that an intact CH3 region is required for the binding of IgG to Fc receptors. Since both IF1 and IF3 possess intact C_H2 regions, this region may not be required for binding to Fc receptors. It is possible, however, that the absence of the CH3 region in IF1 and IF3 can result in a rearrangement of the C_H2 region, destroying an active site wholly or partly within this region. The inability to detect the C_H3 region of membrane-bound IgG on lymphocytes, with a fluorescent antiglobulin possessing an anti-C_H3 region

Table 1 Inhibition of rosette formation with IgG proteins

		Percentage reacting cells
Inhibi	tor	
Experiment 1		i s.e
No inhibitor		33.8 : 2.3
MOPC 21 (parent IgG) 0.58 mg ml ⁻¹		18.0 + 2.9
IF1 (C _H 3 deletion)	0.58 mg ml ⁻¹	30.9 ± 1.9
IF2 (C _H 1 deletion)	0.58 mg ml ⁻¹	3.3 ± 3.7
Experiment 2		
No inhibitor		34.1 ± 2.5
MOPC 21 (parent IgG) 10 mg ml ⁻¹		0.1 + 0.1
IF1 (C _H 3 deletion)	10 mg ml^{-1}	35.9 ± 1.1
Experiment 3		
No inhibitor		43.6
MOPC 21 (parent IgC	3) 5 mg ml^{-1}	9.4
IF3 (C _H 3 deletion)	5 mg ml ⁻¹	41.4 - 1.8
IF1 (C _H 3 deletion)	5 mg ml ⁻¹ (hea	t
	aggregated)	43.8 + 2.9

Pooled lymph node cells from BALB/c mice were used in the experiments. Fc rosette formation was carried out as previously described5.6. For inhibition of Fc rosettes, lymphocytes were pretreated with inhibitor protein at the concentrations stated for 15 min at 0°C before rosette formation. For one experiment IF1 protein was heat-aggregated at 63° C. The results are the means of more than three replicate preparations when the standard error of the mean is given. Two observations in experiment 3 were the results of counting over 200 cells in a single preparation.

specificity, also suggests that it is the C_H3 region which interacts with the lymphocyte membrane9.

In an extension of the domain hypothesis, Edelman et al.¹⁰ proposed that each of the homology regions of the polypeptide chains of immunoglobulin molecules folds to form a compact domain which has evolved to perform independent functions. Thus the V_H and C_H2 regions of IgG possess the antigen binding and complement fixing sites11, respectively. The binding of IgG to human monocytes12 and guinea pig macrophages13, possibly through a site in the CH3 region, has been suggested as evidence for the domain hypothesis. The binding of IgG to Fc receptors on K cells14, neutrophils14 and a non-secreting plasmacytoma¹⁵ also requires an intact C_H3 region. The C_H3 region of IgG seems therefore to possess the property of binding IgG to Fc receptors, which may be functionally relevant, on many types of cells.

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Basten, A. B., Miller, J. F. A. P., Sprent, J., and Pye, J., J. exp. Med., 135, 610-626 (1972).
 Bloch, K. J., Prog. Allergy, 10, 84-150 (1967).
 Cline, M. J., Sprent, J., Warner, N. L., and Harris, A. W., J. Immunol., 108, 1126-1128 (1972).
 Lay, W. H., and Nussenweig, V., J. exp. Med., 128, 991-1009 (1968).
 Hallberg, T., Gurner, B. W., and Coombs, R. R. A., Int. Archs Allergy appl. Immun., 44, 500-513 (1974).
 Ramasamy, R., and Munro, A. J., Immunology, 26, 563-570 (1974).
 Cotton, R. G. H., Secher, D. S., and Milstein, C., Eur. J. Immunol., 3, 135-140 (1973).
 Milstein, C., Adetugbo, K., Cowan, N. J., and Secher, D. S., in Progress in

(1973).
Milstein, C., Adetugbo, K., Cowan, N. I., and Secher, D. S., in Progress in Immunology, 2 (edit by Brent, L., and Holborow, E. J.), 157-168 (North-Holland, Amsterdam, 1974).
Froland, S. S., Natvig, J. B., and Stavem, P., Scand. J. Immunol., 1, 351-360 (1972).

Kehoe, J. M., Cunningham, B. A., Gall, W. E., Gottlieb, P. D., Rutishauser, U., and Waxdal, M. J., Proc. natn. Acad. Sci. U.S.A., 63, 78-85 (1969).
 Kehoe, J. M., and Fougereau, M., Nature, 224, 1212-1213 (1969).
 Okafor, G. O., Turner, M. W., and Hay, F. C., Nature, 248, 228-230 (1974).
 Yasmeen, D., Ellerson, J. R., Dorrington, K. J., and Painter, R. M., J. Immunol., 110, 1706-1709 (1973).
 MacLennan, I. C. M., Connel, G. E., and Gotch, F. M., Immunology, 26, 303-310 (1974).
 Ramasamy, R., Munro, A. J., and Milstein, C., Nature, 249, 573-574 (1974).
 Secher, D. S., Cotton, R. G. H., and Milstein, C., FEBS Lett., 37, 311-316 (1973)