a molecular weight in the IgG and IgM area detected in the indirect test is an antibody or antibody–antigen complex or a low molecular weight T-cell product combined with a high molecular weight picrylated protein.

The following experiments provide immunochemical evidence for the specificity of suppressor factor and show that it can be specifically absorbed by and eluted from picrylated but not by 'oxazolonated' protein bound to Sepharose beads.

Suppressor supernatant (16 ml) was absorbed on to picrylated or oxazolonated beads. For elution the beads used for the first absorption (8 ml) were washed four times in 200 ml ice cold saline and resuspended in 3 ml tissue culture medium with 1% foetal calf serum and heated at 56° C for 30 min. The supernatant was removed and 3 ml of 2 M NaCl added. This was removed after 15 min at room temperature and followed by 0.1 M, *p*H 2.7 glycylglycine-hydrochloric acid buffer for 15 min. The eluates were dialysed against phosphate buffered saline, *p*H 7.2, followed by tissue culture medium. Foetal calf serum was added to 5% and the eluate assessed in the direct test.

Table 2 shows that suppressor activity was specifically absorbed by and eluted from picrylated beads. This specific absorption shows that this suppressor factor is not an anti-receptor site factor of the type described by Ramseier[4]. It is not known whether the suppressor factor is a specific T cell product or a complex of the specific T cell product and antigen. In the latter case the molecular weight of the T cell product without antigen is less than about 50,000 daltons and may be much lower.

The suppressor factor described here has a similar molecular weight to the suppressor factors which specifically prevent a tumour rejection (S. Fujimoto, M. Green, and A. H. Sehon, unpublished) and specifically depress the homocytotrophic (presumptive IgE) antibody response[5]. Purification by gel filtration and absorption and elution from antigen opens up the possibility of detailed immunochemical studies.

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Errata

In the article "Stimulation of synaptosomal dopamine synthesis by veratridine" by R. L. Patrick and J. D. Barchas (*Nature*, **250**, 737; 1974) the following corrections are necessary. Paragraph 2, line 6 should read . . . fraction has been shown to be associated with the synaptosomal component . . . and not as printed. In line 4 of the last paragraph interact should read intact. There were also errors in the Tables which, for the sake of clarity, are reprinted in full below.

Table 1 Effect of tetrodotoxin on the veratridine-induced increase in dopamine synthesis

	Dopamine synthesis (nmol h^{-1} g^{-1})
Controls	9.54 ± 0.53 (12)
Tetrodotoxin	9.50 ± 0.57 (12)
Veratridine	14.7 ± 0.46 (12)*
Tetrodotoxin + veratridine	10.5 ± 0.43 (12)

Aliquots of the striatal P_2 fraction were incubated for 5 min at 37° C either without further additions or in the presence of tetrodotoxin (2×10^{-7} M) followed either by the simultaneous addition of veratridine (7.5×10^{-5} M) plus L-1-^{14}C-tryosine (1×10^{-5} M) or by the addition of tyrosine alone, and incubated for an additional 5 min. The apparent rate of synthesis was calculated by dividing the d.p.m. of product formed per hour per gram of original tissue by the specific activity of the tyrosine added to the medium. The normal incubation medium had the following composition: NaCl, 125 mM; KCl, 5 mM; $CaCl_2$, 1 mM; $MgCl_2$, 1 mM; glucose, 10 mM; ascorbic acid, 1 mM (made fresh daily); and Tris-HCl, 50 mM *p*H 7.4. Values represent the mean ± s.e.m. The number of observations is in parentheses.
*$P<0.001$ controls, $P<0.001$ tetrodotoxin plus veratridine (*t* test).

Table 2 Calcium-dependence of the veratridine-induced increase in dopamine synthesis

	Dopamine synthesis (nmol h^{-1} g^{-1})		
	$+Ca^{2+}$	$-Ca^{2+}$ + EGTA	$-Ca^{2+}$
Controls	13.8 ± 0.22 (18)	13.8 ± 0.41 (18)	12.8 ± 0.72 (10)
Veratridine	18.1 ± 0.30 (18)*	14.4 ± 0.44 (18)	12.0 ± 0.82 (10)

Aliquots of the striatal P_2 fraction were incubated for 5 min at 37° C either in control medium, Ca^{2+}-free medium containing 1 mM EGTA, or simply Ca^{2+}-free medium, followed by the simultaneous addition of veratridine (7.5×10^{-5} M) plus L-1-^{14}C-tyrosine (1×10^{-5} M) or by the addition of tyrosine alone, and incubated for an additional 5 min. Values represent the mean ± s.e.m. The number of observations is in parentheses.
*$P<0.001$ controls.

In the article "Dissociation of EEG and behavioural effects of ethanol provides evidence for a noncholinergic basis of intoxication" by W. R. Klemm (*Nature*, **251**, 234; 1974) the following corrections should be made. In the legend to Fig. 1, the dose of eserine (Fig. 1*b*) should be 0.2 mg kg^{-1}, and of alcohol (Fig. 1*c*) should be 2 g kg^{-1}. Likewise on page 235, right-hand column, line 27, the dose should be 2 g kg^{-1}.

In the article "Radiocarbon chronology for Seibal, Guatemala" by R. Berger, S. de Atley, R. Protsch and G. R. Willey (*Nature*, **252**, 472; 1974) the first word in line 4, para 1 should read 'stages' and we erroneously gave the address of R. Protsch as Frankfurt am Main, DDR, when in fact it should be West Germany.