

cholamines promote REM sleep⁴; our data indicate that in man catecholamines tend to inhibit and serotonin tends to enhance REM sleep.

RICHARD J. WYATT
THOMAS N. CHASE
JIMMY SCOTT¹
FREDERICK SNYDER

National Institute of Mental Health,

KARL ENGLEMAN

National Heart Institute,
9000 Rockville Pike,
Bethesda,
Maryland 20014.

Received May 20, 1970.

- ¹ Aserinsky, E., and Kleitman, N., *Science*, **116**, 273 (1953).
² Dement, W. C., and Kleitman, N., *EEG Clin. Neurophysiol.*, **8**, 673 (1957).
³ Snyder, F., Hobson, J. A., Morisson, D. F., and Goldfrank, F., *J. Appl. Physiol.*, **19**, 417 (1964).
⁴ Jouvet, M., *Science*, **163**, 32 (1969).
⁵ Jouvet, M., and Delorme, J., *CR Soc. Biol.*, **159**, 895 (1963).
⁶ Jouvet, M., Bubillier, P., Pugol, J. F., and Renault, J., *CR Soc. Biol.*, **160**, 2343 (1966).
⁷ Delorme, F., Froment, J. L., and Jouvet, M., *CR Soc. Biol.*, **160**, 2347 (1966).
⁸ Oswald, I., *Pharmacol. Rev.*, **20**, 273 (1968).
⁹ Constantinidis, J., Bartholm, G., Tissot, R., and Pletscher, A., *Experientia*, **42**, 130 (1968).
¹⁰ Rechtschaffen, A. F., in *A Manual of Standardized Terminology, Techniques and Scoring System for Sleep Stages of Human Subjects* (edit. by Land, A., and Kales, A.) (US Department of Health, Education and Welfare, PHS, NIH and NINDS, Bethesda, Maryland, 1968).
¹¹ Butcher, L. L., and Engle, J., *Brain Res.*, **15**, 233 (1969).
¹² Chase, T. N., Schnur, J., and Gordon, E. K., *Intern. J. Neuropharmacol.* (in the press).
¹³ Gebrode, F., and Bowers, M., *J. Neurochem.*, **15**, 1053 (1968).
¹⁴ Wyatt, R. J., Engleman, K., Kupfer, D. J., Scott, J., Sjoerdsma, A., and Snyder, F., *EEG Clin. Neurophysiol.*, **27**, 526 (1969).
¹⁵ Mandell, M. P., Mandell, A. J., and Jacobson, A. M., *Rec. Adv. Biol. Psychiat.*, **7**, 115 (1964).
¹⁶ Wyatt, R. J., Engleman, K., and Snyder, F., *Psychophysiology* (in the press).

Synthesis of γ -Aminobutyric Acid in Fish Erythrocytes

WE have found evidence of high activity of the enzyme glutamate decarboxylase (GAD) outside the nervous system. This enzyme catalyses the formation of γ -aminobutyric acid (GABA), which is believed to be an inhibitory neurotransmitter¹. Recently, however, a high concentration of GABA (11 mmol/kg wet weight) was found in flounder (*Pleuronectes flesus* L.) erythrocytes, where GABA may participate in the maintenance of a steady cell volume during changes in plasma osmolality^{2,3}.

The formation of GABA from glutamate was demonstrated as follows. Erythrocytes were homogenized in isotonic saline and incubated for 1 h at 10° C with 0.6 μ mole/ml. homogenate of DL-[5-¹⁴C]glutamate (specific activity 3.7 mCi/mmol, New England Nuclear Corp.). Amino-acids were extracted with alcohol and separated by thin-layer chromatography³. Spots were visualized by autoradiography and by treatment with ninhydrin reagent. The main labelled ninhydrin-positive product could not be distinguished from GABA (Fig. 1, a5 and b5). Radioactivity was also found in three other substances, one of which (Fig. 1, b6) was tentatively identified as α -ketoglutarate. No metabolism of GABA was detected by incubating with 5-¹⁴C-GABA in similar conditions.

GAD activity was measured at 25° C by a micro method⁴ depending on the production of ¹⁴CO₂ from L-[1-¹⁴C]-glutamate (California Biochemical Corp.). The incubation mixture contained (final concentration): 3.5 mM sodium L-glutamate (75,000 c.p.m.), 20 mM sodium phosphate buffer (pH 6.5), 1 mM dithiothreitol, 0.1 mM pyridoxal phosphate and 0.25 per cent of 'Triton X-100'. In those conditions the activity of rat brain was 4.70 \pm 0.25

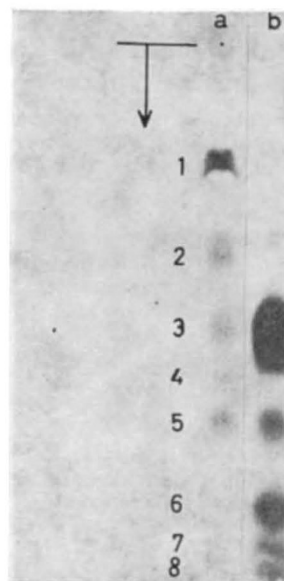


Fig. 1. a, Chromatogram of alcohol extract from flounder erythrocytes, showing the ninhydrin-positive spots (1-5); 3=glutamate; 5=GABA. b, Autoradiogram of the chromatogram a, showing the radioactive spots (3 and 5-8); 3=glutamate; 5=GABA; 6= α -ketoglutarate.

μ moles/h/g wet weight (mean \pm s.e., three experiments). In flounder, the following GAD activities (μ moles/h/g wet weight) were found: erythrocytes, 0.93 \pm 0.08 (mean \pm s.e., six animals); brain, 2.65 \pm 0.27 (mean \pm s.e., six animals). Other tissues contained low activities (liver, 0.06; kidney, 0.03—serum not detectable, one animal) some of which is accounted for by the presence of retained blood cells.

Like GAD of rat brain, the enzymes from flounder erythrocytes and brain were extractable with water, and were inhibited by high concentrations of Cl⁻ and by inactivation of pyridoxal phosphate. Thus, the erythrocyte enzyme was inhibited 77 per cent by 300 mM KCl and 69 per cent by 2 mM NaCN. The figures for the brain enzyme were 88 per cent and 72 per cent. *K_m* values measured at 25° C were for flounder erythrocytes 2.3 mM, for flounder brain 24 mM and for rat brain 12 mM (means of two experiments). It is interesting that flounder erythrocytes contain a low concentration of glutamate (less than 0.2 mmole/kg wet weight)³.

GAD had been thought to occur solely in nervous tissue¹, but has recently been observed in mammalian kidney^{5,6}. The kidney enzyme, however, differs from GAD of brain and of flounder erythrocytes for it is activated by Cl⁻ and probably does not require pyridoxal phosphate⁵.

Part of this work was supported by grants from the Nansen Foundation, Norway, and the Norwegian Research Council for Science and the Humanities.

K. FUGELLI

Institute of Zoophysiology,
University of Oslo,
Oslo 3.

J. STORM-MATHISEN
F. FONNUM

Norwegian Defence Research Establishment,
Division for Toxicology,
N-2007 Kjeller.

Received January 2; revised February 9, 1970.

¹ Roberts, E., and Kuriyama, K., *Brain Res.*, **8**, 1 (1968).

² Fugelli, K., *Comp. Biochem. Physiol.*, **22**, 253 (1967).

³ Fugelli, K., *Experientia*, **26**, 361 (1970).

⁴ Albers, R. W., and Brady, R. O., *J. Biol. Chem.*, **234**, 926 (1959).

⁵ Haber, B., Kuriyama, K., and Roberts, E., *Fed. Proc.*, **577** (1969).

⁶ Whelan, D. T., Scriver, C. R., and Mohyuddin, F., *Nature*, **224**, 916 (1969).