still. The chemical nature of the acceptors of GalNAc groups in the red cell membrane is under investigation. H

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- ¹ Watkins, W. M., Science, 152, 172 (1966).
- Watkins, W. M., and Morgan, W. T. J., Vox Sang., 4, 97 (1959).
- ⁴ Tuppy, H., and Schenkel-Brunner, W. L., Nature, 210, 316 (1966).
 ⁵ Tuppy, H., and Schenkel-Brunner, H., Europ. J. Biochem., 10, 152 (1969). V. M., Smith, Z. G., and Watkins, W. M., Biochem. J., 109, 315 ⁶ Hearn.
- (1968).
- ⁷ Kobata, A., Grollman, E. F., and Ginsburg, V., Arch. Biochem. Biophys., 124, 609 (1968).
- ⁸ Tuppy, H., and Schenkel-Brunner, H., Vox Sang. (in the press).
 ⁹ Race, C., Ziderman, D., and Watkins, W. M., Biochem. J., 107, 738 (1968).
- ¹⁰ Kobata, A., Grollman, E. F., and Ginsburg, V., Biochem. Biophys. Res. Commun., 32, 272 (1968).
- ¹¹ Bray, G. A., Anal. Biochem., 1, 279 (1960).
- ¹² Economidou, J., Hughes-Jones, N. C., and Gardner, B., Vox Sang., 12, 321 (1967).

High Cell Wall Galactosamine Content and Virus Particles in Penicillium stoloniferum

RECENTLY it has been shown^{1,2} that the active component of Statolon, an antiviral agent isolated from 5 day old culture filtrates of Penicillium stoloniferum (strain American Type Culture Collection-ATCC 14586), is double stranded RNA of viral origin, and not a complex polysaccharide as originally suggested³. During this investigation it was discovered that Statolon contained three discrete polysaccharides, readily separable by high voltage paper electrophoresis: a heteroglycan, containing galacturonic acid, galactose, xylose, arabinose and rhamnose; a glucan of the amylopectin-glycogen type (unpublished results of K. W. B., E. B. C. and J. M. Tyler) and a polymer of galactosamine, derived from lysis of the fungal cell wall. We now report that galactosamine is a major constituent of the cell wall of the virus-containing strain of P. stoloniferum (ATCC 14586); in contrast, cell walls of four naturally occurring virus-free* strains of P. stoloniferum (ATCC 10111, Commonwealth Mycological Institute 31200, 91960 and 92219) and a virus-free* isolate (ATCC 14586 B3/1) derived from the infected strain by heat treatment of spores¹ contained only very small amounts of galactosamine. (It is possible that this heat treatment procedure resulted in selection of either a heat resistant or a mutant strain of ATCC 14586.)

Amino-sugars were liberated from cell walls, prepared from 2 day old mycelium, by hydrolysis with 5.7 N HCl at 100° C for 4 h. Two amino-sugars (A and B), which gave positive reactions with the ninhydrin⁴, silver nitrate⁵ and Elson-Morgan⁶ sprays, were detected by paper chromatography (ethyl acetate : pyridine : water : acetic acid 5:5:1:3, by vol.)⁷. On oxidative deamination with ninhydrin⁸ they gave lyxose and arabinose, respectively, and were identified as galactosamine and glucosamine by paper chromatography and gas chromatography⁹ (Table 1). Talosamine and mannosamine, which also give lyxose and arabinose, respectively, on ninhydrin degradation, are included for comparison.

Maximal yields of both hexosamines were obtained after 4 h hydrolysis. The amounts of anhydrohexosamine in the cell walls of the strains examined were determined by gas chromatography as described here (Table 2).

The cell walls of the virus-containing strain have between eighteen and forty-five times more galactosamine

* No virus particles could be detected in extracts from these strains by electron microscopy, sucrosc density gradient centrifugation and serology, but the possibility that particles were present below the level of detection or that the virus was present in the "prophage" state cannot be excluded.

Table 1. IDENTIFICATION OF AMINO-SUGARS IN P. stoloniferum CELL WALLS

	Ninhydrin degradation product	Paper chromato- graphic mobility*	Gas chro- matographic retention time †
Amino-sugar A	Lyxose	1.00	3.36
Galactosamine	Lyxose	1.00	3.34
Talosamine	Lyxose	1.30	2.65
Amino-sugar B	Arabinose	1.18	2.87
Glucosamine	Arabinose	1.18	2.85
Mannosamine	Arabinose	1.53	3.53
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* Expressed as $R_{galactosamine}$ values in ethyl acetate : pyridine : water (5:2:5, by vol.).

* Retention time of hexosaminitol hexa-acetate relative to sorbitol hexa-acetate on a 5 foot column of 10 per cent neopentylglycol sebacate polyester on acid-washed 'Chromosorb W' at 240° C, essentially as described by Perry and Webb⁹.

Table 2.	AMINO-SUGAR	COMPOSITIC	ON OF P.	stoloniferun	<i>i</i> cell	WALLS
Culture	AT 145	CC ATCC 86 14586 B3/1	ATCC 10111	CMI 31200	CMI 91960	CMI 92219
Per cent anhy galactosam	ydro- ine 18 [.]	1 0.7	0.6	0.7	1.0	0.4
glucosamine	e 16	6 13·7	23·0	22.9	22.6	16.4

than the virus-free strains; in contrast, the amounts of glucosamine do not vary widely. Galactosamine has been detected in the cell walls of a number of Ascomycetes¹⁰ and is a major component of the cell walls of the Trichomycete, Amoebidium parasiticum¹¹. Only small amounts of galactosamine were found, however, in the cell walls of P. notatum $(0.9-1.8 \text{ per cent})^{12}$ and P. patulum $(0.5 \text{ per cent})^{13}$, and none was reported in the cell walls of P. chrysogenum¹⁴, P. italicum¹⁵ and P. digitatum¹⁵. Our analyses of P. stoloniferum suggest that the large amount of galactosamine in the cell walls of strain ATCC 14586 may be associated with the presence of virus particles. Although such an effect has not been described previously in fungi, alterations of cell wall structure as a result of phage infection in bacteria have been widely reported¹⁶. The possibility that galactosamine forms part of a receptor site in the cell wall to which the virus binds in the initial stages of infection is being investigated. It has been suggested that galactosamine forms part of the phage receptor sites of group C Streptococcus¹⁷.

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- ¹ Banks, G. T., Buck, K. W., Chain, E. B., Himmelweit, F., Marks, J. E., Tyler, J. M., Hollings, M., Last, F. T., and Stone, M. M., *Nature*, **218**, 542 (1968).
- ⁸ Kleinschmidt, W. J., Ellis, L. F., Van Frank, R. M., and Murphy, E. B., ⁶ Kleinschnidt, W. J., and Probst, G. W., Antibiot. Chemother., 12, 298 (1962).
 ⁸ Kleinschmidt, W. J., and Probst, G. W., Antibiot. Chemother., 12, 298 (1962).
 ⁴ Aminoff, D., and Morgan, W. T. J., Nature, 162, 579 (1948).
- ⁶ Trevelyan, W. E., Procter, D. P., and Harrison, J. S., Nature, 155, 444
- (1950).⁶ Smith, I., Chromatographic and Electrophoretic Techniques, 1, 252 (Heinemann, London, 1960).
 ⁷ Fischer, F. G., and Nebel, H. J., Z. Physiol. Chem., 302, 10 (1955).
- ⁸ Stoffyn, P. J., and Jeanloz, R. W., Arch. Biochem. Biophys., 52, 373 (1954).
- ⁹ Perry, M. B., and Webb, A. C., Canad. J. Biochem., 46, 1163 (1968).
- 10 Bartnicki-Garcia, S., Ann. Rev. Microbiol., 22, 97 (1968).
- ¹¹ Trotter, M. J., and Whistler, H. C., Canad. J. Bot., 43, 869 (1965).
- ¹² Applegarth, D. A., Arch. Biophys. Biochem., 120, 471 (1967).
- 13 Applegarth, D. A., and Bozoian, G., J. Bact., 94, 1787 (1967).
- 14 Hamilton, R. B., and Knight, S. G., Arch. Biochem. Biophys., 99, 282 (1962).
- ¹⁶ Grisaro, V., Sharon, N., and Barkai-Golan, R., J. Gen. Microbiol., 51, 145 (1968).
- ¹⁰ Lüderitz, O., Jann, K., and Wheat, R., in Comprehensive Biochemistry (edit. by Florkin, M., and Stotz, E. H.), 26A, 106 (Elsevier, Amsterdam, (edit. 1968).
- 17 McCarty, M., and Morse, S. I., Adv. Immunology, 6, 249 (1964).