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J. R.	BACKHURST
F. Go	ODRIDGE
R. E.	PLIMLEY

M. FLEISCHMANN

Department of Chemical Engineering, University of Newcastle upon Tyne.

Department of Chemistry, University of Southampton.

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Reconstitution Mechanisms in the Radiolysis of Aqueous Biochemical Systems: Inhibitive Effects of Thiols

The hydrated electron, e_{aq}^{-} , and the OH radical are the principal products of the radiation induced decomposition of liquid water; with gamma rays the 100 eV yields¹ of these reactive intermediates are closely approximated by the values $G_{\cdot,q}^{e} \simeq G_{OH} \simeq 2.6$. While certain classes of organic compounds are known to react quantitatively with both eaq and OH in oxygen-free solution, the observed yield for net destruction of such solutes may be quite low compared with the G value for water decomposition. In interpreting the radiation chemistry of such systems the concept of a reconstitution or back-reaction must be invoked.

For example, in the gamma radiolysis of oxygen-free solutions of the pyrimidine bases, both eaq and OH add preferentially to the 5,6 double bond^{2,3}

$$e_{a0}^{-} + B + H_2O \rightarrow BH + OH^{-}$$
 (1)

$$OH + B \rightarrow BOH$$
 (2)

where the rate constants k_2 , k_3 fall in the range 10⁹ to 10^{10} M⁻¹ s⁻¹ (refs. 4 and 5). Yet the observed yield for base destruction in millimolar solutions is uniformly low with G(-B) < 0.9. The evidence is that a back-reaction³

$$\dot{B}H + \dot{B}OH \rightarrow 2B + H_{*}O$$
 (3)

leads to a reconstitution of the base.

We have found that certain labile compounds such as ascorbic acid and the thiol, cysteine, are effective at low concentrations in blocking reaction (3) by virtue of the H atom transfer reaction

$$BH + R'H \rightarrow BH_2 + R'$$
(4)

where BH₂ represents the dihydro derivative of the pyrimidine nucleus. The effects of added cysteine on the yields for radiolytic reduction of cytosine in oxygen-free solution are shown in Table 1.4. Note that dihydrocytosine is unstable and spontaneously hydrolyses to yield ammonia and dihydrouracil⁶ as the experimentally observed product. The product dihydrouracil was isolated and characterized both chromatographically and spectrophotometrically. The data of Table 1A show that the yield for the radiolytic reduction of the 5,6 double bond of cytosine is essentially quantitative in the presence of either cysteine or ascorbic acid at low concentrations.

Table 1. EFFECT OF CYSTEINE AND ASCORBIC ACID (R'H) on the GAMMA RAY INDUCED REDUCTION OF PYRIMIDINE (C=C) AND PEPTIDE (C=O) LINKAGES IN OXYGEN-FREE SOLUTIONS^{*}

(A) Cytosine \rightarrow dihydrouraeil (BH₂) + ammonia †

Solution	<i>R</i> 'H ‡	$G(\mathrm{BH}_2)$
0.05 M cytosine, <i>p</i> H 7	None	<0.9
0.05 M cytosine, <i>p</i> H 7	Cysteine, 2·5 × 10 ⁻³ M	2.8
0.05 M cytosine, <i>p</i> H 7	Ascorbic acid, 1·5 × 10 ⁻³ M	2.9

(B) N-ethylacetamide \rightarrow acetaldehyde (RCHO) + ethylamine G(RCHO) Solution R'H

1 M N-ethylacetamide, pH 7 None 1 M N-ethylacetamide, pH 7 Cysteine, 4×10^{-4} M <0·1 2·8

* At dosages below $\sim 2.5 \times 10^{18} \text{ eV/g}$.

Similarly, in the radiolysis of primary amides and monosubstituted primary amides (peptides) in oxygen-free neutral solution, the reducing and oxidizing species e_{aq} and OH are removed through reactions of the type

$$e_{aq}^{-} + RCONHR + H_2O \rightarrow RC(OH)NHR + OH^{-}$$
 (5)

$$OH + RCONHR \rightarrow P + H_2O \tag{6}$$

where k_6 , k_7 fall in the range 10⁷ to 10⁸ M⁻¹ s⁻¹ (refs. 7 and 8). Combination of RC(OH)NHR with like species or with the radical P would lead to formation of ketonic products. The combined yield of such products is low, however, with $G(>CO) \simeq 0.2$; the evidence is that the reconstitution reaction

$$RC(OH)NHR + P \rightarrow 2RCONHR$$
(7)

represents the major stoichiometry for removal of organic radical in such systems⁸.

In accord with this, we find that cysteine effectively blocks the reconstitution reaction (7) through the step

$$R(COH)NHR + R'H \rightarrow RCHO + NH_2R + R'$$

and, as shown in Table 1B, the reductive deamination of the peptide bond in the presence of cysteine at low concentrations is essentially quantitative with $G(CH_3CHO)$ $\simeq 2.5.$

We find then that (a) the presence of cysteine or ascorbic acid at low concentrations leads to a very marked enhancement in the radiolytic lability of the pyrimidine and peptide moieties in oxygen-free solution, and that (b) such enhancement arises as a consequence of the blocking by the second solute of the reconstitution reactions formulated in equations 3 and 7.

These results seem to have interesting applications in the study and identification of reductive processes involving reactions of e_{aq} with unsaturated organic functions both in vitro and in vivo.

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> JOHN HOLIAN WARREN M. GARRISON

Lawrence Radiation Laboratory, University of California, Berkeley, California.

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