

Hormonal Control of Lipid Concentration in Rat Heart and Gastrocnemius

In perfused rat heart and isolated rat diaphragm the rate of release and oxidation of fatty acids from muscle glycerides is increased by alloxan-diabetes¹. These changes in diabetes involve actions of growth hormone and corticosteroids in insulin-deficient rats because they are not seen in muscles of hypophysectomized diabetic rats unless the animals are treated with growth hormone and cortisol¹. As part of an investigation of the mechanisms of these changes we have investigated the effects of diabetes, hypophysectomy and treatment with growth hormone and cortisol on the concentrations of glycerides and phospholipids in rat heart and gastrocnemius (the latter as a representative skeletal muscle). The methods and procedure were as follows.

Rat hearts were perfused for 10 min with medium containing glucose (5.5 mM) and insulin (0.1 unit/ml.), and the excised ventricles frozen with a tissue clamp and powdered in a percussion mortar². Gastrocnemius muscle was frozen in acetone-solid carbon dioxide mixture and powdered after 10 min of hind-limb perfusion through the abdominal aorta with the foregoing medium containing glucose (11 mM) but no insulin. These perfusions remove blood and blood lipids. Details of induction of alloxan-diabetes and treatment with growth hormone and cortisol were as described elsewhere². The frozen muscle powder was extracted with chloroform/methanol (2:1 v/v) (15–30 ml./g) in a Potter homogenizer, allowed to stand at 0° for 2 h and centrifuged. The supernatant was shaken with 0.3 vol. of 4 mM magnesium chloride and phases separated by centrifugation after 30 min. The chloroform phase was evaporated to dryness at 70° and the residue taken up in about 7 ml. of chloroform and a sample (0.5 ml.) taken for determination of phospholipid-P (ref. 3), after evaporation of chloroform and oxidation with 0.15 ml. concentrated H₂SO₄/HClO₄ (2:1 v/v). The remainder was shaken with 0.5 g silicic acid to remove phospholipid⁴ (confirmed by assay of phospholipid-P). A sample of supernatant (4 ml.) was evaporated to dryness and glycerides separated by thin-layer chromatography on 'Silica Gel G' (Merck) using as solvent petroleum ether (fraction 40°–60°)/di-ethyl ether/methanol/acetic acid (90:20:2:3 v/v). As markers crude triolein (British Drug Houses, Ltd.) containing mono- and di-olein, and pure tripalmitin and mono-olein (Hormel Institute, University of Minnesota), were used (about 0.5 μmole). The R_F values of the separated classes were: mono-glycerides 0.05; diglycerides 0.45; triglycerides 0.90. The spots, located by markers, were scraped from the plate and extracted twice with 2 ml. of chloroform/methanol (2:1 v/v). The extract was evaporated to dryness at 70°, saponified with 0.5 ml. 4 per cent potassium hydroxide in 95 per cent ethanol for 30 min at 60°, acidified with 0.5 ml. 10 per cent HClO₄, neutralized with saturated KHCO₃, and KClO₄ separated by centrifugation at 0°. Glycerol was assayed enzymatically in the supernatant⁵. Recoveries (using 1-[¹⁴C]-palmitate labelled tripalmitin from the Radiochemical Centre, Amersham, added to extracts of heart) were in excess of 90 per cent. Recovery of mono-olein was in excess of 80 per cent. Contamination with phospholipid-P was less than 0.05 per cent.

The results are shown in Table 1. In the glyceride series the concentration of triglyceride was greatly in excess of that of di- and mono-glyceride, the concentration ratios being 140:1:6 in normal rat heart and 120:1:4 in normal rat gastrocnemius. These values indicate that loss of the first fatty acid may limit triglyceride breakdown in muscle. The concentration of triglyceride in heart and gastrocnemius was significantly increased by alloxan-diabetes. In hypophysectomized rats alloxan-diabetes failed to increase the triglyceride concentration unless the animals were treated with growth hormone

Table 1. GLYCERIDE AND PHOSPHOLIPID CONCENTRATIONS IN RAT HEART AND GASTROCNEMIUS

Rat	Tissue	Concentration μmoles/g dry wt. (Mean ± S.E.)			Phospholipid phosphorus
		Triglyceride	Diglyceride	Monoglyceride	
Normal (6)	Heart	13.6 ± 1.4	0.1	0.6	184 ± 8
Alloxan-diabetic (6)		* 28.2 ± 1.9	0.6	0.7	163 ± 8
Hypophysectomized (6)		10.3 ± 1.3	0.4	0.7	143 ± 4
Hypophysectomized diabetic (6)		10.6 ± 0.9	0.3	0.7	154 ± 4
Hypophysectomized diabetic treated with growth hormone and cortisol (4)	Gastrocnemius	* 22.9 ± 3.7	—	—	142 ± 5
Normal (8)		12 ± 1.5	0.1	0.4	64 ± 2
Alloxan-diabetic (8)		26 ± 2.7	0.4	0.5	63 ± 1

* P < 0.01 versus control. No. of muscle samples given in parentheses.

and cortisol. The concentration of diglyceride (measured on pooled extracts) may also have been increased in alloxan-diabetes whereas the concentrations of mono-glyceride and phospholipids appeared to be unchanged. When hearts from normal rats were perfused for 60 min without substrate the triglyceride concentration fell from 18.7 ± 0.8 (8) to 8.7 ± 0.7 (8) (μmoles/g dry wt.; mean ± S.E. with number of hearts). No significant change in phospholipid was detected (186 ± 6–171 ± 7).

These observations lead us to conclude that heart muscle triglyceride is broken down during perfusion, thus providing fatty acid for oxidation⁶. Evidence for the location of this glyceride in muscle cells has been presented^{1,7}. The concentration of triglyceride, but not that of phospholipid, is increased by actions of growth hormone and cortisol in an insulin-deficient rat. Loss of the first fatty acid from triglyceride appears to limit breakdown in both normal and diabetic rats. The role of increased triglyceride concentration in accelerated lipolysis in diabetic muscle¹ and the source of muscle glyceride (that is, whether derived from plasma glyceride or plasma non-esterified fatty acids) is under investigation.

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R. M. DENTON
P. J. RANDLE

Department of Biochemistry,
University of Bristol.

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Acrylamide-gel Electrophoresis of β-Lactoglobulins stored in Solutions at pH 8.7

NATIVE β-lactoglobulins-A and -B, mol. wt. 35,500, can reversibly dissociate into sub-units of mol. wt. 17,750 in acidic¹ or alkaline² solutions. Aggregation of β-lactoglobulin-A to a tetramer structure has been observed¹ in solutions of pH 3.5–5.2 stored at 4° C. Irreversible denaturation of β-lactoglobulin occurs on heating³ neutral solutions above 65° C or on storage^{4,5} in alkaline solution at 25° (slowly in the pH range 8–9.5, more rapidly in solutions of higher pH). This denaturation was shown by changes in iso-electric solubility, optical rotation and light scattering; Tiselius electrophoresis in 'Veronal' (pH 8.4) buffer separated one, or occasionally two, faster-migrating components in addition to native β-lactoglobulin. An initial reversible splitting followed by irreversible aggregation was postulated³, but the nature and number of products formed were not studied further. Bingham, Krugman and Estermann⁶ recently noted that stored samples of β-lactoglobulin were separated by acrylamide disc electrophoresis⁷ into five bands whereas the fresh material split into only two bands; they gave no indication