

Shrinkage Temperature of Eye Collagen

DURING a study of the shrinkage temperature of skin collagen, normal human eye collagen was included in the control material. A measure of difference was found between the sclera, which behaved as ordinary mammalian collagen¹⁻³, and the cornea, which showed a lower hydrothermal stability, in the sense that it shortened at a lower temperature than sclera or skin collagen.

Although the structure of the transparent cornea is well documented^{6,7}, and descriptions exist of the analytical differences between it and the sclera⁸, there seems to be no ready reference to a lower shrinkage temperature.

Using a modified Theis apparatus⁹ the results were as shown in Table 1.

Table 1
Shrinkage temperature (° C)

| Specimen | Human eye | | Shrinkage temperature (° C) | | | | | | | | | | | | |
|----------|---------------|-----|-----------------------------|----|----|----|----|----|----|----|----|----|----|---|--|
| | Sex | Age | 55 | 56 | 57 | 58 | 59 | 60 | 61 | 62 | 63 | 64 | 65 | | |
| 314 | Normal cornea | F | 66 | | | | 1 | | | | | | | | |
| 314 | Normal sclera | F | 66 | | | | | | | | 1 | | | | |
| 318 | Normal cornea | M | 84 | | 1 | | | | | | | | | | |
| 318 | Normal sclera | M | 84 | | | | | | | | 1 | | | | |
| 321 | Normal cornea | F | 81 | | 1 | | | | | | | | | | |
| 321 | Normal sclera | F | 81 | | | | | | | | 1 | | | | |
| 323 | Normal cornea | M | 62 | | 1 | | | | | | | | | | |
| 323 | Normal sclera | M | 62 | | | | | | | | 1 | | | | |
| 324 | Normal cornea | M | 1 | | | | | | | | | | | | |
| 324 | Normal sclera | M | 1 | | | | | | | | 1 | | | | |
| 325 | Normal cornea | F | 60 | | 1 | | | | | | | | | | |
| 325 | Normal sclera | F | 60 | | | | | | | | | 1 | | | |
| 330 | Normal cornea | M | 18 | | | | | | | | | | 1 | | |
| 330 | Normal sclera | M | 18 | | | | | | | | | | | 1 | |
| 333 | Normal cornea | M | 1 | | | | | | | | | | | | |
| 333 | Normal sclera | M | 1 | | | | | | | | | | | | |
| 291 | Normal cornea | M | | | | | 1 | | | | | | | | |
| 272 | Normal cornea | F | 86 | | 1 | | | | | | | | | | |
| STB1 | Normal cornea | M | 69 | | 1 | | | | | | | | | | |
| STB2 | Normal cornea | M | 58 | | | | 1 | | | | | | | | |
| STB3 | Normal cornea | F | 82 | | 1 | | | | | | | | | | |
| STB3 | Normal sclera | F | 82 | | | | | | | 1 | | | | | |
| STB4 | Normal cornea | M | 65 | | | | | | | | | | 1 | | |
| STB4 | Normal sclera | M | 65 | | | | | | | | | | | 1 | |

These findings relate only to the translucent area of normal eyes; investigation of pathological specimens is pending.

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H. STRINGER
J. PARR

The Wellcome Medical Research Institute,
and the Department of Ophthalmology,
University of Otago Medical School,
Dunedin, New Zealand.

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Biochemical Chlorination of *Streptomyces aureofaciens*

EARLIER work on the chlorination mechanism of *Streptomyces aureofaciens* established that chlorine of chlorinated fatty acids was incorporated into the molecule of 7-chlorotetracycline (CTC)^{1,2}.

Recently, Sekizawa has found that the chlorine of chloropropanediols was also incorporated into CTC and, moreover, this compound was able to overcome the effect of known chlorination inhibitors³⁻⁵. Sekizawa also reported that after the addition of chloropropanediols to the media of oxytetracycline-producing strains, CTC could be detected⁶. He therefore supposed chloropropanediols to enter directly into the tetracycline (TC) molecule during the course of biosynthesis preceding the chlorination reaction.

The conditions of chlorine incorporation were investigated by the application of Na³⁶Cl, using 2,5-dimercaptothiadiazole^{1,3,4} as chlorination inhibitor. Regarding organic chloro-compounds, the incorporation of chlorine from 1-¹⁴C-acetate (MCA), 2-¹⁴C-MCA and 3-chloropropanediol^{1,2} was investigated. Incorporation of the carbon of monochloroacetates into TC was also detected.

Strains of CTC-producing *Str. aureofaciens* 'B-28' (ref. 7) and the tetracycline producer *Str. aureofaciens* 'CDS-314' (ref. 8) were used for the present experiments. The synthetic medium and experimental conditions were the same as reported earlier^{7,9}.

Using isotopically labelled NaCl it was shown that TDA exerts its inhibitory action in the first oxidative stage of chlorination. Na³⁶Cl could be detected in the broth of chlortetracycline fermentations inhibited by TDA. Similarly, chloride could be detected about the end of fermentations of TC-producing 'CDS-314' strain. These facts support the assumption that inhibition of chlorination was effected in the first phase of the biosynthetic chain, genetically manifest during the course of strain production⁸. But, in contrast to the inhibition of TDA, in the case of strain 'CDS-314' copper did not suspend the inhibition of CTC production; the genetic block has been found to be irreversible (Table 1).

Table 1 shows that, in the case of *Streptomyces aureofaciens* 'CDS-314', the 'residual reaction' supposed by Sekizawa⁶ to be general in chlorination reactions could not be found. In our opinion the appearance of a 'residual reaction' in the case of predominantly TC-producing strains depends on the localization of the genetic block. We suppose, however, that at the very first stage of chlorination there is no alternative pathway in the biogenesis of CTC; later, there may be a partial separation of the reaction routes.

By the use of 1-¹⁴C-MCA and 2-¹⁴C-MCA it could be shown that, in the case of strain 'B-28', the chlorine of MCA, decomposed enzymatically, was incorporated into the CTC molecule. It was found that TDA did not inhibit the incorporation of carbon from MCA 1-C and 2-C into the tetracycline molecule. Similarly, in the case of strain 'CDS-314', independently from the genetic block, carbon atoms of MCA were incorporated into TC (Table 2). We suppose that the first step of chlorine utilization is an enzymatic cleavage of the chlorine of MCA to inorganic chloride during the metabolism of both strains. Afterwards, evidently, chloride will be incorporated into CTC only in the case of the CTC-producing strain 'B-28'.

The possibility reported by Sekizawa that propanediol may be a direct precursor for CTC would have been confirmed if the production of CTC could have been detected at the end of the fermentation, after giving chloropropanediol to the culture of the exclusively TC-producing strain 'CDS-314'.

Experimental results obtained in our laboratory did not support the assumption of Sekizawa. CTC could not be detected in the fermentation broth of strain 'CDS-314', thus excluding the possibility that chloropropanediol acts as a direct precursor of CTC, if it is assumed that, in general, the biosynthesis is similar in TC- and CTC-producing

Table 1

| Strain | Materials* | Total antib. activity mcg/ml. | Breaking per mg of antibiotics | CTC (%) | Chloride det. at the end of fermentation |
|---------|---------------------------|-------------------------------|--------------------------------|---------|--|
| B-28 | — | 1,080 | 1,512 | 100 | 32 |
| B-28 | 0.2 mM TDA | 1,040 | 396 | 26 | 80 |
| B-28 | 0.2 mM TDA | — | — | — | — |
| CDS-314 | +0.3 mM CuSO ₄ | 825 | 1,180 | 74 | 38 |
| CDS-314 | — | 1,370 | 38 | 0 | 98 |
| CDS-314 | 0.2 mM TDA | 1,280 | 32 | 0 | 96 |
| CDS-314 | 0.2 mM TDA | — | — | — | — |
| — | +0.2 mM CuSO ₄ | 1,010 | 36 | 0 | 99 |
| — | — | — | 40† | — | 96 |

* As an isotope source 0.4 μc./90 ml., Na³⁶Cl was added at 0 h for each experiment.

† Activity was related to an equal quantity of butanolic extract. The small activity observed in the control flask may be due to a small amount of ³⁶Cl dissolved in butanol.