of the myometrium to oxytocin rather than to an increase in the amounts of oxytocin released into the blood. Since the hypothalamus is functionally connected with both neurohypophysis and adenohypophysis it is tempting to speculate that the relay we have demonstrated between uterus and hypothalamus may be concerned not only with the release of oxytocin but also with effecting increased myometrial sensitivity to it, factors which together underly the mechanism of normal parturition.

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PHARMACOLOGY

Isolation and Identification of the 20_β-Hydroxy Derivatives of 6β -Hydroxycortisol and 6β -Hydroxycortisone in Liquor Amnii

The presence of C-6 oxygenated steroids in neonatal urine was first reported by Ulstrom $et \ al.^1$. It has been demonstrated^{2,3} that placental tissue incubated under standard conditions, is able to oxygenate at the C-6 position, while the work of Frantz et al.4 has indicated that during pregnancy the urinary concentrations of the polar steroid 6β -hydroxycortisol is elevated above the normal non-pregnant state. It appeared likely, therefore, that similar polar steroids might be present in liquor amnii.

In a previous communication⁵, the isolation and identi- $6\beta, 11\beta, 17\alpha, 21$ - tetrahydroxy - pregn - 4 - enefication of 3-20-dione (6\beta-hydroxycortisol) in liquor amnii were reported. At this time the presence of three additional polar steroids was recognized. Two of these compounds have now been identified as $6\beta, 17\alpha, 20\beta, 21$ -tetrahydroxy-pregn-4-ene-3,11-dione (compound 3) and 63,- $11\beta, 17\alpha, 20\beta, 21$ -pentahydroxy-pregn - 4 - ene - 3 - one (compound 4).

Liquor amnii, obtained at the time of labour, was extracted with ethyl acetate, using the methods previously described⁵. The purified extract was chromatographed in a modified BuC system⁶ and the chromatogram treated with a 2: 1 solution of 10 per cent aqueous sodium hydroxide in 50 per cent methanol and 0.02 per cent blue tetrazolium. It was dried in an oven at 60° C for 10 min. When the chromatogram was viewed under ultra-violet light, using a minus blue 4 gelatine filter (Ilford), three brilliant fluorescent zones were observed. The least polar of these has been previously identified as 6β-hydroxycortisol

Table 1. PROPERTIES OF THE 20β -Hydroxy DERIVATIVES OF 6β -Hydroxy-CORTISOL AND 6β -HydroxyCORTISONE

CORTISON AND OP HIDROALCORLISOND							
		Compound 206-66-OHF		Compound 206-66-OHE			
		Isolated	Ref.	Isolated	Ref.		
1	Ultra-violet absorption in ethanol	$238 m\mu$	$238 \ \mathrm{m}\mu$	$238 m\mu$	238 mµ		
2	Reaction with blue tetrazolium	Neg.	Neg.	Neg.	Neg.		
3	Mobility in chromato- graphic system Mod. BuC (ref. 6)	0.01	0.01	0.03	0.03		
4	Mobility after the oxidation of the product from Reaction 3 with sodium bismuthate B/50 syst. (ref. 9) LB21/80 (ref. 9)	0·24 0·01	0·24 0·01	0·32 0·03	0-32 0-03		
5	Mobility after acetylation of the product from Reaction 4 <i>LT</i> /2185 (ref. 9)	0.15	0.15	0-31	0.31		
6	Mobility after further oxidation of the product from Reaction 5 using chromic acid LT/2185 (ref. 9)	0.31	0.31	0.31	0.31		
7	Mobility after reduction of the product from Reac- tion 4 using Zn/HAc (ref. 9 LT/2185 (ref. 9)) 0·24	0-24	0.38	0.38		

(compound 1) (ref. 5). Quantities of the other two zones (compounds 3 and 4) were collected from subsequent chromatograms, and were carried through a series of chemical and enzymatic steps. Identification of these 203-hydroxy derivatives of 63-hydroxycortisol and 6β-hydroxycortisone was based on the properties as listed in Table 1. The products from each procedure were compared chromatographically with reference steroids. In each case an agreement of R_F values was found.

The configuration of the side-chain was elucidated using the enzyme 20β-hydroxysteroid dehydrogenase7, and by an examination of the chromatographic properties of the steroids. The reference steroids for this comparison were obtained by treating 6β -hydroxycortisol and 63-hydroxycortisone with sodium borohydride at 0° C (ref. 8).

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Formation of Tetrahydropapaveroline from Dopamine in vitro

IN 1938, Holtz et al.¹ reported that dopamine, which is pressor in the cat, was converted to a depressor substance after incubation with monoamine oxidase (MAO), for example, extracts of guinea pig kidneys. These authors suggested that the depressor agent might be dihydroxyphenyl acetic aldehyde, the primary deamination product of dopamine, since the addition of semicarbazide prevented the formation of the depressor substance.